

The role of redox regulation in the normal physiology and inflammatory diseases of skin

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1. ABSTRACT

Skin is the largest organ which contains complex and tightly regulated redox network of the reactive oxygen/nitrogen/lipid species producing components as well as the redox damage protective systems. This redox balancing system has evolved to regulate normal physiological processes and to protect skin and the internal organs against environmental damage. Exposure to some physical, chemical, and biological agents results in the excessive formation of free radicals and non-radical redox active species within the skin. Normally, skin reacts to this overproduction by sacrificing non-enzymatic antioxidants and by adaptive induction of both protective detoxifying and damage-eliminating systems. Thus, fast restoration of redox balance necessary to maintain normal skin structure and functioning occurs. In the case of excessive exposure or defects in the adaptive reactions, redox damage to skin components occurs. Here, we focus on the role of redox status in the acute inflammatory response to wounding and chronic inflammatory skin diseases such as psoriasis, atopic and contact dermatitis. Redox-mediated chronic inflammation and immunosuppression as risk factors for tumorigenesis are also reviewed.

2. REDOX-MEDIATED PROCESSES ESSENTIAL FOR SKIN PHYSIOLOGY

Skin is the biggest and extremely complex organ of the human body having a “sandwich-like” structure. The uppermost layer bordering environment is stratum corneum, a unique, highly lipophilic two-compartment system of enucleated cells embedded in a lipid-enriched intercellular matrix, forming stacks of bilayers that are rich in fibrous proteins, mainly, keratins, ceramides, cholesterol and free fatty acids (1). Notably, the stratum corneum of normal human skin contains non-enzymatic water and lipid-soluble antioxidants such as GSH, vitamin C, uric acid, vitamin E, squalene, and coenzyme Q10, distributed in a gradient with the highest concentration on the deepest stratum corneum layers (2).

The underlying layer (epidermis) contains keratinocytes at different stages of differentiation. Moving upward from the deepest basal level of epidermis, young keratinocytes become mature and more differentiated, ending up in stratum corneum as corneocytes. The bricks of epidermal keratinocytes are cemented by lipids and free fatty acids. Lipid-soluble antioxidants, mainly alpha-

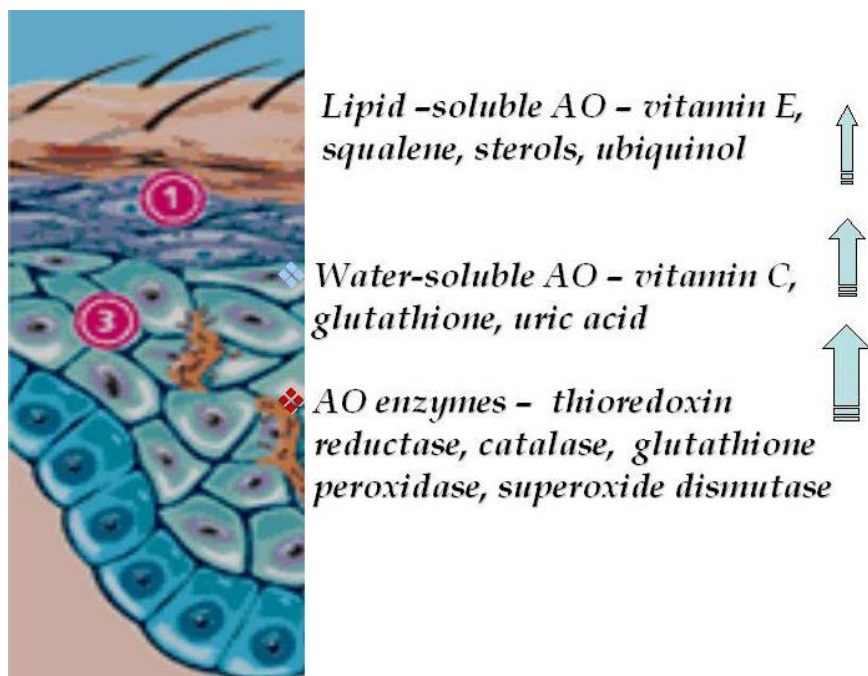


Figure 1. Distribution of low molecular and enzymatic antioxidants in the skin epidermis. The uppermost layer (*stratum corneum*) contains mainly lipid soluble low molecular antioxidants. The deeper layers contain also high levels of water-soluble non-enzymatic antioxidants as well as antioxidant enzymes. The concentrations of practically all low molecular antioxidants in the skin epidermis decrease gradually from the lower to the upper layers.

tocopherol and antioxidant enzymes like CAT, SODs, GPx, PRx were found in the epidermis (3). Beneath the epidermis, the dermal layer is rich of various types of cells such as fibroblasts, melanocytes, endothelial cells, smooth muscle cells, immune cells (lymphocytes, granulocytes, monocytes, and dendritic cells, Langerhans cells, and neurons). The extracellular space of skin derma is filled up with specific proteins (collagens and fibrins) and polysaccharides (hyaluronic acid, glucosaminoglycans, etc.). Skin derma contains large amounts of water-soluble antioxidants such as ascorbic acid, uric acid, and GSH. The distribution of major antioxidants along skin layers is shown in Figure 1. The richness in a wide variety of antioxidants in the skin could be explained, first of all, by the necessity to counteract endogenously produced redox reactive species.

Normally, practically all types of skin cells produce reactive oxygen (ROS) and nitrogen (RNS) species (Figure 2). For example, both melanocytes and keratinocytes produce H_2O_2 and superoxide radicals in the reaction of pheomelanin with UV light (4). Fibroblasts and keratinocytes produce small amount of superoxide as a by-product of electron transfer in the mitochondrial respiratory chain. All phagocytic cells have a well-characterized superoxide anion-generating plasma membrane NADPH-oxidase capable of producing the large amounts of superoxides required for their function in host defense (5). In addition, granulocytes express both cNOS and iNOS producing NO. Singlet oxygen release was detected in the activated skin granulocytes. The singlet oxygen was a secondary product in the reaction of H_2O_2 , chloride, and

myeloperoxidase. Hypochlorite, formed in the reaction, interacts with another molecule of H_2O_2 producing singlet oxygen (6). The endothelial cells of small blood vessels located in the dermal layer are equipped with eNOS, which releases NO to regulate the vasculature relaxation.

Upon the interaction of skin components with numerous physical factors (ionizing irradiation, UV-, visible, and infrared light, low and high temperatures, mechanical wounding, ultrasound and electromagnetic fields), chemical agents (ozone, tobacco smoke, organic and non-organic toxins, and heavy metal ions), and biological invaders (microbes, viruses, parasites, and allergens) additional production of reactive species occurs (2). Skin reacts by switching on adaptive processes as well as by sacrificing pre-existent antioxidants. Thus, in the normal skin, the redox balance will be gradually restored. In this sense, the redox system of the skin could be considered as a sensor of environmental signals able of translating them into second messengers (reactive redox species) to start the skin adaptive machinery.

Skin-generated ROS comprise singlet oxygen, superoxide anion, hydrogen peroxide and hydroxyl radicals (7). Although dismutation of superoxide anion probably accounts for much of the hydrogen peroxide produced by eukaryotic cells, it can also be formed by direct two-electron reduction of oxygen, a mechanism shared by a number of flavoprotein oxidases (8). Nitric oxide (NO) and peroxynitrite are major reactive nitrogen species in biological systems. NO within the skin is produced by two constitutive nitric oxide synthase (cNOS) isoforms,

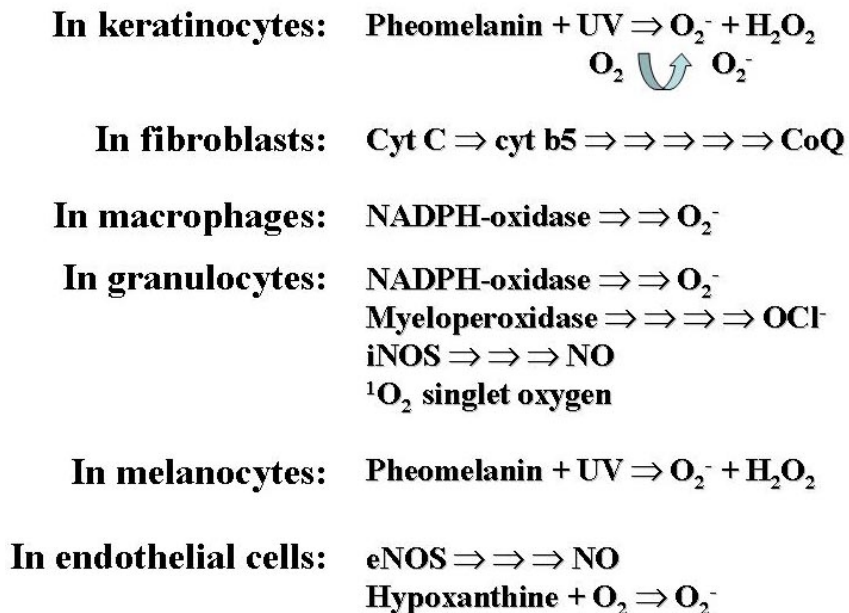


Figure 2. Reactive oxygen and nitrogen species production by different skin cells. Keratinocytes and melanocytes produce superoxide radical and hydrogen peroxide in the reaction of photo-oxidation of pheomelanin. Fibroblasts produce superoxide intracellularly as a result of leakage of the electrons from the respiratory chain in mitochondria. Macrophages have a superoxide-producing NADPH-oxidase bound to cellular membrane. Granulocytes produce a wide spectrum of primary ROS and RNS due to the activity of NADPH-oxidase, myeloperoxidase, cNOS and iNOS. Endothelial cells are a source of NO (eNOS) and superoxide produced in the reaction of xanthine with xanthine oxidase.

identified as endothelial (eNOS) and neuronal (nNOS), and one inducible (iNOS) isoform. In the skin, both fibroblasts and keratinocytes constitutively express eNOS. NO easily reacts with oxidative species, including ROS, transition metals and thiols to yield various reactive nitrogen species (RNS). In particular, NO reacts with superoxide anion at near diffusion-limited rates, leading to the extremely rapid production of the peroxynitrite ion. In the presence of limiting amount of the precursor L-arginine, uncoupled cNOS dimers become a source of both NO and superoxide anion, reaction between them giving rise to the peroxynitrite ion (9). When generated at high concentrations, peroxynitrite can diffuse and undergo transformation into other powerful oxidants, including oxidative (OH[•]) and other reactive nitrogen species (NO₂, NO₂⁺) (10). Superoxide could be also formed in the skin non-enzymatically as a by-product of neurotransmitters (adrenalin and nor-adrenalin) auto-oxidation (11). Non-enzymatic NO release from its pre-formed complexes with GSH and hemoglobin seems to be an important source of RNS in the skin under some circumstances (11). By reacting with lipid molecules directly or affecting redox-sensitive lipid-metabolizing enzymes (phospholipases, lipoxygenases, and cyclooxygenases), ROS induce production of reactive lipid species (lipid radicals, peroxides, hydroperoxides, aldehydes, etc.). The interaction of primary ROS and RNS with some molecules, like thiols, alpha-tocopherol, ascorbic acid, amino acids, *etc.* may lead to the formation of secondary radicals or new redox reactive species (11). Collectively, the following free radicals and reactive species have been identified in the

skin so far: superoxide radicals, hydroxyl radicals, singlet oxygen, hydrogen peroxide, hypochlorite, nitric oxide, peroxynitrite, alkoxyl radicals, lipid alkyl radicals, hydroxyalkyl radicals, glutathionyl radical, other thiol radicals, tocopheryl, carbon-centered radicals (12), ascorbyl radical (13), and melanin radicals (14).

It appears that the complex redox system in the skin is, probably, the most evolutionary ancient and efficient intrinsic mean for the maintenance of structural and functional integrity of skin itself, which in turn, provides proper protection of the whole organism. The redox system controls barrier and immune functions of the skin, its self-repair, and removal of cellular and molecular debris. All ROS, RNS and lipid reactive species may have both cytotoxic effects and regulatory functions depending on the enzymatic source, vicinity to biologically essential molecules/structures, relative concentration and duration of the reactive species generation. Numerous regulatory functions of redox reactive species in the skin could be exerted at both genetic (regulation of intracellular and intercellular signaling pathways) and epigenetic (metabolic regulation of enzymatic activities) levels. The physiological functions of redox reactive species in the skin are listed in Table 1. To exert physiological functions, ROS and RNS are typically generated by tightly regulated enzymes such as cNO synthase and NADPH oxidase isoforms (15). On the other hand, excessive amounts of ROS may arise either from over-stimulation of NADPH oxidases or from less well-regulated sources such as the mitochondrial electron transport chain. For instance, NO is thought to function as a

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Table 1. Physiological role of reactive oxygen and nitrogen species in the skin

ROS, RNS	Function
Superoxide anion radical	Intracellular and intercellular messenger Regulator of cell growth and proliferation Regulator of stress protein induction Regulator of cytokine synthesis Enzyme activity regulator
Hydroxyl radical	Anti-microbial, anti-viral, and anti-tumor (??) action Cytokine destruction Dead cell destruction Destruction of extracellular matrix
Singlet oxygen	Intracellular and intercellular messenger Enzyme activity regulator Anti-microbial and anti-viral action Regulator of stress protein induction
Nitric oxide	Intracellular and intercellular messenger Regulator of cell growth and proliferation Regulator of cell senescence and death Regulator of vascular tonus Regulator of neurotransmitter release
Peroxynitrite	Anti-microbial, anti-viral, and anti-tumor (??) action Regulator of cell senescence and death Enzyme activity regulator Cytokine destruction Dead cell destruction Destruction of extracellular matrix
Hydrogen peroxide	Intracellular and intercellular messenger Regulator of cell growth and proliferation Regulator of cell senescence and death Regulator of stress protein induction including melanogenesis Enzyme activity regulator Regulator of cytokine synthesis

signaling molecule for the fine, steady-state control of crucial cell processes, including proliferation (10), or as a source of highly toxic oxidants utilized for cytotoxic and microbicidal activity of macrophages (16). Although we will mainly focus on the essential role of free radicals and other oxidants in the adaptation reaction and regulation of diverse cell functions, they have to be promptly detoxified in order to prevent oxidative damage to cells and extracellular matrix. As compare to the extracellular environment, the cytosol is normally maintained under strict reducing conditions. This is accomplished by the redox buffering capacity of the major intracellular thiols GSH and TRX. The high ratios of reduced to oxidized GSH and TRX are maintained by the activity of GSH reductase and TRX reductase, respectively. Both of these thiol redox systems can actively reduce hydrogen peroxide or lipid peroxides through reactions that are catalysed by GSH and TRX peroxidases, whereas GSH scavenges hydroxyl radical and singlet oxygen directly. Notably, GSH can regulate redox signaling by balancing the ratio between the level of its oxidized form (GSSG) and its reduced (GSH) form. GSH is present at millimolar concentrations in distinct intracellular compartments, including the cytosol,

the mitochondria and the nucleus. GSH itself participates in a number of physiologically important redox reactions such as regeneration of some cutaneous antioxidants (vitamin C, vitamin E and carotenoids) back to their active forms (17). Indeed, the ratio of GSH/GSSG represents a valid measurement of oxidative stress in cells, tissues, and in the whole organism (18). Activities of many skin enzymes are directly affected by reactive species. For example, both CAT and tetrahydrobiopterin dehydrogenase activities are susceptible to high levels of H₂O₂ (19), MnSOD activity drops drastically upon its exposure to peroxynitrite (20), and cyclooxygenase could be activated by NO (21).

2.1. Redox regulation of skin cell growth, differentiation, senescence, and death

The identification of redox species in the transmission of physiological processes now clearly indicates that a regulated, temporary shift of the intracellular redox state toward more oxidizing conditions represents an indispensable step along the signaling pathways that eventually lead cells to survive and proliferate, or to progress into their functional differentiation. An imbalance of its redox state can however be used by the cell to program its senescence or death. For example, apoptosis of keratinocytes plays a critical role in regulating normal epidermal physiology. In particular, programmed cell death balances proliferation to maintain epidermal thickness, contributes to stratum corneum formation and eliminates pre-malignant cells. Although the rates of keratinocyte proliferation and apoptosis are high in newborn skin and decrease with aging, they remain matched throughout the whole life to maintain epidermal homeostasis. The apoptotic gene program, unlike those associated with proliferation and differentiation, is not localized to one region but is manifested throughout all skin layers. Importantly, the integration of the apoptotic machinery into keratinocyte developmental program must be tightly regulated so that apoptosis does not precede differentiation. External stimuli may dramatically affect normal redox balance within the skin, thus, inducing a shift in the skin life circle “growth-differentiation-senescence-death”.

In cell cultures, depletion of GSH was associated to increased proliferation of fibroblasts but decreased proliferation of vascular endothelial cells (8), hence underlining the existence of a cell-specific control on the proliferation driven by GSH-GSSG balance. By contrast, a permanent shift toward an oxidizing environment in the cell predisposes to cell death due to apoptosis or, in the presence of an intense oxidizing stimulus, to necrosis (22).

TRX is a small multifunctional protein with two redox-active cysteines within a conserved active site, and it can regulate the activity of some proteins by directly binding to them. An increase in its expression was shown in keratinocytes under conditions of oxidative stress (23). There is also evidence that TRX can translocate from the cytosol to the nucleus in response to oxidant stress to regulate gene expression through Ref-1. Binding and activation of Ref-1 by TRX facilitates DNA binding of the Jun-Fos complex to the AP-1 consensus site to mediate

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transcription of AP-1-dependent genes, which include critical components for the initiation of cell proliferation (24).

The epidermal growth factor receptor (EGFR) signaling pathway governs the homeostatic maintenance and repair of epithelial tissues (25). The major EGFR-activated molecular effectors are the mitogen-activated protein kinases (MAPKs) identified as ERK1 and 2, whose induction is required for cell cycle entry (26) as well as for EGFR-dependent pro-survival mechanisms (27, 28). EGFR-initiated signals are also critical in maintaining a proliferative pool of undifferentiated cells responsible for epidermal self-renewal in the basal epidermal layer (29). There is growing experimental evidence that ROS serve as intracellular second messengers in the transmission of EGFR-initiated signals. In particular, it has been initially demonstrated that ligand-mediated EGFR activation led to a transient increase in H₂O₂ generation in the human epidermoid carcinoma cells A431, and that H₂O₂ generation was required for inhibition of the protein tyrosine phosphatases and hence activation of EGFR tyrosine kinase activity (30). Accordingly, incorporation of catalase into the cells by electroporation abrogated ligand-induced EGFR tyrosine phosphorylation and consequently the activation of this intracellular pathway. An increase in intracellular ROS concentration following EGFR activation was also reported in the human keratinocyte cell line HaCaT (31). Notably, even exogenous H₂O₂ at concentrations lower than 2 mM actively induced EGFR phosphorylation in HeLa cells, with consequent up-regulation of the PI3K-Akt-mediated survival pathway (32). These data suggest that, at these concentrations, H₂O₂ acts by promoting epithelial cell survival and proliferation. In normal keratinocytes, cell-permeable antioxidants (N-acetyl-L-cysteine, ascorbic acid) or the H₂O₂-degrading enzyme catalase can successfully inhibit the specific ligand-induced EGFR activation and the subsequent activation of ERK1/2, clearly demonstrating that H₂O₂ is an endogenous mediator required for EGFR-mediated cell activation (33). These antioxidant treatments also prevent the fast activation of EGFR due to exposure to physiological doses of UVB radiation or to exogenously added H₂O₂ (33). Active protein-tyrosine phosphatases (PTP) are known to limit EGFR functions through its dephosphorylation (34). In turn, the generation of ROS secondary to EGFR activation leads to the reversible inactivation of crucial PTPs by oxidizing the catalytic cysteine in their active site. Noteworthy, oxidative inhibition of PTP activity by ROS is the mechanism underlying the activation of EGFR by UV irradiation (35), which is implicated in keratinocyte resistance to apoptosis and enhanced proliferation, and eventually in the onset of nonmelanoma skin cancer. Concomitant to the EGFR-dependent up-regulation of ERK1/2, which represents the main protective mechanism in keratinocytes, both UVB and exogenous H₂O₂ led to the EGFR-independent and rather persistent activation of the stress-induced MAPKs identified as p38alpha/beta (36). In these experimental conditions, the selective abrogation of ERK1/2 activity by specific inhibitors resulted in the cellular death (Peus *et al.*, 1999), due to the prevalence of pro-apoptotic, oxidative

stress-specific signaling components. Indeed, a lethal cascade initiated by the early generation of UVB-induced ROS followed by activation of the Apoptosis signal regulating kinase-1 (Ask-1) was demonstrated in human keratinocytes (37). The central role of UVB-generated H₂O₂ in the initiation of the apoptotic process has been further highlighted in a recent paper reporting that over-expression of catalase, but not of CuZn-SOD, could successfully protect keratinocytes from caspase-9 activation and death (38).

Peroxideroxins (Prxs) that reduce peroxides in the presence of thioredoxin are expressed in the epidermis and the elevated expression of two isotypes of the Prx family, namely Prx I and II, characterizes its suprabasal layers. The elegant study from a Korean group (39) has shown that PrxI/II up-regulation was induced by keratinocyte differentiation. ROS seem to be signaling molecules for both the processes of PrxI/II expression and keratinocyte differentiation. As an exogenous source of ROS, UVB radiation induced the differentiation of human keratinocytes by diacylglycerol-mediated translocation of the protein kinase C (PKC) fractions (40). In human dermal fibroblasts, H₂O₂ treatment caused a slowing down of cell proliferation as it induced differentiation and the increase of SOD and CAT activities (41). Still, the regulatory mechanisms by which ROS and the antioxidants including Prxs control the differentiating processes in normal skin should be better understood. Vitamin C in its function of antioxidant seems to be a signaling molecule to trigger skin differentiation and protect against the differentiation-dependent oxidative stress (13). The vitamin C-triggered differentiation proceeds through a PKC-dependent activation of the AP-1 DNA binding activity, since vitamin C promotes protein kinase C translocation from cytosol to the membrane. The induction of PKC is strictly redox dependent because the enzyme contains redox-sensitive cysteine moieties both in its regulatory and catalytic domains (42). Vitamin C activates PKC by preventing its auto-inhibition *via* the regulatory domain or by sparing the free sulphhydryl groups necessary for catalytic activity. Skin keratinocyte death by apoptosis is regulated by AP-1 pathway and connected with p53 mitochondrial translocation. The pre-apoptotic activation of p53 induced by TPA in mouse keratinocytes is mediated by O₂⁻ produced by membrane-bound NADPH-oxidase (43). Nitric oxide and peroxynitrite are thought to be involved in the regulation of keratinocyte proliferation. This subject will be discussed in detail in the 3.3 paragraph of the present review since the NO-dependent mechanism has recently become a leading hypothesis for the keratinocyte hyperproliferation that characterizes the psoriatic plaques.

2.2. Reactive oxygen and nitrogen species and antioxidants in the skin immunity

Anatomically, skin is the first barrier to microbiological invasion. The capacity of the skin to protect the host against potentially lethal threat depends not only on its physical and structural properties but also includes an immune component. Today, the skin is regarded as the biggest immunological organ consisting of two parts. The first one is represented by static

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(continuously present in the skin) cell populations of keratinocytes, mast cells, and endothelial cells. The second part consists of numerous migrating immune cells, which move to and from the skin upon microbial stimuli. Thus, discrete T-lymphocyte subsets, granulocytes, and macrophages are recruited into the skin after its infection. On the other hand, dendritic antigen-presenting cells called Langerhans cells in the epidermis and dermal dendritic cells in dermis, being stimulated and loaded with microbial peptidic antigens, leave the skin and migrate to regional lymphoid tissues (44)

Upon a microbial, fungal, or parasite invasion into the skin, keratinocytes can immediately mount an innate immune response against invading microorganisms, since they express a variety of TLRs, the most investigated recognition receptors and the primary sensors of innate immunity (45). Each TLR is activated by distinct conserved microbial structures, including LPS, flagellin, bacterial DNA or lipoarabinomannan, and its triggering activates an inflammatory cascade, launches a quick antimicrobial reaction and directs adaptive immunity to mount a protective response. In particular, activation of the TLRs leads to enhanced expression of genes encoding for an array of antimicrobial peptides, including defensins and cathelicidins, which possess chemoattractive properties for phagocytic cells (46). In their turn, granulocytes and macrophages attracted to the infection site produce an array of antimicrobial agents like ROS, RNS, and proteases (5, 11, 15). The oxidative burst in phagocytes starts from the receptor-dependent activation of cellular membrane bound NADPH-oxidase, which primarily produces superoxide radicals. The superoxide production gives rise to H_2O_2 formation. The latter is a substrate for myeloperoxidase (MPO), an enzyme of cytoplasm granules in neutrophilic leucocytes. MPO catalyses H_2O_2 decomposition and, in the presence of chloride anions, a hypochlorite (ClO^-) formation takes place (11). Epidermal granulocytes destroy invading microorganisms *via* a potent antimicrobial arsenal of oxidants and antimicrobial agents. Thus, granulocytes being challenged with bacterial LPS over-express iNOS, which become a source of large amounts of NO. Another extremely effective anti-microbial agent is peroxynitrite, formed in the presence of NO, superoxide, H_2O_2 , and MPO. The cathelicidin LL-37 kills a broad spectrum of microbes, activates granulocyte chemotaxis, production of cytokines/chemokines and ROS. LL-37-stimulated ROS release occurs through NADPH-oxidase activation and intracellular Ca^{+2} mobilization (47). By contrast, granulocytes with impaired functions such as chemotaxis and ROS production were not able to prevent skin from recurrent bacterial infections, as was observed in the population of mentally retarded adults (48). Also knockout mice lacking expression of iNOS (iNOS $^{-/-}$) animals) or NADPH-oxidase (p47(phox $^{-/-}$) animals) showed extreme susceptibility to intradermal infection with *Francisella tularensis* (49). Granulocytes from the iNOS deficient mice produced more superoxide than wild type mice when stimulated with *Porphyromonas gingivalis* bacteria. However, iNOS ($^{-/-}$) mice inoculated with these bacteria developed more skin characteristic lesions and showed higher bacterial survival rate than the wild type animals

(50). It is interesting to note that some endogenously formed oxidized phospholipids function as a negative feedback to suppress innate immune reactions induced by LPS through blocking of inflammatory gene upregulation (51). This could partly explain the regulatory role of redox products in both initiation and cessation of the immune reaction to microbial invaders in the skin.

Dendritic cells (DCs), including epidermal Langerhans cells and dermal DCs, are specialized in the recognition and capture of foreign antigens as well as in the activation of naive T cells, all essential steps in the induction of the immune response. Finally, T lymphocytes translate antigen recognition into effector mechanisms to eliminate pathogens. The recruitment of leukocytes at the site of skin inflammation is the critical step for an efficient response to potentially dangerous signals (52). The question of life and death of DCs is closely connected with redox control of the balance between Nf κ B and JNK/AP-1 activities. Thus, specific blockade of Nf κ B in DCs induces highly elevated JNK/AP-1 activity with consequent DCs death because of increased levels of ROS and carbonyl proteins within the cells (53). There is growing evidence that the redox equilibrium of DCs influences their ability to induce T-cell activation. For example, GSH depletion interferes in IL-12 and IFN- γ expression, impairs DCs maturation, and as a result, inhibits the delayed-type hypersensitivity response (54). Pre-transplant treatment of donor or recipient skin with manganese-based complexes known as SOD- and catalase-mimicking agents leads to significant reduction of type 1 cytokine producing T-cells and delayed allograft rejection (55). More than 30 years ago, the discovery of the connection between UV radiation and the immune system triggered the field of photoimmunology. Pioneering work (56) suggested that H_2O_2 was a signal to UV-induced impairment of epidermal-derived DCs. Later, nitric oxide was widely recognized as a mediator of UV-induced immunosuppression in humans due to NO-mediated depletion of Langerhans cells in the epidermis (57, 58). A recent report from an Australian group has shown that a combination of inhibitor of NO production, iron chelator, and antioxidant polyphenol was extremely effective in the protection against UV-induced immunosuppression in the skin (59).

2.3. Antioxidants against environment-induced redox imbalance

The skin is permanently exposed to physical, chemical, and biological aggression by the environment. In general, endogenous ROS are generated in the course of metabolic processes that tend to detoxify environmental hazards. The abnormal levels of ROS induce an adaptive reaction such as expression of cytoprotective proteins, which include ROS-detoxifying enzymes. In order to protect the skin structural and functional integrity, a wide spectrum of phase I enzymes, active both in oxidation and reduction, and phase II enzymes, active in conjugation, can be found up-regulated in the skin, or can be rapidly, transcriptionally induced in response to different physical and chemical agents. The transcriptional response to these agents is typically mediated by the *cis*-acting antioxidant response element (ARE), found in the promoter of the

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encoding genes for diverse products such as several glutathione S-transferases (GSTs), metalloproteinases, NADPH:quinone oxidoreductase 1 (NQO1), UDP glucuronotransferase (UGT), gamma-glutamyl cysteine ligase, HO-1 and peroxiredoxin VI (60, 61). The major ARE-binding transcription factor is NF-E2-related factor 2 (Nrf2), which, through heteromeric interaction with the small Maf proteins, binds the ARE and initiates the *de novo* expression of detoxifying enzymes (62).

In the skin, although several Nrf2-dependent ROS-detoxifying enzymes are found to be up-regulated in wound healing, abrogation of Nrf2 expression in transgenic mice is substantially irrelevant for the healing process, whereas it is essential for the effective detoxification of chemical carcinogens (63). These observations emphasize the special chemopreventive role of Nrf2-control genes in the skin. The transcriptional activation of Nrf2-dependent genes can be induced by various chemicals, including redox active compounds such as quinones, isothiocyanates, peroxides, mercaptans, transition metals, trivalent arsenicals, and also by chemopreventive antioxidants such as dietary polyphenols (62). Some of these compounds induce Nrf2 translocation in epithelial cells via intracellular generation of ROS, in particular H₂O₂, as demonstrated for inorganic arsenic, a well-characterized carcinogen (64). By contrast, chemopreventive polyphenols may prevent inflammation and cancer by enhancing cellular antioxidant and detoxifying enzymes *via* activation of Nrf2. They may also suppress the induction or over-amplification of pro-inflammatory and growth-promoting genes driven by the major transcription factors NFkappaB and AP-1 (65).

Oxidative stress in the skin, induced by various agents, up-regulates trans-activating AP-1 components such as Fos and Jun and down-regulates its inhibitory components (Fra-1 and Fra-2). The shift of redox balance towards oxidative conditions facilitates phosphorylation and activation of JNK, a kinase phosphorylating c-jun and hence promoting its translocation into the nucleus. As a result, transcription of AP-1-dependent genes takes place and cells die by apoptosis. Vitamin C is an anti-apoptotic antioxidant, which provides the redox-dependent inhibition of JNK as well as redox-dependent activation of Fra-1 and Fra-2, two inhibitory components of AP-1 machinery (13).

Organic peroxides, widely used in the chemical and pharmaceutical industries, promote skin cancers and cause epidermal hyperplasia. Their molecular mechanism is thought to be tightly connected with the induction of AP-1 pathway through free radical formation and oxidative stress, revealed by decreased levels of GSH, accumulation of peroxidative products, and chronic inflammation in the skin exposed to peroxides (66). Peroxiredoxin 6 is an enzyme that detoxifies H₂O₂ and organic peroxides. Transgenic mice over-expressing this enzyme showed enhanced resistance to the toxicity of various exogenous inducers of oxidative stress and strongly reduced the number of apoptotic cells after UVA and UVB irradiation *in vivo* and *in vitro* (67).

3. SKIN INFLAMMATION AND REDOX IMBALANCE

The inflammatory reaction in various organs has long been connected with overproduction of ROS, RNS, and high levels of products of lipid peroxidation. Despite of a cascade of publications on the topic, the cellular and molecular mechanisms of redox regulation of pro-inflammatory and anti-inflammatory reactions in the skin are still quite obscure. Therefore therapeutic implications of pro- and antioxidants have not brought significant clinical results as yet. Here, we are reviewing works on the involvement of ROS and RNS in the acute after-wound skin inflammatory reaction, in the UV- and other environment hazards-induced cutaneous inflammation as well as in some socially relevant chronic inflammatory skin diseases such as psoriasis, atopic and contact dermatitis.

3.1. Acute inflammatory reaction to skin wounding

Damage to the skin triggers a cascade of events that leads to rapid repair of the wound through the concerted effort of the wounded cell layers which is accompanied, and also partially regulated, by a robust, transient inflammatory response, in which first neutrophils and then macrophages migrate from the nearby tissues and the circulation to the wound site (68). Epidermal cells have a major role in mounting the wound healing response through a complex program of *de novo* gene expression, which include upregulation of autocrine growth factors, cytokines, chemokines and adhesion molecules (52).

During the inflammatory phase of wound healing, a variety of molecular mechanisms guarantee that a massive infiltrate of neutrophils and macrophages invade the wound. These phagocytic cells possess a well-characterized superoxide anion-generating oxidase identified as NADPH-oxidase, localized at the phagosomal membranes and the plasma membrane, and capable of producing large amounts of ROS, in turn required for their microbicidal functions. Superoxide anion can dismutate spontaneously or by the action of SOD to generate easily diffusible H₂O₂, which is converted to the highly reactive hydroxyl radicals in the presence of transition metals through the Fenton reaction. Activated macrophages also express high levels of iNOS, and consequently provide to robustly increase NO, another microbicidal agent, at the wound site (8, 16). These leukocytic populations release high levels of pro-inflammatory cytokines, in particular IL-1 and TNF-alpha in the epidermal and dermal environment, and hence provide the strongest stimulus for the enhancement of iNOS expression also by resident cells (69). Hence, the inflammatory phase of wound healing can be conceived as a condition of intense oxidative stress for the skin. Due to their special reactivity with the macromolecular constituents of the cell, high levels of ROS and RNS however may represent a serious danger for tissue integrity. As a consequence, the wound is involved in an active program of up-regulated expression of ROS-detoxifying enzymes, as demonstrated by experiments on animal models (70). These enzymes include both the cytosolic CuZn-SOD and the mitochondrial Mn-SOD, active in the detoxification of superoxide generated by

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phagocytes. Distinct enzymes involved in H₂O₂ or phospholipid hydroperoxide degradation, including PRx VI, the Se-containing enzyme GPx and CAT were found strongly up-regulated in the healing epidermis (71, 72). Several antioxidant therapies using naturally occurring antioxidants have shown promising results in enhancing wound repair. For instance, flavonoid catechins (73) and phenylpropanoids (74), which exert effective antioxidant, free-radical-scavenging, and iron chelating effects, significantly accelerate full-thickness wound closure.

Despite the complex, conflicting understanding of its possible contribution to skin inflammation (69), NO signaling certainly plays a beneficial role in the distinct aspects of wound closure (10, 75, 76, 77). In particular, iNOS-generated NO is crucially involved in the re-epithelialization of the wound (78, 79), and in tissue remodeling, with increased collagen production, and hence tensile strength, at the wound site (80). In addition, a relevant contribution of NO to wound healing is linked to its regulatory effect on angiogenesis. Indeed, the wound repair process is angiogenesis-dependent, and new blood vessel formation must occur for an injured tissue to progress toward complete tissue regeneration. Notably, there is evidence that iNOS drives the expression of VEGF, so that impaired VEGF expression and delayed wound healing can be observed in iNOS knockout mice (81). Moreover, topically applied gaseous NO accelerate substantially wound healing (82).

During the early inflammatory phase of wound healing, the oxidative metabolism of arachidonic acid (AA), whose release from the membrane phospholipids is catalyzed by the activation of phospholipase A₂, provides a number of pro-inflammatory bioactive mediators that are favorably associated with tissue repair. In particular, up-regulated expression of cyclooxygenase-2 and its metabolic products, in particular prostaglandin E₂ (PGE₂), increase the rate of repair in its distinct phases, including re-epithelialization and efficient neo-angiogenesis through up-regulation of VEGF expression (77).

In the healing wound, there is a remarkable up-regulation of the inducible isoform 1 of the enzyme dedicated to the degradation of heme, namely HO-1, suggesting its active contribution to the healing process (70). Expression of HO-1 can be induced not only by its own substrate, heme, but also by a variety of physical and chemical stimuli, including UVB, oxidant species or organic chemicals. Degradation of heme leads to the formation of carbon monoxide, iron and biliverdin, whose product of reduction, bilirubin, is a potent antioxidant. Notably, HO-1 can be found up-regulated not only in the hyperproliferative epithelium, but also in the inflammatory cells of the granulation tissue (83). Apart from its direct antioxidant cytoprotection and detoxification of heme, HO-1 appears involved in the up-regulation of VEGF expression in epithelial cells (84), and hence could participate in the mechanisms that

tend to guarantee neo-angiogenesis during tissue repair.

3.2. UV-induced chronic inflammation in the skin

The majority of skin damaging effects of UV light occurs *via* the generation of free radicals. The UV-induced ROS formation starts from absorption of UV light by endogenous chromophores such as trans-urocanic acid, melanins, porphyrins, flavins, quinines, tryptophan, hydroxypyridines, and glycation end-products (85). After absorption of the light energy, the chromophores become activated (excitation state). The activated chromophores may directly react with the target molecule, thus, transforming it in a corresponding free radical. In the presence of oxygen, superoxide radicals or singlet oxygen are formed. They further induce oxidative modifications of lipids, proteins, and DNA. Chronic inflammation connected with UV exposure has been reported long time ago (86). The heliodermatitis was morphologically characterized by dermal infiltrate of mast cells, histiocytes, and CD4⁺ T-cells. The role of oxidative stress in the generation of UV-induced pro-inflammatory skin environment has been extensively investigated (87). Notably, UV radiation increases the activity of xanthine oxidase in human keratinocytes, with a consequent boost in the generation of the superoxide anion (88). Sustained production of ROS in the epidermal keratinocytes following exposure to UV depletes these cells of their free radical scavenger systems (13, 89). On the other hand, supplementation with antioxidants protects keratinocytes and skin in general against UV-induced ROS-mediated damage (13, 90). UV radiation at doses lower than the minimal erythematous dose enhances phospholipase activity and upregulates COX, thus increasing prostaglandin production (mainly, PGE₂). PGE₂ is known to play a key role in the induction of erythematous and edematous skin responses (91). In addition, UV radiation up-regulates keratinocyte expression and release of potent pro-inflammatory cytokines such as IL-1 and TNF-alpha, which establish an overt pro-inflammatory environment also through autocrine stimulation of distinct chemokines, the major chemoattractants of the diverse leukocyte populations into the skin (92). In the mouse skin, a single inflammatory dose of UV radiation causes infiltration by granulocytes and macrophages (93), and permanence of intense inflammatory infiltrate is determinant for tumor growth promotion (94). A large amount of literature supports the role of inflammation in driving tumor progression, and, consistently, anti-inflammatory drugs were shown to reduce the incidence of cancer. Inhibition of the PGE₂-generating enzyme COX2 can prevent skin cancers in the mouse, including UV-induced carcinogenesis (95). Experimental evidence collected on human skin demonstrates that NO is a major contributor to the expression of UV-induced erythema (96). Indeed, intradermal administration of L-NAME prior or after UVB exposure prevents the inflammatory response. In the hours following UV exposure, iNOS protein is strongly up-regulated in both dermis and epidermis, reasonably as a consequence of direct UV induction combined with the increased concentration of local pro-inflammatory cytokines involved in iNOS expression (69). Notably, iNOS prolongs and boosts the local release of NO, which is

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also involved in the reduction of CD1a+ Langerhans cells in the irradiated skin and consequently in its immunosuppression (97). In their whole, these independent observations point at the complexity of NO effects in the skin, only partially defined so far. Topical application of some botanical antioxidants such as green tea polyphenols prevent a number of UV-induced inflammatory features, for example, formation of inflammatory infiltrate, erythema, the increase in prostaglandin synthesis and the induction of myeloperoxidase activity (98). Two other botanical antioxidants, silimarín and genistein decrease significantly the inflammatory edema and suppress skin hypersensitivity as a consequence of solar UV irradiation (99, 100). A recent publication shows that vitamin C reduces the inflammatory response to UVB radiation in terms of reduced transcript and protein expression of the pro-inflammatory cytokines IL-8 and MCP-1 (101).

3.3. Psoriasis

Affecting 2-3% of the population, psoriasis is a common chronic inflammatory skin disorder. It is characterized by complex epidermal abnormalities and a prominent inflammatory cell infiltration (102). Keratinocyte hyperproliferation and disturbed differentiation are hallmarks of psoriatic lesions. In addition, evidence of aberrant expression of apoptosis-related molecules and senescence-associated signals further complicates the understanding of the concomitant processes going on in the psoriatic skin. T cell-derived IFN- γ is overexpressed in psoriasis, consistent with the predominant Th1 immunopathology observed in this disease. Also leukocyte-released TNF- α certainly plays a central role in its pathogenesis in the majority of patients, as currently demonstrated by the clinical efficacy of anti-TNF- α medicinal preparations (103). Both TNF- α and IFN- γ are potent inducers of ROS generation in non-phagocytic cells (8). In particular, there is evidence that a mitochondrial source of ROS is required for the strong TNF- α -driven activation of NF- κ B, which represents an essential step in the pro-inflammatory process of cytokine and chemokine up-regulation by the resident cells (104). In addition, TNF- α and IFN- γ upregulate keratinocyte expression of EGFR ligands (105), thus, promoting EGFR-dependent generation of ROS. In their whole, these mechanisms conceivably precipitate a condition of intense oxidative stress in the psoriatic lesion, which is dramatically aggravated by the “respiratory burst”, the large amount of superoxide anion released by the numerous neutrophils and macrophages that infiltrate the chronically inflamed skin. Strong up-regulation of HO-1 in the psoriatic lesions may represent a cytoprotective mechanism against local oxidative stress (83).

Indeed, there are many features of severe oxidative stress in the patients with active psoriasis. In the plasma and red blood cells of patients with active psoriasis, increased levels of malonyl dialdehyde (MDA) were interpreted as the fingerprint of the exhaustion of natural enzymatic and non-enzymatic antioxidant defenses and consequently the prevalence of deleterious peroxidative processes in the cell membranes and plasma lipids (106, 107). Another study has shown that erythrocytes from

psoriatic patients present a statistically significant decrease in erythrocyte SOD and GPx, but failed to find any abnormality in the MDA levels in the serum of these patients. MDA levels were increased exclusively in the lesional tissues (108). Significantly decreased antioxidant potential of the plasma, higher-than-normal expression of SOD, and elevated MDA levels were also reported. However there was no correlation between these parameters and the disease severity (109). Taken together, these results support the hypothesis that an imbalance in the oxidant-antioxidant system is a characteristic feature in psoriatic patients (110). Its role in the pathogenesis of psoriasis remains unclear so far.

It is noteworthy that pro-inflammatory mediators remarkably up-regulated in the psoriatic lesions, including TNF- α , IFN- γ , and IL-8, are strong inducers of iNOS expression and NO release from the epidermal keratinocytes (111, 1120). Being a potent regulator of keratinocyte growth and differentiation, NO has been recently considered a key player in the pathogenesis of psoriasis. Indeed, a large body of work suggests that NO is pro-inflammatory in the skin. Application of a NO-releasing cream on the skin of healthy subjects elicits an inflammatory response and the characteristic loss of Langerhans cells, this last recently correlated to up-regulated iNOS activity during the UVB-induced erythematous response (97, 113). In contrast with these observations is the evidence that application of a NO-releasing ointment on active psoriatic lesions leads to disease regression, possibly due to prominent NO-dependent down-regulation of chemokine expression and consequently to inhibition of T cell and macrophage attraction into the skin (114). This report emphasizes a possible beneficial effect of NO on this chronic disorder and suggests that the actual levels of NO are lower-than-normal in the lesions. The evidence that arginase 1 is aberrantly over-expressed in the lesional skin of psoriatic patients suggests the possibility that arginine, which is the substrate for NO generation by iNOS, might be not adequately available for iNOS activity (115). In addition, the presence of abundant superoxide anion could rapidly trap NO to generate peroxynitrite, a cytotoxic agent that nitrosylates thiol groups and leads to DNA break (116). However, recent studies have demonstrated that endogenously produced NO increase the expression of CuZn SOD mRNA in keratinocytes (117). With regards to pathogenic keratinocyte proliferation in psoriasis, NO has shown to exhibit biphasic action providing a proliferative signal at low concentrations, and inducing cell cycle arrest at high levels.

Surprisingly, the clinical evidence that modulation of the skin's redox state can be used therapeutically to modulate the inflammatory response in psoriasis shows positive clinical outcomes with therapies mainly with pro-oxidant and pro-inflammatory mechanisms of action. For example, PUVA therapy widely used in the treatment of psoriasis leads to the formation of singlet oxygen in the skin (6). Solar irradiation is also known to improve substantially the clinical conditions of the patients with psoriasis, diminishing inflammatory skin lesions. The

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whole spectrum sun light induces pro-inflammatory IL-6 production accompanied by increased levels of 8-isoprostane, a marker of oxidative damage to skin lipids (118). Broad band UVB phototherapy was successfully introduced into therapeutic protocols for psoriasis. The UVB phototherapy is also associated with a sharp increase in the levels of TBA products and nitrite/nitrate concentration in the plasma of patients subjected to a 1-2 month long phototherapy (119). Another relatively effective systemic treatment of psoriasis includes anti-cancer cytostatic preparations such as cyclosporine and methotrexate. After the course of systemic therapy with methotrexate, elevated serum nitrite/nitrate levels were found (120). Fumaric acid esters are used for the systemic therapy of psoriasis with high clinical efficacy. The therapy with fumaric acid esters result in the induction of superoxide production by circulating blood monocytes (121). Anthralin is a well-established topical therapeutic agent for psoriasis. Its positive clinical effect seems to be mediated by the activation of EGF receptor in keratinocytes, which leads to H₂O₂ generation (122). In general, pro- and antioxidant properties of anthralin and some of its derivatives are reviewed (123).

3.4. Atopic and contact dermatitis

Oxidative stress is widely implicated in the molecular mechanisms leading to the hypersensitivity reaction identified as contact dermatitis, a very common disease in general and occupational dermatology (124). Irritant contact dermatitis, which accounts for 50-80% of all cases, is a non-immunological, local inflammatory skin reaction in response to the transcutaneous penetration of irritant substances, whereas allergic contact dermatitis is a cell-mediated immune-specific type IV hypersensitivity reaction.

In the case of irritant contact dermatitis, the leukocyte infiltrate is dominated by neutrophils and macrophages attracted into the skin by keratinocyte-released chemokines, essentially represented by IL-8 and MCP-1. Induction of chemokine expression is a direct response of the resident cell populations to the irritant or sensitizer (125). Molecular triggers of chemokine release from keratinocytes are pro-inflammatory cytokines, in particular IL-1 and TNF- α . In allergic contact dermatitis, numerous antigen-specific memory T cells invade the epidermis and dermis upon skin re-exposure to the antigen, attracted by T cell-specific chemokines locally released by keratinocytes and professional antigen-presenting cells. Similar to the psoriatic lesion, high levels of TNF- α and IFN- γ are released by activated leukocytes, thus, stimulating pro-inflammatory program in the resident cells of the contact dermatitis skin.

Examples of irritants or antigens with oxidizing properties are numerous, and include hydroperoxides and peroxides, metal salts such as Nickel (II) or Chromium (VI), quinones and primary amines (126). All these substances may determine an oxidative stress in the cell and affect its molecular mechanisms. Moreover, some molecules to become antigenic should be chemically modified in redox reactions. The pro-inflammatory activity of ROS-

generating chemicals in the skin is exemplified by the induction of irritant contact dermatitis through intradermal injection of hydrogen-peroxide producing enzymes including glucose oxidase. The inflammatory reaction in the skin was neutralized by simultaneous administration of CAT or SOD (127). Systemic or topical treatment with N-acetylcysteine prior to epicutaneous application of 2,4,6-trinitro-1-chlorobenzene, a strong sensitizer and oxidant, reduces all the inflammation-related parameters, including expression of pro-inflammatory cytokines, edema and leukocyte infiltration (128). The group of sensitizers with a dinitrohalogenbenzene structure, namely 2,4-dinitro-1-fluorobenzene and 2,4-dinitro-1-chlorobenzene, were shown to irreversibly inhibit mammalian thioredoxin reductase (129), thus, eliminating thioredoxin from the maintenance of intracellular redox balance. In the positive patch test for allergic contact dermatitis to Nickel (II), an abnormal elevation of the GSSG/GSH ratio and an increase in the levels of iron have been detected in the skin lesion (130). In contrast, induction of allergic contact dermatitis to polyaromatic hydrocarbons requires the biosynthesis of the reactive oxidative intermediate on the cytochrome P450-dependent enzymes (131). Also paraphenylenediamine, a known trigger of allergic contact dermatitis in the industrialized world, requires a cytochrome P450-dependent transformation to become reactive and elicit allergic inflammatory reaction (132). Finally, the depletion of the enzymatic and/or non-enzymatic antioxidant systems may create oxidative stress, which triggers pro-inflammatory metabolism. In particular, activation of the redox-sensitive NF κ B pathway leads to the *de novo* expression of a plethora of cytokines and chemokines by resident cells (126, 133), and eventually to tissue damage due to massive leukocyte infiltration.

An increase in iNOS was found immunohistochemically both in irritant and allergic contact dermatitis (134). The increased levels of NO found in the contact dermatitis skin may partly explain enhanced proliferation of epidermal cells, neutrophil accumulation and local vasodilatation (135). Indeed, the iNOS inhibition by aminoguanidine prior to immune challenge reduces the inflammatory response to both picryl chloride and 2,4-dinitro-1-fluorobenzene, known as skin sensitizers (136, 137). In humans, topical application of a NO-releasing cream provides a stimulus to induce an irritant contact dermatitis (113), although another report suggests that NO had a limited role in irritant contact dermatitis (138).

3.5. Chronic skin inflammation and immunosuppression as risk factors for skin tumorigenesis

Skin cancer is currently the most common type of cancer in humans, with an alarming increase in its incidence everywhere around the globe. The fundamental role of redox changes favoring oxidative stress in all crucial steps of carcinogenesis has been extensively investigated (139, 140). The concept that chronic inflammation is a critical component of tumor promotion is now widely accepted and the mechanisms underlying inflammation-sustained tumor promotion have been partially identified (141). The observation that tumors arise in sites of

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persistent infection, chronic irritation and inflammation is particularly strong in the skin (142). Chronic infiltration of the skin by activated phagocytes producing excessive amounts of ROS and RNS induces oxidative DNA damage in rapidly proliferating skin cells. In turn, high and chronic local overproduction of superoxide and nitric oxide results in peroxynitrite formation. The latter is recognized as a potent oxidant of DNA and a direct mutagen (143). Moreover, nitrosylation of biological amines may result in the formation of carcinogenic nitrosamines. Sustained generation of nitrosamines in the course of chronic inflammation is thought to provide a strong carcinogenic stimulus (144, 145). Peroxynitrite reacts with thiols yielding S-nitrosothiols. This crucially affects enzymes with thiol groups in their active center including DNA repair enzymes and zinc finger transcription factors.

In humans, the progression from the benign actinic keratosis to squamous cell carcinoma is associated with persistent inflammation (146). Noteworthy, the inflammation subsides once the skin tumor has progressed into squamous cell carcinoma. Potent anti-oxidant principles such as black tea polyphenols were shown to protect the mouse skin both from inflammation and following carcinogenesis induced by tumor promoters or UV (147).

The relevance of immunosuppression in carcinogenesis was first demonstrated by Fisher and Kripke, who had shown that transplantation of UV-induced skin tumours to syngenic mice resulted in tumour rejection (148). At the same time, if recipient mice have been pre-treated by sub-carcinogenic doses of UV, the rejection did not occur. These data allowed to suggesting that a low-dose UV irradiation possess an immuno-suppressive action through the development of transferable antigen-specific suppressor T-lymphocytes. The same cellular mechanism was found for acute exposure of skin to UV light, which induces a transient suppression of local immunity, including contact hypersensitivity and delayed-type hypersensitivity (149). UV-dependent immunosuppression can be reproducibly observed in humans after a single or short-term exposure to UV irradiation, although the mechanisms underlying this phenomenon are not clear as yet (150). There is a hypothesis that UV-induced DNA damage to epidermal Langerhans cells is an initial event in the cascade of UV-induced local suppression of contact hypersensitivity (151). It has long been known that UV exposure destroys Langerhans cells in the skin that results in a lack of contact hypersensitivity response. Liposome-based trans-cutaneous administration of a DNA repair enzyme in humans prevents acute effects of UV irradiation including up-regulation of TNF-alpha and IL-10 (152). Another photosensitive regulator of immune response in the skin is *trans*-urocanic acid subjected to photo-isomerization to *cis*-urocanic acid, in turn known as immunosuppressor (153).

It seems that alterations in cell redox homeostasis are of extreme importance for the UV-induced local and systemic immuno-suppression (150). For example, UV-induced free radical generation contributes to

the oxidation of membrane phosphatidylcholine leading to the formation of its derivative platelet activating factor (PAF) with strong immuno-modulating properties (154). Besides UV light, activation of PAF synthesis can be triggered by a variety of cell stressors, including DNA damaging agents and powerful oxidants (152). The main cellular targets for PAF in the skin are dermal mast cells. In response to PAF, these cells release large amounts of downstream mediators like IL-10, which is currently considered centrally responsible for the systemic immune suppression (150).

An increasing body of evidence now clearly indicates that antioxidants can efficiently oppose redox-mediated gene mutations, chronic inflammation in the skin, and immunosuppression. As a result, they may be feasible for chemoprevention of carcinogenesis in the skin. Accordingly, addition of ROS scavengers or NO inhibitors to sunscreen preparations significantly reduces UV-associated inflammatory and carcinogenic features in the mouse skin (156). Vitamin E protects mouse skin against UV-induced DNA damage, inflammation and carcinogenesis (157). Topical application of tannic acid (158) and epigallocatechin-3-gallate (159), polyphenolic antioxidants, reduce UV carcinogenesis in animals. Protective effects of topical/systemic administration of antioxidants against UV-associated impairment of antigen presentation (160) and local immuno-suppression have been revealed by the restoration of contact hypersensitivity (161) and inhibition of antigen-specific tolerance (162).

4. CONCLUSIONS AND PERSPECTIVES

In this review we attempted to show vitally essential functions of redox reactive species such as ROS, RNS and reactive lipid species as well as antioxidants to maintain the normal structure and physiology of the skin. The physiological role of free radicals and corresponding reactive species as second messengers from cell receptors to the nucleus is becoming more and more evident. This issue is presently discussed in the literature more extensively than historically dominant issues concerning the toxicity of free radical species. The redox regulated molecular events in a single cell are now translated into cellular behavior and interplay between different cells, coordinated actions which provide proper protection of the whole organism against hostile environment. According to a steadily growing mountain of evidence, the complex system of redox balancing in the skin may have also a key role in the biochemical translation of external signals and in the adaptation of the skin to continuously changing exposure to physical, chemical, and biological hazards. Depending on the nature, intensity and duration of the external factor exposure and genetic peculiarities of the organism, sometimes, normally tightly regulated systems of redox balance fail to provide an adequate adaptive protection. Then, the redox-mediated skin pathology such as chronic inflammation or cancer develops. There are many examples of positive clinical effects of regulators of redox balance (both pro- and antioxidants) in the prevention and the cure of such pathologies. To develop new preventive and therapeutic strategies, a better

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knowledge on the molecular and cellular mechanisms of redox sensitivity and cross-talk between different skin cell types should be obtained.

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6. REFERENCES

1. P.M. Elias: Stratum corneum defensive functions: an integrated view. *J Invest Dermatol* 125, 183-200 (2005)
2. J.J. Thiele, C. Schroeter, S.N. Hsieh, M. Podda and L. Packer: The antioxidant network of the stratum corneum. *Curr Probl Dermatol* 29, 26-42 (2001)
3. L. Packer and G. Valacchi: Antioxidants and the response of skin to oxidative stress: vitamin E as an indicator. *Skin Pharmacol Appl Skin Physiol* 15, 282-290 (2002)
4. E. Pelle, T. Mammone, D. Maes and K. Frenkel: Keratinocytes act as a source of reactive oxygen species by transferring hydrogen peroxide to melanocytes. *J Invest Dermatol* 124, 793-797 (2005)
5. B.M. Babior: NADPH oxidase: an update. *Blood* 93, 1464-1476 (1999)
6. L.-O. Klotz, N.J. Holbrook and H. Sies: UVA and singlet oxygen as inducers of cutaneous signaling events. *Curr Probl Dermatol* 29, 95-113 (2001)
7. R. Rodriguez and R. Redman: Balancing the generation and elimination of reactive oxygen species. *Proc Natl Acad Sci USA* 102, 3175-3176 (2005)
8. V.J. Thannickal and B.L. Fanburg: Reactive oxygen species in cell signaling. *Am J Physiol Lung Cell Mol Physiol* 279, L1005-L1028 (2000)
9. A.J. Kozak, F. Liu, P. Funovics, A. Jacoby, R. Kubant and T. Malinski: Role of peroxynitrite in the process of vascular tone regulation by nitric oxide and prostanoids: a nanotechnological approach. *Prostaglandins Leukot Essent Fatty Acids* 72, 105-113 (2005)
10. A. Soneja, M. Drewes and T. Malinski: Role of nitric oxide, nitrooxidative and oxidative stress in wound healing. *Pharmacol Rep* 57, 108-119 (2005)
11. E.T. Denisov and I.B. Afanas'ev: Oxidation and Antioxidants in Organic Chemistry and Biology. CBC Taylor & Francis Group, Boca Raton-London- New York-Singapore (2005)
12. J. Fuhs, T. Herrling. and N. Groth: Detection of free radicals in skin: a review of the literature and new developments. In: Oxidants and Antioxidants in Cutaneous Biology. *Curr Probl Dermatol*. Basel, Karger. Eds.: J. Thiele and P. Elsner, 29, 1-17 (2001)
13. M.V. Catani, I. Savini, A. Rossi, G. Melino and L. Avigliano: Biological role of vitamin C in keratinocytes. *Nutrition Reviews* 63, 81-90 (2005)
14. M.A. Pathack and K. Stratton: Free radicals in human skin before and after exposure to light. *Arch Biochem Biophys* 123, 463-476 (1968)
15. W. Droge: Free radicals in the physiological control of cell function. *Physiol Reviews* 82, 47-95 (2002)
16. J.D. MacMicking, C. Nathan, G. Hom, N. Chartrain, D.S. Fletcher, M. Trumbauer, K. Stevens, Q.W. Xie, K. Sokol, N. Hutchinson, H. Chen and J.S. Mudgett: Altered responses to bacterial infection and endotoxic shock in mice lacking inducible nitric oxide synthase. *Cell* 81, 641-650 (1995)
17. R. Masella, R. Di Benedetto, R. Vari, C. Filesi and C. Giovannini: Novel mechanisms of natural anti-oxidant compounds in biological systems: involvement of glutathione and glutathione-related enzymes. *J Nutr Biochem* 16, 577-586 (2005)
18. A. Pastore, G. Federici, E. Bertini and F. Piemonte: Analysis of glutathione: implication in redox and detoxification. *Clin Chim Acta* 333, 19-39 (2003)
19. K.U. Schallreuter: A review of recent advances on the regulation of pigmentation in the human epidermis. *Cell Mol Biol* 45, 943-949 (1999)
20. L.A. Macmillan-Crow and D.L. Cruthirds: Invited review: manganese superoxide dismutase in disease. *Free Rad Res* 34, 325-336 (2001)
21. D. Salvemini, M.G. Currie and V. Mollace: Nitric oxide-mediated cyclooxygenase activation. A key event in the antiplatelet effects of nitrovasodilators. *J Clin Invest* 97, 2562-2568 (1996)
22. M. Valko, D. Leibfritz, J. Moncol, M.T.D. Cronin, M. Mazur and J. Telse: Free radicals and antioxidants in normal physiological functions and human diseases. *Int J Biochem Cell Biol* 39, 44-84 (2007)
23. Y. Sachi, K. Hirota, H. Masutani, K. Toda, T. Okamoto, M. Takigawa and J.Yodoi: Induction of ADF/TRX by oxidative stress in keratinocytes and lymphoid cells. *Immunol Lett* 44, 189-193 (1995)
24. K. Hirota, M. Matsui, S. Iwata, A. Nishiyama, K. Mori and J. Yodoi: AP-1 transcriptional activity is regulated by a direct association between thioredoxin and Ref-1. *Proc Natl Acad Sci USA* 94, 3633-3638 (1997)
25. M. Jost, C. Kari and U. Rodeck: The EGF receptor - an essential regulator of multiple epidermal functions. *Eur J Dermatol* 10, 505-510 (2000)
26. R. Wetzker and F.D. Böhmer: Transactivation joins multiple tracks to the ERK/MAPK cascade. *Nat Rev Mol Cell Biol* 4, 651-857 (2003)

Redox processes in skin physiology and inflammation

27. M. Jost, T.M. Huggett, C. Kari and U. Rodeck: Matrix-independent survival of human keratinocytes through an EGF receptor/MAPK-kinase-dependent pathway. *Mol Biol Cell* 12, 1519-1527 (2001)
28. B. Canguilhem, A. Pradines, C. Baudouin, C. Boby, I. Lajoie-Mazenc, M. Charveron and G. Favre: RhoB protects human keratinocytes from UVB-induced apoptosis through epidermal growth factor receptor signaling. *J Biol Chem* 280, 43257-43263 (2005)
29. I. Haase and N. Hunzelmann: Activation of epidermal growth factor receptor/ERK signaling correlates with suppressed differentiation in malignant acanthosis nigricans. *J Invest Dermatol* 118, 891-893 (2002)
30. Y.S. Bae, S.W. Kang, M.S. Seo, I.C. Baines, E. Tekle, P.B. Chok and S.G. Rhee: Epidermal growth factor (EGF)-induced generation of hydrogen peroxide. *J Biol Chem* 272:217-221, 1997
31. R. Goldman, S. Moshonov, X. Chen, A. Berchansky, G. Furstenberger and U. Zor: Crosstalk between elevation of intracellular $[Ca^{2+}]$, reactive oxygen species and phospholipase A2 stimulation in a human keratinocyte cell line. *Adv Exp Med Biol* 433, 41-45 (1997)
32. X. Wang, K.D. McCullough, T.F. Franke and N.J. Holbrook: Epidermal growth factor receptor-dependent Akt activation by oxidative stress enhances cell survival. *J Biol Chem* 275, 14624-14631 (2000)
33. D. Peus, R.A. Vasa, A. Meves, M. Pott, A. Beyerle, K. Squillace and M.R. Pittelkow: H_2O_2 is an important mediator of UVB-induced EGF receptor phosphorylation in cultured keratinocytes. *J Invest Dermatol* 110, 966-971 (1998)
34. Y. Xu, V. Grachtchouk, J.J. Voorhees and G.J. Fisher: Receptor-type protein-tyrosine phosphatase¹ regulates epidermal growth factor receptor function. *J Biol Chem* 280, 42694-42700 (2005)
35. Y. Xu, Y. Shao, J.J. Voorhees and G.J. Fisher: Oxidative inhibition of receptor-type protein-tyrosine phosphatase¹ by ultraviolet irradiation activates epidermal growth factor receptor in human keratinocytes. *J Biol Chem* 281, 27389-27397 (2006)
36. D. Peus, R.A. Vasa, A. Beyerle, A. Meves, C. Krautmacher and M.R. Pittelkow: UVB activates ERK1/2 and p38 signaling pathways via reactive oxygen species in cultured keratinocytes. *J Invest Dermatol* 112, 751-756 (1999)
37. A. Van Laethem, K. Nys, S. Van Kelst, S. Claerhout, H. Ichijo, J.R. Vanderheede, M. Garmyn and P. Agostinis: Apoptosis signal regulating kinase-1 connects reactive oxygen species to p38 MAPK-induced mitochondrial apoptosis in UVB-irradiated human keratinocytes. *Free Radic Biol Med* 41, 1361-1371 (2006)
38. H.R. Rezvani, F. Mazurier, M. Cario-André, C. Pain, C. Ged, A. Taïeb and H. de Verneuil: Protective effects of catalase overexpression on UVB-induced apoptosis in normal human keratinocytes. *J Biol Chem* 281, 17999-18007 (2006)
39. S.J. Yun, J.J. Seo, J.Y. Chae and S.-C. Lee: Peroxiredoxin I and II are up-regulated during differentiation of epidermal keratinocytes. *Arch Dermatol Res* 10, 403-418 (2005)
40. M.S. Matsui, N. Wang and V.A. DeLeo: Ultraviolet radiation B induces differentiation and protein kinase C in normal human epidermal keratinocytes. *Photodermatol Photoimmunol Photomed* 12, 103-108 (1996)
41. E. Bazso-Dombi, K. Oravec, F. Jeney, K. Nagy and I. Zs-Nagy: On the useful role of OH & other free radicals in differentiation of cultured human fibroblasts. *Arch Gerontol Geriatr* 31, 233-242 (2000)
42. R. Gopalakrishna and S. Jaken: Protein kinase C signaling and oxidative stress. *Free Rad Biol Med* 28, 1349-1361 (2000)
43. Y. Zhao, L. Chaiswing, V. Bakthavatchalu, T.D. Oberley and D.K. St. Clair: Ras mutation promotes p53 activation and apoptosis of skin keratinocytes. *Carcinogenesis* 27, 1692-1698 (2006)
44. S. Pastore, F. Mascia, V. Mariani and G. Girolomoni: Keratinocytes in skin inflammation. *Expert Rev Dermatol* 1, 279-291 (2006)
45. J.E. McInturff, R.L. Modlin and J. Kim: The role of toll-like receptors in the pathogenesis and treatment of dermatological disease. *J Invest Dermatol* 125, 1-8 (2005)
46. M. Dürr and A. Peschel: Chemokines meet defensins: the merging concepts of chemoattractants and antimicrobial peptides in host defense. *Infect Immun* 70, 6515-6517 (2002)
47. Y. Zheng, F. Niyonsaba, H. Ushio, I. Nagaoka, S. Ikeda, K. Okumura and H. Ogawa: Cathelicidin LL-37 induces the generation of reactive oxygen species and release of human alpha-defensins from neutrophils. *Br J Dermatol* Epub ahead of print (2007)
48. J. Gilad, A. Borer, R. Smolyakov, K. Riesenber, F. Schlaeffer and R. Levy: Impaired neutrophil functions in the pathogenesis of an outbreak of recurrent furunculosis caused by methicillin-resistant *Staphylococcus aureus* among mentally retarded adults. *Microbes Infect* 8, 1801-1805 (2006)
49. H. Lindgren, S. Stenmark, W. Chen, A. Tarnvik and A. Sjöstedt: Distinct roles of reactive nitrogen and oxygen species to control infection with the facultative intracellular bacterium *Francisella tularensis*. *Infect Immun* 72, 7172-7182 (2004)

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50. R. Gyurko, G. Boustany, P.L. Huang, A. Kantarci, T.E. Van Dyke, C.A. Genco, and F.C. Gison 3rd: Mice lacking inducible nitric oxide synthase demonstrate impaired killing of *Porphyromonas gingivalis*. *Infect Immun* 71, 4917-4924 (2003)
51. V.N. Bochkov, A. Kadl, J. Huber, F. Gruber, B.R. Binder and N. Leitinger: Protective role of phospholipid oxidation products in endotoxin-induced tissue damage. *Nature* 419, 77-81 (2002)
52. S. Pastore, A. Cavani, C. Albanesi and G. Girolomoni: Chemokines of human skin. In: *Skin Immune System. Cutaneous Immunology and Clinical Immunodermatology*. Ed.: J.D. Bos. CRC Press, Boca Raton, FL, 373-392 (2005)
53. E. Kriehuber, W. Bauer, A.S. Charbonnier, D. Winter, S. Amatschek, D. Tamandl, N. Schweifer, G. Stingl and D. Maurer: Balance between NF-kappaB and JNK/AP-1 activity controls dendritic cell life and death. *Blood* 106, 175-183 (2005)
54. H.J. Kim, B. Barajas, R.C. Chan and A.E. Nel: Glutathione depletion inhibits dendritic cell maturation and delayed-type hypersensitivity: implications for systemic disease and immunosenescence. *J Allergy Clin Immunol* 119, 1225-1233 (2007)
55. G. Tocco, B.M. Illigens, B. Malfroy and G. Benichou: Prolongation of alloskin graft survival by catalytic scavengers of reactive oxygen species. *Cell Immunol* 241, 59-65 (2006)
56. G. Caceres-Dittmar, K. Ariizumi, S. Xu, F.J. Tapia, P.R. Bergstresser and A. Takashima: Hydrogen peroxide mediates UV-induced impairment of antigen presentation in a murine epidermal-derived dendritic cell line. *Photochem Photobiol* 62, 176-183 (1995)
57. J.M. Kuchel, R.S. Barnetson and G.M. Halliday: Nitric oxide appears to be a mediator of solar-simulated ultraviolet radiation-induced immunosuppression in humans. *J Invest Dermatol* 121, 587-593 (2003)
58. K.S. Yuen, M.R. Nearn and G.M. Halliday: Nitric oxide-mediated depletion of Langerhans cells from the epidermis may be involved in UVA radiation-induced immunosuppression. *Nitric Oxide* 6, 313-318 (2002)
59. P. A. Russo and G.M. Halliday: Inhibition of nitric oxide and reactive oxygen species production improves the ability of a sunscreen to protect from sunburn, immunosuppression and photocarcinogenesis. *Br J Dermatol* 155, 408-415 (2006)
60. H. Steinbrenner, M.C. Ramos, D. Stuhlmann, D. Mitic, H. Sies and P. Brenneisen: Tumor promoter TPA stimulates MMP-9 secretion from human keratinocytes by activation of superoxide-producing NADPH oxidase. *Free Rad Res* 39, 245-253 (2005)
61. T. Nguyen, C.S. Yang and C.B. Pickett: The pathways and molecular mechanisms regulating Nrf2 activation in response to chemical stress. *Free Radic Biol Med* 37, 433-441 (2004)
62. J.S. Lee and Y.J. Surh: Nrf2 as a novel molecular target for chemoprevention. *Cancer Lett* 224, 171-184 (2005)
63. U. auf dem Keller, M. Huber, T.A. Beyer, A. Kümin, C. Siemes, S. Braun, P. Bugnon, V. Mitropoulos, D.A. Johnson, J.A. Johnson, D. Hohl and S. Werner: Nrf transcription factors in keratinocytes are essential for skin tumor prevention but not for wound healing. *Mol Cell Biol* 26, 3773-3784 (2006)
64. J. Pi, J.M. Reece, Y. Kumagai and M.P. Waalkes: Transcription factor Nrf2 activation by inorganic arsenic in cultured keratinocytes: involvement of hydrogen peroxide. *Exp Cell Res* 290, 234-245 (2003)
65. Y.J. Surh, J.K. Kundu, H.K. Na and J.S. Lee: Redox-sensitive transcription factors as prime targets for chemoprevention with anti-inflammatory and antioxidative phytochemical. *J Nutr* 135, 2993S-3001S (2005)
66. A. Murray, E. Kisin, C. Kommineni, V. Vallyathan, V. Castranova and A. Shvedova: Pro/antioxidant status and AP-1 transcription factor in murine skin following topical exposure to cumene peroxide. *Carcinogenesis* 28: 1582-1588 (2007).
67. A. Kumin, C. Huber, T. Rulicke, E. Wolf and S. Werner: Peroxiredoxin 6 is a potent cytoprotective enzyme in the epidermis. *Am J Pathol* 169, 1194-1205 (2006)
68. P. Martin and S.J. Leibovich: Inflammatory cells during wound repair: the good, the bad and the ugly. *Trends Cell Biol* 15, 599-607 (2005)
69. M.M. Cals-Grierson and A.D. Ormerod: Nitric oxide function in the skin. *Nitric Oxide* 10, 179-193 (2004)
70. U. auf dem Keller, A. Kümin, S. Braun and S. Werner: Reactive oxygen species and their detoxification in healing skin wound. *J Invest Dermatol Symp Proc* 11, 106-111 (2006)
71. B. Munz, S. Frank, G. Hubner, E. Olsen and S. Werner: A novel type of glutathione peroxidase: expression and regulation during wound healing. *Biochem J* 326, 579-585 (1997)
72. H. Steiling, B. Munz, S. Werner and M. Brauchle: Different types of ROS-scavenging enzymes are expressed during cutaneous wound repair. *Exp Cell Res* 247, 484-494 (1999)
73. M. Kapoor, R. Howard, I. Hall and I. Appleton: The effects of epicatechin gallate on wound healing and scar formation in a full-thickness incisional wound healing model in rats. *Am J Pathol* 165, 299-300, (2004)
74. L. Korkina, E. Mikhal'chik, M. Suprun, S. Pastore and R. Dal Toso: Molecular mechanisms underlying wound

Redox processes in skin physiology and inflammation

healing and anti-inflammatory properties of naturally occurring bio-technologically produced phenylpropanoid glycosides. *Cell Mol Biol* 53, 78-83 (2007)

75. S. Frank, H. Kämpfer, C. Wetzler and J. Pfeilschifter: Nitric oxide drives skin repair: novel functions of an established mediator. *Kidney Int* 61, 882-888 (2002)

76. A. Schwentker, Y. Vodovotz, R. Weller and T.R. Billiar: Nitric oxide and wound repair: role of cytokines? *Nitric Oxide* 7, 1-10 (2002)

77. M. Kapoor, F. Kojima, I. Appleton, S. Kawai and L.J. Crofford: Major enzymatic pathways in dermal wound healing: current understanding and future therapeutic targets. *Curr Opin Investig Drugs* 7, 418-422 (2006)

78. K. Yamasaki, H.D. Edington, C. McClosky, E. Tzeng, A. Lizonova, I. Kovcsdi, D.L. Steed and T.L. Billiar: Reversal of impaired wound repair in iNOS-deficient mice by topical adenoviral-mediated iNOS gene transfer. *J Clin Invest* 101, 967-971 (1998)

79. B. Stallmeyer, H. Kampfer, N. Kolb, J. Pfeilschifter and S. Frank: The function of nitric oxide in wound repair: inhibition of inducible nitric oxide synthase severely impairs wound reepithelialization. *J Invest Dermatol* 113, 1090-1098 (1999)

80. M.R. Schaffer, U. Tantry, F.J. Thronton and A. Barbul: Inhibition of nitric oxide synthesis in wounds: pharmacology and effect on accumulation of collagen in wounds in mice. *Eur J Surg* 165, 262-267 (1999)

81. D. Most, D.T. Efron, H.P. Shi, U.S. Tantry and A. Barbul: Characterization of incisional wound healing in inducible nitric oxide synthase knockout mice. *Surgery* 132, 866-876 (2002)

82. A.B. Shekhter, V.A. Serezhenkov, T.G. Rudenko, A.V. Pekshev and A.F. Vanin: Beneficial effect of gaseous nitric oxide on the healing of skin wounds. *Nitric Oxide* 12, 210-219 (2005)

83. C. Hanselmann, C. Mauch and S. Werner: Haem oxygenase-1: a novel player in cutaneous wound repair and psoriasis? *Biochem J* 353, 459-466 (2001)

84. A. Jazwa, A. Loboda, S. Golda, J. Cisowski, M. Szelag, A. Zagorska, P. Sroczynska, J. Drukala, A. Jozkowicz and J. Dulak: Effect of heme and heme oxygenase-1 on vascular endothelial growth factor synthesis and angiogenic potency of human keratinocytes. *Free Radic Biol Med* 40, 1250-1263 (2006)

85. C.E. Cross, A. van der Vliet, S. Louie, J.J. Thiele and B. Halliwell: Oxidative stress and antioxidants at biosurfaces: plants, skin, and respiratory tract surfaces. *Environ Health Perspect* 106, 1241-1251 (1998)

86. R.M. Lavker and A. Kligman: Chronic heliodermatitis: a morphologic evaluation of chronic actinic dermal damage

with emphasis on the role of mast cells. *J Invest Dermatol* 90, 325-330 (1988)

87. G.M. Halliday: Inflammation, gene mutation and photoimmunosuppression in response to UVR-induced oxidative damage contributes to photocarcinogenesis. *Mutat Res* 571, 107-120 (2005)

88. G. Delicostantinos, V. Villiotou and J.C. Stavrides: Alterations of nitric oxide synthase and xanthine oxidase activities of human keratinocytes by ultraviolet B radiation. *Biochem Pharmacol* 51, 1727-1738 (1996)

89. M. Podda, M.G. Traber, C. Weber, L.J. Yan and L. Packer: UV radiation depletes antioxidants and causes oxidative damage in a model of human skin. *Free Radic Biol Med* 24, 55-65 (1998)

90. W. Stahl, H. Mukhtar, F. Afaq and H. Sies: Vitamins and polyphenols in systemic photoprotection. In: *Skin Aging*. Eds.: B.A. Gilchrest and J. Krutmann, Springer, Berlin-Heidelberg-New York, 113-122 (2006)

91. C.H. Kangrotondo, C.C. Miller, A.R. Morrison and A.P. Pentland: Enhanced keratinocyte prostaglandin synthesis after UV injury is due to increased phospholipase activity. *Am J Physiol* 264, C396-C401 (1993)

92. S. Pastore, F. Mascia and G. Girolomoni: The contribution of keratinocytes to the pathogenesis of atopic dermatitis. *Eur J Dermatol* 16, 125-31 (2006)

93. K.D. Cooper, N. Duraiswamy, C. Hammerberg, E. Allen, C. Kimbroughgreen, W. Dillon and D. Thomas: Neutrophils, differentiated macrophages and monocyte/macrophage antigen presenting cells infiltrate murine epidermis after UV injury. *J Invest Dermatol* 101, 155-163 (1993)

94. R. Sluyter and G.M. Halliday: Enhanced tumor growth in UV-irradiated skin is associated with an influx of inflammatory cells in the epidermis. *Carcinogenesis* 21, 1801-1807 (2000)

95. S.M. Fischer, H.H. Lo, G.B. Gordon, K. Seibert, G. Kelloff, R.A. Lubet and C.J. Conti: Chemopreventive activity of celecoxib, a specific cyclooxygenase-2 inhibitor, and indomethacin against ultraviolet light-induced skin carcinogenesis. *Mol Carcinog* 25, 231-240 (1999)

96. L.E. Rhodes, G. Belgi, L. McLoughlin, G.F. Clough and P.S. Friedmann: Ultraviolet-B-induced erythema is mediated by nitric oxide and prostaglandin E₂ in combination. *J Invest Dermatol* 117, 880-885 (2001)

97. J.M. Kuchel, R.C. Barnetson and G.M. Halliday: Nitric oxide appears to be a mediator of solar-simulated ultraviolet radiation-induced immunosuppression in humans. *J Invest Dermatol* 121, 587-593 (2003)

98. C.A. Elmetts, D. Singh, K. Tubesing, M. Matsui, S.K. Kotiyar and H. Mukhtar: Cutaneous photoprotection from

Redox processes in skin physiology and inflammation

- ultraviolet injury by green tea polyphenols. *Am Acad Dermatol* 44, 425-432 (2001)
99. S.K. Katiyar: Treatment of silymarin, a plant flavonoid, prevents ultraviolet light-induced immune suppression and oxidative stress in mouse skin. *Int J Oncol* 21, 1213-1222 (2002)
100. S. Widyarini, N. Sprinks, A.J. Husband and V.E. Reeve: Isoflavonoid compounds from red clover (*Trifolium pratense*) protect from inflammation and immune suppression induced by UV radiation. *Photochem Photobiol* 74, 465-470 (2001)
101. J.S. Kang, H.N. Kim, J. Jung, J.E. Kim, G.H. Mun, Y.S. Kim, D. Chi, D.H. Shin, Y.I. Hwang and W.J. Lee: Regulation of UVB-induced IL-8 and MCP-1 production in skin keratinocytes by increasing vitamin C uptake via the redistribution of SVCT-1 from the cytosol to the membrane. *J Invest Dermatol* 127, 698-706 (2007)
102. M.P. Schön and W.H. Boehncke: Psoriasis. *New Engl. J Med* 352, 1899-1912 (2005)
103. A. Jacobi, V. Mahler, G. Schuler and M. Hertl: Treatment of inflammatory dermatoses by tumour necrosis factor antagonists. *J Eur Acad Dermatol Venereol* 20, 1171-87 (2006)
104. P.P. Simeonova, S. Leonard, L. Flood, X. Shi and M.I. Luster: Redox-dependent regulation of interleukin-8 by tumor necrosis factor- α in lung epithelial cells. *Lab Invest* 79, 1027-1037 (1999)
105. F. Mascia, V. Mariani, G. Girolomoni and S. Pastore: Blockade of the EGF receptor induces a deranged chemokine expression in keratinocytes leading to enhanced skin inflammation. *Am J Pathol* 163, 303-312 (2003)
106. S. Briganti and M. Picardo: Antioxidant activity, lipid peroxidation and skin diseases. What's new. *J Eur Acad Dermatol Venereol* 17, 663-669 (2003)
107. B.V. Kural, A. Örem, G. Cimsit, Y.E. Yandi and M. Calapoglu: Evaluation of the etherogenic tendency of lipids and lipoprotein content and their relationships with oxidant-antioxidant system in patients with psoriasis. *Clin Chim Acta* 328, 71-82 (2003)
108. M. Yildirim, H.S. Inaloz, V. Baysa and N. Delibas: The role of oxidants and antioxidants in psoriasis. *J Eur Acad Dermatol Venereol* 17, 34-36 (2003)
109. K. Baz, M.Y.B. Cimen, A. Kokturk, A.C. Yazici, G. Eskandari, G. Ikizoglu, H. Api and U. Atik: Oxidant/antioxidant status in patients with psoriasis. *Yonsei Med J* 44, 987-990 (2003)
110. Y. Okayama: Oxidative stress in allergic and inflammatory skin diseases. *Curr Drugs Targets Inflamm Allergy* 4, 517-519 (2005)
111. D. Bruch-Gerharz, K. Fehsel, C. Suschek, G. Michel, T. Ruzicka and V. Kolb-Bachofen: A proinflammatory activity of interleukin 8 in human skin: expression of the inducible nitric oxide synthase in psoriatic lesions and cultured keratinocytes. *J Exp Med* 184, 2007-2012 (1996)
112. A.D. Ormerod, R. Weller, P. Copeland, N. Benjamin, S.H. Ralston, P. Grabowski and R. Herriot: Detection of nitric oxide and nitric oxide synthases in psoriasis. *Arch Dermatol Res* 290, 3-8 (1998)
113. A.D. Ormerod, P. Copeland, I. Hay, A. Husain and S.W. Ewen: The inflammatory and cytotoxic effects of a nitric oxide releasing cream on normal skin. *J Invest Dermatol* 113, 392-297 (1999)
114. M.L. Giustizieri, C. Albanesi, C. Scarponi, O. De Pità and G. Girolomoni: Nitric oxide donors suppress chemokine production by keratinocytes *in vitro* and *in vivo*. *Am J Pathol* 161, 1409-1418 (2002)
115. D. Bruch-Gerharz, O. Schnorr, C. Suschek, K.F. Beck, J. Pfeilschifter, T. Ruzicka and V. Kolb-Bachofen: Arginase 1 overexpression in psoriasis. *Am J Pathol* 162, 203-211 (2003)
116. E. Szabo, L. Virag, E. Bakondi, L. Gyure, G. Hasko, P. Bai, J. Hunyadi, P. Gergely and C. Szabo: Peroxynitrite production, DANN breakage and poly(ADP-ribose) polymerase activation in a mouse model of oxazolone-induced contact hypersensitivity. *J Invest Dermatol* 117, 74-80 (2001)
117. S. Frank, H. Kampfer, M. Podda, R. Kaufmann and J. Pfeilschifter: Identification of copper/zinc superoxide dismutase as a nitric oxide-regulated gene in human (HaCaT) keratinocytes: implications for keratinocyte proliferation. *Biochem J* 346, 719-728 (2000)
118. M. Kuhn, R. Weber, L. Kolbe, O. Schnorr and H. Sies: Solar-simulated radiation induces secretion of IL-6 and product of isoprostanes in human skin *in vivo*. *Arch Dermatol Res* 297, 477-479 (2006)
119. K.I. Kilinc, S.F. Girgin, I. Ertam, S. Alper, G. Ozturk and S.E. Yildirim: Broad-band ultraviolet B phototherapy is associated with elevated serum thiobarbituric acid reactive substance and nitrite-nitrate levels in psoriatic patients. *J Eur Acad Dermatol Venereol* 20, 1226-1231 (2006)
120. T.S. Tekin, N. Ilter, B. Sancak, M.G. Ozden and M.A. Gurer: Nitric oxide levels in patients with psoriasis treated with methotrexate. *Mediators Inflamm* 3, 16-43 (2006)
121. K. Zhu and U. Mrowietz: Enhancement of antibacterial superoxide-anion generation in human monocytes by fumaric acid esters. *Arch Dermatol Res* 297, 170-176 (2005)
122. D. Peus, A. Beyerle, M. Vasa, M. Pott, A. Meves and M.R. Pettelkow : Antipsoriatic drug anthralin induces EGF

Redox processes in skin physiology and inflammation

receptor phosphorylation in keratinocytes : requirement for H(2)O(2) generation. *Exp Dermatol* 13, 78-85 (2004)

123. J. Fuhs, W. Nitschman and L. Packer: Antioxidant and prooxidant effects of the antipsoriatic compound anthralin in skin and subcellular fractions. *Adv Exp Med Biol* 267, 537-541 (1990)

124. D.R. Bickers and M. Athar: Oxidative stress in the pathogenesis of skin disease. *J Invest Dermatol* 126, 2565-2575 (2006)

125. S. Grabbe, M. Steinert, K. Mahnke, A. Schwarz, T.A. Luge and, T. Schwarz: Dissection of antigenic and irritative effects of epicutaneously applied haptens in mice. Evidence that not the antigenic component but nonspecific proinflammatory effects of haptens determine the concentration-dependent elicitation of allergic contact dermatitis. *J Clin Invest* 98, 1158-1164 (1996)

126. J. Fuchs, T.M. Zollner, R. Kaufmann and M. Podda: Redox-modulated pathways in inflammatory skin diseases. *Free Radic Biol Med* 30, 337-353 (2001)

127. C.W. Trenam, A.J. Darbagh, C.J. Morris and D.R. Blake: Skin inflammation induced by reactive oxygen species (ROS): an *in vivo* model. *Br J Dermatol* 125, 325-329 (1991)

128. G. Senaldi, P. Pointaire, P.F. Piguet and G.E. Grau: Protective effect of N-acetylcysteine in hapten-induced irritant and contact hypersensitivity. *J Invest Dermatol* 102, 934-937 (1994)

129. J. Nordberg, L. Zhong, A. Holmgren and E.S. Arner: Mammalian thioredoxin reductase is irreversibly inhibited by dinitrohalobenzenes by alkylation of both the redox active selenocysteine and its neighboring cysteine residue. *J Biol Chem* 273, 10835-10842 (1998)

130. S. Kaur, M. Zilmer, M. Eisen, A. Rehema, T. Kullisaar, T. Vihalemm and K. Zilmer: Nickel sulphate and epoxy resin: differences in iron status and glutathione redox ration at the time of patch testing. *Arch Dermatol Res* 295, 517-520 (2004)

131. C. Anderson, A. Hehr, R. Robbins, R. Hasan, M. Athar, H. Mukhtar and C.A. Elmet: Metabolic requirements for induction of contact hypersensitivity to immunotoxic polyaromatic hydrocarbons. *J Immunol* 155, 3530-2537 (1995)

132. S. Sieben, Y. Kawakubo, T. Al Masaoudi, H.F. Merk and B. Blomeke: Delayed-type hypersensitivity reaction to paraphenylenediamine is mediated by 2 different pathways of antigen recognition by specific alphabeta human T-cell clones. *J Allergy Clin Immunol* 109, 1005-1011 (2002)

133. J.J. Haddad: Redox regulation of pro-inflammatory cytokines and I κ B/NF- κ B nuclear translocation and activation. *Biochem Biophys Res Commun* 296, 847-856 (2002)

134. A.D. Ormerod, C.M. Dwyer, A. Reid, P. Copeland and W.D. Thomson: Inducible nitric oxide synthase demonstrated in allergic and irritant contact dermatitis. *Acta Derm Venereol* 77, 436-440 (1997)

135. M.M. Teixeira, T.J. Williams and P.G. Hellewell: Role of prostaglandins and nitric oxide in acute inflammatory reactions in guinea-pig skin. *Br J Pharmacol* 110, 1515-1521 (1993)

136. H. Morita, M. Hori and Y. Kitano: Modulation of picryl chloride-induced contact hypersensitivity reaction in mice by nitric oxide. *J Invest Dermatol* 107, 549-552 (1996)

137. R. Ross, C. Gillitzer, R. Klein, J. Schwing, H. Kleinert, U. Forstermann and A.B. Reske-Kunz: Involvement of NO in contact hypersensitivity. *Int Immunol* 10, 61-69 (1998)

138. J. Wallengren and B. Larsson: Nitric oxide participates in prick test and irritant patch test reactions in human skin. *Arch Dermatol Res* 293, 121-125 (2001)

139. J.E. Klaunig and L.M. Kamendulis: The role of oxidative stress in carcinogenesis. *Annu Rev Pharmacol Toxicol* 44, 239-267 (2004)

140. M. Valko, C.J. Rhodes, J. Moncol, M. Izakovic and M. Mazur: Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem Biol Interact* 160, 1-40 (2006)

141. L.M. Coussens and Z. Werb: Inflammation and cancer. *Nature* 420, 860-867 (2002)

142. M.M. Mueller: Inflammation in epithelial skin tumors: old stories and new ideas. *Eur J Cancer* 42, 735-744 (2006)

143. M. Lechner, P. Lirk and J. Rieder: Inducible nitric oxide synthase (iNOS) in tumor biology: the two sides of the same coin. *Semin Cancer Biol* 15, 277-289 (2005)

144. S.H. Yuspa: Cutaneous chemical carcinogenesis. *J Am Acad Dermatol* 15, 1031-1044 (1986)

145. H. Ohshima and H. Bartsch: Chronic infections and inflammatory processes as cancer risk factors: possible role of nitric oxide in carcinogenesis. *Mutat Res* 305, 253-264 (1994)

146. T. Berhane, G.M. Halliday, B. Cooke and R.S. Barnetson: Inflammation is associated with progression of actinic keratoses to squamous cell carcinomas in humans. *Br J Dermatol* 146, 810-815 (2002)

147. H. Mukhtar, S.K. Katiyar and R. Agarwal: Green tea and skin--anticarcinogenic effects. *J Invest Dermatol* 102, 3-7 (1994)

148. M.S. Fisher and M.L. Kripke: Systemic alteration induced in mice by ultraviolet light irradiation and its

Redox processes in skin physiology and inflammation

relationship to ultraviolet carcinogenesis. *Proc Natl Acad Sci USA* 74, 1688-1692 (1977a)

149. D.L. Damian and G.M. Halliday: Measurement of ultraviolet radiation-induced suppression of recall contact and delayed-type hypersensitivity in humans. *Methods* 28, 34-45 (2002.)

150. S.E. Ullrich: Mechanisms underlying UV-induced immune suppression. *Mutat Res* 571, 185-205 (2005)

151. A.A. Vink, A.M. Moodycliffe, V. Shreedar, S.E. Ullrich, L. Roza, D.B. Yarosh and M.L. Kripke: The inhibition of antigen-presenting activity of dendritic cells resulting from UV irradiation of murine skin is restored by *in vitro* photorepair of cyclobutane pyrimidine dimers. *Proc Natl Acad Sci USA* 94, 5255-5260 (1997)

152. P. Wolf, H. Maier, R.R. Mullegger, C.A. Chadwick, R. Hofmann-Wellenhof, H.P. Soyer, A. Hofer, J. Smolle, M. Horn, L. Cerroni, D. Yarosh, J. Klein, C. Bucana, K. Dunner, C.S. Potten, H. Honigsmann, H. Kerl and M.L. Kripke: Topical treatment with liposomes containing T4 endonuclease V protects human skin *in vivo* from ultraviolet-induced upregulation of interleukin-10 and tumor necrosis factor- α . *J Invest Dermatol* 114, 149-156 (2000)

153. M. Norval and A.A. El-Ghorr: Studies to determine the immunomodulating effects of cis-urocanic acid. *Methods in Enzymology* 28, 63-70 (2002)

154. L.A. Barber, D.F. Spandau, S.C. Rathman, R.C. Murphy, C.A. Johnson, S.W. Kelley, S.A. Hurwitz and J.B. Travers: Expression of the platelet-activating factor receptor results in enhanced ultraviolet B radiation-induced apoptosis in a human epidermal cell line. *J Biol Chem* 273, 18891-18897 (1998)

155. J.P. Walterscheid, S.E. Ullrich and D.X. Nghiem: Platelet-activating factor, a molecular sensor for cellular damage, activates systemic immune suppression. *J Exp Med* 195, 171-179 (2002)

156. G.M. Halliday, P.A. Russo, K.S. Yuen and B.O. Robertson: Effect of inhibitors of oxygen radical and nitric oxide formation on UV radiation-induced erythema, immunosuppression and carcinogenesis. *Redox Rep* 4, 316-318 (1999)

157. K.E. Burke, J. Clive, G.F. Combs, J. Commisso, C.L. Keen and R.M. Nakamura: Effects of topical and oral vitamin E on pigmentation and skin cancer induced by ultraviolet irradiation in Skh:2 hairless mice. *Nutr Cancer* 38, 87-97 (2000)

158. H.L. Gensler, K.E. Gerrish, T. Williams, J. Rao and J. Kittelson: Prevention of photocarcinogenesis and UV-induced immunosuppression in mice by topical tannic acid. *Nutr Cancer* 22, 21-130 (1994)

159. H.L. Gensler, B.N. Timmermann, S. Valcic, G.A. Wachter, R. Dorr, K. Dvorakova and D.S. Alberts: Prevention of photocarcinogenesis by topical administration of pure epigallocatechin gallate isolated from green tea. *Nutr Cancer* 26, 325-335 (1996)

160. C. Caceres-Dittmar, K. Ariizumi, S. Xu, F.J. Tapia, P.R. Bergstresser and A. Takashima: Hydrogen peroxide mediates UV-induced impairment of antigen presentation in a murine epidermal-derived dendritic cell line. *Photochem Photobiol* 62, 176-83 (1995)

161. L.T. Van der Broeke and G.M. Beijersbergen van Henegouwen: Topically applied N-acetylcysteine as a protector against UVB-induced systemic immunosuppression. *J Photochem Photobiol B*, 27, 61-65 (1995)

162. T. Nakamura, S.R. Pinnell, D. Darr, I. Kurimoto, S. Itami, K. Yoshikawa and J.W. Streilein: Vitamin C abrogates the deleterious effects of UVB radiation on cutaneous immunity by a mechanism that does not depend on TNF- α . *J Invest Dermatol* 109, 20-24 (1997)

Abbreviation: AP1: activator protein 1, ARE: antioxidant response element, Ask-1: Apoptosis signal regulating kinase-1, CAT: catalase, cNOS: constitutive nitric oxide synthase, COX: cyclooxygenase, DNA: deoxyribonucleic acid, EGFR: epidermal growth factor receptor, eNOS: endothelial nitric oxide synthase, ERKs: extracellular signal-regulator kinases, GPx: glutathione peroxidase, GSF: , GSH: reduced form of glutathione, GSSG: oxidized form of glutathione, GST: glutathione-S-transferase, HO-1: heme oxygenase, , IL1: interleukin 1, IL8: interleukin 8, INF-gamma: interferon gamma, , iNOS: inducible nitric oxide synthase, JNK: c-jun N-terminal kinase, LPS: lipopolysaccharide, MAPKs: mitogen-activated protein kinases, MCP1: monocyte chemoattractant protein 1, MDA: malonyl dialdehyde, MMPs: matrix metalloproteinases, mRNA: messenger ribonucleic acid, NADPH-oxidase: nicotinamide adenine dinucleotide phosphate oxidase, NFkappaB: nuclear factor kappa B, nNOS: neuronal nitric oxide synthase, Nrf2: nuclear factor E2-related factor 2, PI3K: phosphoinositol-3-kinase, PKC: protein kinase C, PRx: peroxyredoxin, PTPs: protein-tyrosine phosphatases, PUVA: 5-methoxypsoralen + UVA, RNS: reactive nitrogen species, ROS: reactive oxygen species, SOD: superoxide dismutase, TGF- α : transforming growth factor, TLRs: toll-like receptors, TNF- α : tumor necrosis factor alpha, TPA: 12-O-tetradecanoylphorbol-13-acetate, TRX: thioredoxin, UGT: uridine diphosphate glucuronotransferase, UVA, UVB: ultraviolet A and B, respectively, VEGF: vascular endothelial growth factor,

Key Words: Skin Physiology, Skin Inflammation, Reactive Oxygen Species, Nitric Oxide, Antioxidants, Redox Balance, Psoriasis, Atopic Dermatitis, Contact Dermatitis, Skin Wounds, UV-Induced Inflammation, Review

Redox processes in skin physiology and inflammation

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