

Amino acids in healthy aging skeletal muscle

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1. ABSTRACT

Life expectancy in the U.S. and globally continues to increase. Despite increased life expectancy quality of life is not enhanced, and older adults often experience chronic age-related disease and functional disability, including frailty. Additionally, changes in body composition such as the involuntary loss of skeletal muscle mass (i.e. sarcopenia) and subsequent increases in adipose tissue can augment disease and disability in this population. Furthermore, increased oxidative stress and decreased antioxidant concentrations may also lead to metabolic dysfunction in older adults. Specific amino acids, including leucine, cysteine and its derivative taurine, and arginine can play various roles in healthy aging, especially in regards to skeletal muscle health. Leucine and arginine play important roles in muscle protein synthesis and cell growth while cysteine and arginine play important roles in quenching oxidative stress. Evidence suggests that supplemental doses of each of these amino acids may improve the aging phenotype. However, additional research is required to establish the doses required to achieve positive outcomes in humans.

2. INTRODUCTION

2.1. Aging and chronic disease

Life expectancy from birth and from the age of 60 years continues to increase with the average life expectancy in the United States being 78.8. years and 83.2. years for men and women, respectively, an approximate 16 year increase since 1940 (1). The percentage of the total population over the age of 65 years is now ~13% (40,229,000 older adults) and is expected to reach ~20% by 2030 (2).

Although human life expectancy is increasing, older individuals are faced with increased risk for chronic disease, reduced ability to perform activities of daily living, and general loss of independence, suggesting a larger window of diminished quality of life for aging adults. Many chronic diseases and physical impairments are associated with changes in body composition such as increases in fat mass and decreases in skeletal and bone masses. Age-related changes in skeletal muscle such as the involuntary loss of muscle mass (sarcopenia) and the associated decrease in strength increase the risk for falls and fractures. These changes have also been linked to loss of mobility and independence and to physical

Table 1. Classification of amino acids in humans

Essential amino acids	Conditionally essential amino acids	Nonessential amino acids
Histidine	Arginine	Alanine
Isoleucine	Cysteine	Aspartate
Leucine	Glutamine	Asparagine
Lysine	Glycine	Glutamate
Methionine	Proline	Serine
Phenylalanine	Tyrosine	
Threonine		
Tryptophan		
Valine		

disabilities in older adults (3-5). Such changes in body composition can lead to debilitating physical conditions that impact the ability of older adults to complete instrumental activities of daily living. In 2010, over 25% of those aged ≥ 65 years reported difficulty walking and climbing stairs, while 18.5% reported difficulty doing errands independently (6). Dietary modifications may improve adverse age-related changes in body composition, and therefore, may impact the development of many chronic conditions (e.g. sarcopenia, obesity). Current recommendations for improving body composition in all populations, but most importantly for older adults, include exercise and consumption of a healthy diet with adequate dietary protein (7, 8).

Approximately 35-45% of the adult human body mass is comprised of skeletal muscle; however, between the ages of 30-80 y, skeletal muscle mass decreases by approximately 30% (i.e. sarcopenia), declining at a rate of 3-8% per decade after the age of 30 y (9, 10). The aging skeletal muscle phenotype is characterized by visible adverse changes in composition and function that begin as early as the 4th decade of life (10). Interestingly, the relationship between age-related loss of skeletal muscle mass and strength is not linear (11). Goodpaster *et al.* reported a 3-4% decrease in leg lean mass but a 5-9% decrease in specific torque among older adults over a 3 y period (12). Changes in amino acid and protein metabolism in response to stimuli may contribute to these changes in skeletal muscle mass and strength in older adults. Evidence suggests that older adults have a blunted anabolic response to amino acid ingestion, although basal muscle protein degradation and turnover remains unchanged (13, 14). Additionally, age-related changes in the skeletal muscle transcriptome and methylome partially underlie and likely precede the observed phenotypic changes in aging muscle (15-18).

Age-related changes in skeletal muscle morphology and physiology accelerate with advancing

age, and are accentuated by intrinsic and extrinsic factors (19-26). For example, sarcopenia is characterized by chronic low-grade inflammation and is associated with increased reactive oxygen species production, altered mitochondrial function, and elevated apoptotic signaling (27, 28). Accumulation of reactive oxygen species damages lipid membranes, nuclear DNA, and mitochondrial DNA. Subsequently, these cellular damages decrease protein synthesis and induce apoptosis and myofiber death. Factors influencing muscular changes have been identified at all levels: the single myofiber, the skeletal muscle organ, and the whole organism. However, skeletal muscle aging is complex and multifactorial, and the molecular mechanisms instituting adverse changes in skeletal muscle with age are yet unclear.

2.2. Endogenous protein and amino acids

Endogenous proteins play many structural and functional roles in the mammalian body. All enzymes and many cellular structures including actin and myosin filaments, membrane carriers, and hormone receptors are proteins. Amino acids are the basic building blocks of all proteins and are often classified as essential, conditionally essential, or non-essential (Table 1), although essentiality is species and age dependent. In humans, nine amino acids are classified as essential. These amino acids have carbon skeletons that cannot be synthesized *de novo* by the body and are therefore required from dietary sources (29). Additionally, in humans, six amino acids are classified as conditionally essential. Conditionally essential amino acids can be synthesized by the human body, but their synthesis is limited by a variety of factors including amino acid precursor availability. When synthesis is limited for any reason, the amino acid becomes an essential part of the diet. Alternatively, five amino acids are known to be nonessential in humans. These amino acids are assumed to be synthesized in adequate amounts as long as the total protein requirement is met. However, whether or not these quantities are sufficient for optimal health and growth remains unknown. Furthermore, although the importance of conditionally essential and nonessential amino acid intake has been explored in other mammals, its research is limited in humans (30). Future research is needed to explore the role and dietary requirements of conditionally essential and nonessential amino acids in human health.

The skeletal muscle is a key organ system for the degradation and synthesis of amino acids. Degradation and synthesis of amino acids depends on both the distribution of macronutrients and the availability of macronutrients for energy production (i.e., fed vs. fasted). For example, skeletal muscle is the major organ for initiating the transamination of branched-chain amino acids (leucine, isoleucine, and valine). Branched chain ketoacids can then be released from muscle for oxidation in the liver and

Table 2. Amino acid dietary reference intakes

Amino acid	DRI Recommendation (mg · g protein ⁻¹ · day ⁻¹)	Grams amino acid per day based on 77.4. kg ¹	Grams amino acid per day based on 90 kg ²	References
Histidine	18	1.11	1.30	42, 62
Isoleucine	25	1.55	1.8	42, 62
Leucine	55	3.4	4.0	42, 62
Lysine	51	3.16	3.67	42, 62
Methionine & cysteine	25	1.5	1.8	42, 62
Phenylalanine & tyrosine	47	2.9	3.38	42, 62
Threonine	27	1.67	1.97	42, 62
Tryptophan	7	0.43	0.50	42, 62
Valine	32	1.98	2.3	42, 62

¹: Based on average weight of a U.S. woman 60-69 years old. ²: Based on average weight of a U.S. man 60-69 years old

other organs. The degradation of branched-chain amino acids provides products for the synthesis of amino acids such as glutamine in the muscle. Additionally, the release of amino acids (e.g., alanine and glutamine) from the skeletal muscle provides necessary products for synthesis of other amino acids and molecules, such as glucose, by other organs. While this is only a brief overview, the importance of skeletal muscle for amino acid metabolism is highlighted.

2.3. Exogenous protein and amino acids

Adequate dietary protein is essential for overall human health with recommendations differing throughout the human lifespan. As there are no true body stores for protein, insufficient protein intake to satisfy body requirements leads to a negative protein balance (i.e., protein synthesis less than breakdown). Imbalanced protein metabolism during inadequate protein intake generally occurs in the skeletal muscle, resulting in clinical manifestations such as skeletal muscle atrophy, impaired muscle growth or regrowth, and functional decline. Some populations, such as older adults, are particularly vulnerable to insufficient protein.

Animals and plants are both sources of exogenous protein and amino acids. Meat, poultry, seafood, dairy, and eggs are excellent sources of high biological value proteins, meaning the protein mixture in the individual food source contains each of the essential and conditionally essential amino acids at a level that equals or exceeds the human requirement for the amino acid as a percentage of the total protein requirement. Whey and casein are two common protein supplements derived from dairy products. When milk is curdled for cheese, it separates into two phases: the watery phase and the curds. Whey protein is derived from the watery phase. It represents about 20% of the proteins in cow's milk, and it is rich in leucine. Alternatively, casein is insoluble at acid pH and is found in the curds.

It represents about 80% of the proteins in cow's milk, but it has less leucine than whey. Legumes, nuts, seeds, vegetables, and whole grains are lower biological protein sources; they provide limited amounts of one or more of the essential amino acids.

The current protein Recommended Dietary Allowance (RDA) is 0.8 g · kg body weight⁻¹ · day⁻¹ for all adults over the age of 19 years. Infants, children, pregnant women, and lactating women have increased protein needs (31). A committee within the Institute of Medicine (the health arm of the United States National Academy of Science) set the adult protein RDA after analyzing several nitrogen balance studies. Furthermore, they also set requirements for many of the essential and conditionally essential amino acids after analyzing results of isotopic tracer studies (see Table 2 for Dietary Reference Intakes).

Adequate protein intake is essential for muscle mass maintenance and growth, especially in older adults. Several studies have shown that when compared to younger adults, older adults may have greater total protein needs and may benefit from higher protein intakes (32-39). In relatively healthy, older adults, acute studies demonstrated that old (vs. young) adult skeletal muscle has an impaired protein synthesis response to protein intakes of 20 g or less, but a comparable protein synthesis response to protein intakes of 25-30 g in a single meal (40). These varying responses could more specifically reflect the availability of specific amino acids (14, 41).

Additionally, two recent studies using the indicator amino acid oxidation method found that the mean protein requirement for elderly women was 0.96 g · kg⁻¹ · day⁻¹ or 0.85 g · kg⁻¹ · day⁻¹ (34, 35). Both of these values are above the current estimated average requirement of 0.66 g · kg⁻¹ · day⁻¹ and even

Table 3. Summary of the effects of single-dose or long-term protein or amino supplementation in older adults

Supplement	Dose and duration	Physiological effect	Conclusion
¹ EAA (52)	6.7 g EAAs (26% (1.7 g) or 41% (2.7 g) leucine); single dose	Following EAA ingestion containing 2.7 g leucine only: ↑ FSR, comparable to the younger adults consuming 2.7 g leucine ↑ Phenylalanine net balance (reflects muscle balance)	Increasing the proportion of leucine in a mixture of EAA can overcome the attenuated MPS response in elderly
Whey protein isolate (53)	0, 10, 20, or 40 g of whey protein isolate; single dose	Following the 20 g dose: ↑ MPS by 65% above basal level ↑ MPS by 90% over basal level No effects observed after ingestion of lower doses	Twenty grams of whey protein is sufficient to increase myofibrillar MPS in non-frail older adults
EAA+arginine (56)	11 g EAA+arginine; 2x per day, between meals for 16 weeks	↑ LBM after 12 weeks ↑ Lower extremity strength measure score ↑ Gait speed ↓ 5-step test time ↓ Floor-transfer test time	Supplementing the diet with EAA+arginine improves lean body mass, strength, and physical function compared to baseline values in glucose intolerant elderly individuals
EAA (57)	8 grams EAA (2.5 g leucine) given at 10 am and 5 pm for 12 months	↑ Whole-body lean mass ↑ Circulating IGF-1 ↓ Circulating TNF-alpha	Nutritional supplements with oral EAA mixture increased whole-body lean mass in elderly subjects with sarcopenia
EAA (58)	7.5 g EAA (1.39 g leucine); 2x per day for 3 months between meals	↑ Basal FSR ↑ Lean body mass ↑ Basal IGF-1 protein expression	EAA improved LBM and basal muscle protein synthesis in older individuals
Leucine (63)	4 g/meal; 3 meals/day; 2 weeks	↑ Postabsorptive FSR ↑ Phosphorylation of mTOR ↑ Phosphorylation of 4E-BP1 ↑ Phosphorylation of p70S6K1	Leucine supplementation during meals may improve MPS and mechanisms underlying anabolic responses in older skeletal muscle
Leucine (64)	2.5 g leucine consumed with meals for 3 months	No change in skeletal muscle mass or strength No change in whole-body insulin No change in glycated hemoglobin content No change in plasma lipids	Long-term leucine supplementation does not augment skeletal muscle mass or strength and does not improve glycemic control or blood lipid profile in healthy elderly men
Leucine (65)	2.5 g leucine consumed with meals for 6 months	No change in lean tissue mass (not different b/w groups) No change in body fat percentage No change in muscle strength No change in muscle fiber type No change in insulin sensitivity or plasma lipid concentration (did not change b/w groups)	Prolonged leucine supplementation does not modulate body composition, muscle strength, muscle mass, glycemic control, or lipidemia in elderly, type 2 diabetic patients who habitually consume adequate dietary protein

¹EAA: Essential amino acids; FSR: Fractional synthesis rate; MPS: Muscle protein synthesis; LBM: Lean body mass

above the current RDA of $0.8 \text{ g} \cdot \text{kg}^{-1} \cdot \text{day}^{-1}$ for adults. Of functional importance, Campbell *et al.* showed that older adults lost muscle mass when placed on a controlled diet with $0.8 \text{ g} \cdot \text{kg}^{-1} \cdot \text{day}^{-1}$ for 14 weeks (32). This study was tightly controlled, and each subject resided at the General Clinical Research Center at The Pennsylvania State University for the 14-week duration. The results of these studies collectively suggest that older adults have protein needs greater than the current RDA.

Consumption of other macronutrients is also important for muscular health. Protein and amino acid requirements are determined under conditions of sufficient dietary energy intake. Amino acids will be increasingly oxidized for energy under conditions of inadequate energy intake from carbohydrate and fat. Although glucose is the primary energy source for both the working muscle and the brain under postprandial conditions, amino acids can be used as a fuel, both directly and after

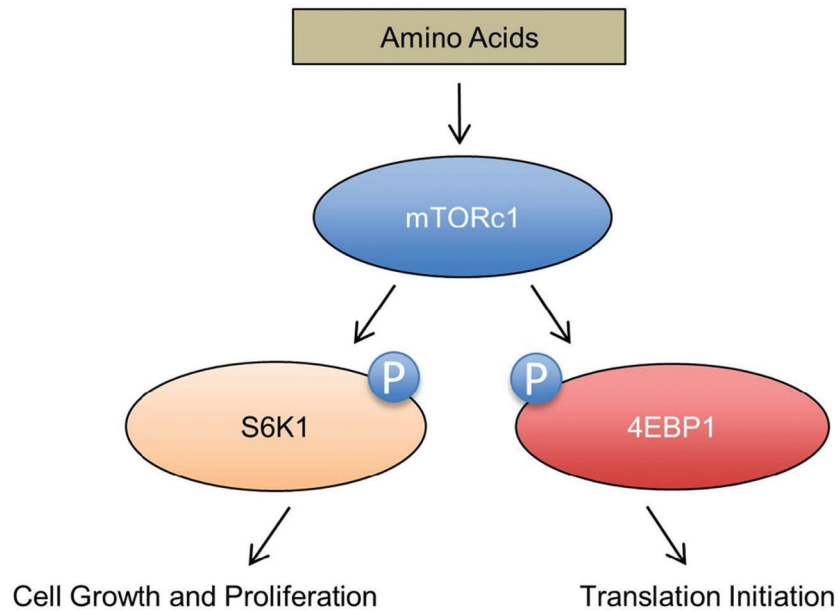


Figure 1. Biochemical mechanisms for the beneficial effects of BCAA supplementation on aging skeletal muscle.

gluconeogenesis from amino acid carbon chains, when energy intake is inadequate. Therefore, adequate energy intake, including some carbohydrate intake, is essential for sparing amino acids for anabolic purposes. With adequate energy and carbohydrate intake, protein can be utilized for its many structural and functional roles.

Specific amino acids are known to play integral roles in muscular health throughout the lifespan; these include leucine, cysteine and its derivative taurine, and arginine. Furthermore, aging affects the availability and physiologic effects of several of these amino acids. The purpose of this review is to summarize the roles of each of these amino acids in healthy aging, with a particular focus on skeletal muscle health.

3. LEUCINE

Leucine is a hydrophobic, branched-chain amino acid metabolized predominately in the liver, adipose tissue, and skeletal tissue (42). The leucine dietary reference intake is 55 mg · g dietary protein⁻¹ · day⁻¹ (~3 grams leucine for a 70 kg individual consuming the RDA for dietary protein (0.8 g · kg⁻¹ · day⁻¹) (43). Food sources include soybeans, beef, chicken, eggs, oats, nuts, and milk. Leucine is predominantly known for its role in the stimulation of muscle protein synthesis (44). However, leucine has several additional functions in the body. Unlike other amino acids, leucine cannot be converted into glucose. Rather, leucine is a significant precursor for lipid biosynthesis in adipose tissue and, to a lesser extent, in muscle and liver (42). Leucine, like other amino acids, is essential for the synthesis of proteins. In the brain, leucine is a component of leucine-enkephalin,

an endogenous opioid peptide neurotransmitter (45). Additionally, on the molecular level, leucine residues are critical components of the leucine zipper structural motif commonly found in DNA binding proteins. Each leucine zipper motif has four leucine residues, each separated by six amino acids (46).

3.1. Leucine-stimulated protein synthesis

The importance of essential amino acids, particularly leucine, in protein synthesis has been well established (47). Ingestion of essential amino acids, leucine in particular, increases mammalian target of rapamycin complex 1 (mTORC1) activity (47). mTORC1 is a serine/threonine protein kinase that increases muscle protein synthesis through translational control. Upon activation, mTORC1 phosphorylates many downstream targets including S6K1 and 4EBP1 (Figure 1). Phosphorylated S6K1 promotes cell growth and cell proliferation through phosphorylation of further downstream targets (48). Phosphorylation of 4EBP1, a translational repressor, results in its dissociation from the inactive eIF4E-4EBP1 complex which frees eIF4E to bind with eIF4G to form the active eIF4G-eIF4E complex (49). The eIF4G-eIF4E complex plays an integral role in eukaryotic translation initiation (50).

Even when isolated from other essential amino acids, leucine can independently activate mTORC1 (51) and enhance muscle protein synthesis (52). Multiple studies have shown that essential amino acid bolus feedings that include 3 grams of leucine significantly increase muscle protein fractional synthetic rate in older adults (53-55). In young adults, 10 grams of essential amino acids (~2 grams of leucine) mixed in a

noncaloric, noncaffeinated carbonated beverage was sufficient to induce maximal postprandial muscle protein synthesis (56). Other human studies have shown that essential amino acid supplements with 1.39 to 3.95 grams of leucine given between meals increase lean body mass, strength, and physical function independently of exercise (57-59).

3.2. Anabolic resistance to protein and leucine ingestion in aging

Aging is associated with a blunted skeletal muscle anabolic response to ingestion of 20 grams or less of high quality protein (14, 53). Additionally, when normalized to body weight, older men require a greater relative protein intake than younger men to reach equivalent muscle protein synthesis rates suggesting the existence of an anabolic threshold (60). However, the anabolic response to the ingestion of 30 grams of protein is similar in old and young skeletal muscle (61). This bolus-dependent anabolic response may be clinically relevant; merely supplementing the diet of older adults with 20 grams of protein twice per day was not sufficient to attenuate skeletal muscle loss with disuse (62). Rather, current evidence suggests that older adults may benefit more from an even meal distribution of protein with 25-30 grams of protein at each of the three daily meals, ~90 grams per day (40). Importantly, 30 grams of high quality protein contains ~3 grams of leucine. Therefore, the amount of leucine recommended to overcome the impaired anabolic response in older adults is ~3 grams per meal, or ~9 grams per day, a level that is 3 times the current leucine RDA (Table 3) (43, 63).

Supplementing older adults whose protein intake was controlled at $0.8 \text{ g} \cdot \text{kg}^{-1} \cdot \text{day}^{-1}$ with additional leucine at meals increased fractional synthetic rate and mTORC1 signaling (64). However, when older adults who were already consuming $\sim 1.0 \text{ g protein} \cdot \text{kg}^{-1} \cdot \text{day}^{-1}$ were supplemented with leucine at meals, no effects on changes to fractional synthetic rate or lean body mass were observed (65, 66). Collectively, this evidence suggests that essential amino acid supplements given between meals or leucine supplements given with meals may only be beneficial for muscle mass maintenance in those consuming less than or equal to the current protein RDA.

Despite the increasing evidence that older adults may have increased dietary protein and/or leucine needs, protein intake, and therefore leucine intake, generally decreases with age (67, 68). This decrease in protein intake may be attributed to changes in appetite, especially in those consuming liquid supplements (68, 69). Essential amino acid or leucine supplements may be an effective strategy for increasing protein intake, muscle protein synthesis, and lean body mass in older adults without adversely affecting satiety or the normal metabolic response to a later meal (70, 71).

3.3. Inactivity and bedrest

Inactivity and bedrest further attenuate the anabolic response to protein/leucine intake (72). In healthy, older adults seven days of bedrest significantly reduces leg lean mass and muscle protein synthesis in response to 12 grams of essential amino acids. Seven days of bedrest is comparable to the average length of a hospital stay for older adults with acute illness (73). Older adults also lose significantly more leg mass after just 10 days of bedrest compared to the amount younger adults lose after 28 days of bedrest (74, 75). These large reductions in muscle mass with inactivity in older adults may result from blunted muscle protein synthesis in response to essential amino acid stimulation. Drummond *et al.* reported that the reduction in leg lean mass and muscle protein synthesis after bed rest is accompanied by a reduction in mTORC1 signaling and a reduction in amino acid transporter protein content (72). Augmentation of anabolic resistance following muscle disuse could explain why protein supplementation alone does not attenuate disuse atrophy. Interestingly, exercise training alone is inadequate to fully regain the muscle mass lost during two weeks of bedrest (76, 77). Therefore, nutritional interventions that stimulate mTORC1 activity, a positive regulator of muscle protein synthesis, may have important implications for skeletal health in older adults, especially when combined with exercise training. Research investigating the therapeutic effects of combined physical activity and protein/leucine supplementation post bedrest or immobilization is needed.

3.4. Leucine and weight loss in aging

Leucine may also play an important role in muscle mass maintenance during weight loss, especially in older adults (78). Along with the age-related decrease of skeletal muscle mass there is an accumulation of fat within and around the muscle (sarcopenic obesity) which, combined with skeletal muscle loss, likely increases one's risk for functional disability and metabolic dysregulation (79, 80). Additionally, there is an age-associated redistribution of fat mass to the abdominal region which is closely linked to morbidity and mortality as well as increased risk of chronic diseases (e.g., cardiovascular disease, insulin resistance, metabolic syndrome, etc.) (81-83). Further, the global rise in obesity is affecting all generations and is a health concern for the older population (84). A major problem that older adults face when trying to combat obesity with caloric restriction is the further loss of skeletal muscle mass; in some older adults, the risks associated with muscle atrophy may outweigh the benefits of weight loss. Sarcopenia itself is independently associated with insulin resistance (79). Therefore, utilizing nutritional strategies that promote muscle mass maintenance is imperative during weight loss, especially in older adults.

Although the ideal macronutrient ratios during weight loss remain controversial, higher protein diets

are likely to promote better body composition during hypocaloric conditions (8, 85-88). Furthermore, because of leucine's unique role in muscle protein synthesis, leucine has the potential to independently stimulate muscle protein synthesis and protect lean body mass during weight loss (89-91). Several studies have reported benefits of whey protein or dairy consumption on adipose tissue loss, lean muscle mass maintenance, and muscle protein synthesis rates during weight loss (78, 92, 93). Preserving muscle mass during weight loss is important for the prevention of sarcopenia as well as for improvements in metabolic function and insulin sensitivity.

3.5. Leucine and insulin resistance

Amino acids, especially the essential amino acids, increase circulating insulin levels, and high protein intakes have been associated with the development of type II diabetes (94-96). Alternatively, higher protein diets have also been associated with improved body composition through greater loss of fat mass and reduced loss of lean muscle mass (85, 97). Improvements in glycemic control often accompany these improvements in body composition (85, 97).

Skeletal muscle plays an important role in insulin sensitivity; it accounts for ~75% of the body's insulin-stimulated glucose uptake (98, 99). Despite the positive data surrounding leucine's beneficial effect on muscle mass maintenance and sarcopenia prevention in older adults, recent epidemiologic data suggests an association between elevated circulating branched-chain amino acids and the development of type II diabetes (100-102). However, evidence from animal feeding studies suggests that the association of branched-chain amino acids with type II diabetes development may only occur in the background of a high-fat diet or in humans who preferentially use fat for energy production (100, 103, 104). Similarly, elevated leucine levels observed in obese individuals decline rapidly following bariatric surgery (105). Interestingly, while excess leucine from dietary protein intake may increase the risk of metabolic disease in adults 50-65 y, higher protein intake may be beneficial at reducing overall and cancer-related mortality in adults over the age of 66 y (106). Whether leucine is the cause or effect of insulin resistance is unclear, but further investigation is warranted, particularly if leucine is protective against mortality in older adults.

Inflammation is known to play a key role in obesity-linked insulin resistance (107). Burrill *et al.* recently reported that proinflammatory cytokine treatments down-regulated the expression of branched-chain amino acid transport and oxidation genes in 3T3-L1 cells (108). Furthermore, in rat liver epithelial cells, insulin inhibits branched-chain α -ketoacid dehydrogenase, the rate-limiting enzyme in branched-chain amino acid catabolism (109). The cumulative

effects of chronic inflammation and hyperinsulinemia may induce gene expression changes that result in decreased branched-chain amino acid uptake in critical tissues. Future research needs to continue elucidating the mechanistic links between branched-chain amino acids and insulin resistance; no causal relationships have been established to date.

4. CYSTEINE

Cysteine is a semi-essential amino acid that may be consumed as such in the diet, or it can be synthesized in the body from the sulfur atom from methionine and the amino acid serine. Because of this, the requirements for methionine and cysteine must be considered collectively. The dietary reference intake for cysteine and methionine is $25 \text{ mg} \cdot \text{g protein}^{-1} \cdot \text{day}^{-1}$ (43). Food sources of cysteine include soy, eggs, dairy, whole grains, meats, and poultry.

Cysteine is a precursor to glutathione, the most abundant intracellular thiol, as well as iron-sulfur proteins, such as NADH dehydrogenase (110). Cysteine residues are often covalently bound to other cysteine residues in disulfide bonds. These disulfide bonds play important roles in crosslinking proteins and supporting a protein's tertiary structure. Although clinically diagnosed cysteine deficiency is rare, evidence suggests that supplemental cysteine may be beneficial, especially in older adults (111).

4.1. Cysteine and glutathione synthesis

Cysteine is required for both synthesis of proteins and synthesis of the tripeptide gamma-glutamylcysteinylglycine, or glutathione. Glutathione is the major intracellular thiol and an important antioxidant molecule, and the availability of cysteine limits the rate of glutathione synthesis. The normal turnover of glutathione in adults has been estimated to be $< 40 \text{ mmol per day}$, which is slightly greater than estimates of the magnitude of cysteine turnover in the body protein pool (112-115). Because tissue glutathione levels become depleted at sulfur amino acid intakes that are marginal but adequate for protein synthesis, marginal protein intakes are likely to be associated with low tissue glutathione levels (116-119).

Sulfur amino acid nutrition is of significant interest in the context of aging because it is known that oxidative stress increases as organisms age, and oxidative stress may contribute to the sarcopenia associated with aging (120, 121). Furthermore, human plasma and erythrocyte glutathione levels decrease in aging, and animal studies have shown that muscle, blood cell, liver, and lung glutathione concentrations decrease with age (122-127). Because cysteine is the limiting substrate for glutathione synthesis, decreased cysteine availability may contribute to lower glutathione levels in older adults.

Several studies have shown associations of glutathione depletion with increased oxidative stress and increased muscle cell apoptosis. Dam *et al.* reported that depletion of glutathione increased reactive oxygen species production, DNA fragmentation, calpain activity, and caspase-independent apoptotic signaling in rat skeletal muscle (128). Sekhar *et al.* reported that, compared to younger adults, older adults had lower cysteine and glutathione levels in red blood cells, lower fractional rates of glutathione synthesis, and higher levels of three markers of oxidative stress (i.e., hydroperoxides, F_2 -isoprostanes, and lipid peroxides) (127). Sinha-Hikim *et al.* observed elevated markers of oxidative stress, inflammation, and muscle cell apoptosis, as well as decreased muscle weight, in aged mice compared with young mice (129).

Short-term cysteine supplementation studies in humans have shown positive effects on glutathione pools (127, 130, 131). Sekhar *et al.* reported that supplementing older subjects with *N*-acetylcysteine equivalent to $0.81 \text{ mmol cysteine} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$ plus $1.33 \text{ mmol} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$ glycine for 14 days increased the rate of intracellular glutathione synthesis and nearly doubled red blood cell glutathione concentrations, resulting in glutathione concentrations in supplemented older adults that were comparable to the levels observed in young subjects (127). Supplementation also led to a significant reduction in concentrations of plasma markers of oxidative stress (reactive oxygen metabolites, F_2 -isoprostane, and lipid peroxides) and in the ratio of oxidized glutathione to reduced glutathione in red blood cells. Sekhar *et al.* also reported similar positive outcomes of the *N*-acetylcysteine + glycine supplementation of patients with uncontrolled type 2 diabetes (131). In addition, Nguyen *et al.* showed that supplementation of older HIV-infected men with this same *N*-acetylcysteine + glycine supplement for 14 days increased red blood cell glutathione level, reduced markers of oxidative stress, increased muscle strength, and improved mitochondrial function and insulin sensitivity (132).

Sinha-Hikim *et al.* and Vidal *et al.* conducted studies with longer term cysteine supplementation in rodents (129, 133). Sinha-Hikim *et al.* found that supplementation of aged male mice with a formulation containing cysteine, glycine, selenomethionine and glutamine for 6 months prevented age-related increases in muscle cell apoptosis (129). Supplementation also prevented increases in oxidative stress and inflammation marker levels; tissue glutathione levels were not reported for this study but presumably were increased by the sulfur amino acid-containing supplement. Vidal *et al.* reported the physiologic effects of long-term cysteine supplementation in rats (133). Twenty-one month old rats were clustered into one of two groups, non-inflamed or low-grade inflamed, according to their natural α_2 -macroglobulin plasma level. Rats with an α_2 -macroglobulin level

greater than two standard deviations above the mean for adult rats were categorized as low-grade inflamed. None of the rats showed any observable signs of disease at baseline. After rats were categorized, they were fed either a nonpurified rodent diet supplemented with $4.0 \text{ g} \cdot \text{kg}^{-1}$ of cysteine or the same diet supplemented with an isonitrogenous amount of alanine (control) for 14 weeks. Cysteine-supplemented rats had higher free cysteine and free glutathione concentrations in the liver than did the alanine-supplemented groups, regardless of inflammation grade. There was no difference in acute-phase protein levels in plasma of cysteine-supplemented and control rats. Interestingly, however, cysteine-supplemented rats did not demonstrate the age-related decline in food intake that was observed in the control rats supplemented with alanine. This association of cysteine and glutathione status with food intake is consistent with previous findings; Hernadfalvi *et al.* reported that the attenuation of anorexia in lipopolysaccharide-pretreated mice was associated with increased glutathione levels in brain and liver (134). No causation between brain glutathione levels and food intake has been proven. However, the effect of tissue glutathione levels on food intake should be further investigated given that decreased food intake has been negatively associated with survival in elderly patients (135).

Sulfur amino acids are present in adequate amounts in most diets that include high-protein foods. However, when dietary sulfur amino acid intake is inadequate, cysteine supplements may be beneficial. Cysteine-rich, un-denatured whey protein can be used as a cysteine supplement. Daily whey protein consumption (2-3 servings of 10-15 grams) has been shown to increase plasma glutathione levels in patients with nonalcoholic steatohepatitis, HIV, and cystic fibrosis (136-138). Because whey protein is also a good source of leucine, whey protein supplements may confer multiple health benefits in older adults. In clinical and research situations, precursors of cysteine are frequently used because cysteine and its precursor methionine are both easily oxidized. As a supplement to research diets, the disulfide form of cysteine, cystine, can be used; much of the cysteine present in dietary proteins is present in disulfide linked form and is released as cystine during proteolytic digestion (129, 139). Clinically, cysteine derivatives, especially *N*-acetylcysteine and 2-oxothiazolidine-4-carboxylate have been used to provide cysteine, especially when intravenous administration is required, because these derivatives are much more water-soluble than the disulfide cysteine (140, 141). Supplementation with cysteine or its various derivatives are not entirely equal because of differences in the uptake and metabolism of cysteine and its derivatives, but cysteine and glutathione depletion or inadequacy favor the utilization of all forms of cysteine for glutathione synthesis as opposed to their catabolism to taurine and inorganic sulfur (142-145).

Overall, these results suggest that increased cysteine intake may be beneficial for older adults in that they may prevent or minimize decreases in tissue glutathione levels and increases in oxidative stress that occur with aging. However, the etiology of decreased cysteine availability in older adults, the effects of restored glutathione levels on inflammatory disease progression in humans, and the long-term effects of cysteine supplementation on clinical outcomes or disease attenuation have received little investigation. The potential therapeutic effects of restored glutathione levels on sarcopenia, chronic disease progression, and mortality remain largely unknown.

4.2. Taurine

Taurine, a derivative of cysteine, has antioxidant, anti-inflammatory, and metabolic actions that may be therapeutic for aging individuals. Muscle cannot synthesize taurine from cysteine; therefore, it is dependent upon taurine uptake. Taurine is found naturally in fish and meat but is absent in plants outside of algae. Although taurine is an acid with an amino group, taurine has a sulfonic acid group instead of a carboxylic acid group like the proteogenic amino acids; taurine is not used as a building block for proteins in the body. However, it is the most abundant free amino acid in skeletal and cardiac tissue, and it plays a role in many cellular functions including muscle excitability, oxidant attenuation, and inflammatory control (146, 147). Over the past decade, taurine's direct and indirect properties have been recognized as important attenuators of skeletal muscle health, cardiovascular disease, and diabetes.

Taurine plays an integral role in chloride ion conductance and calcium homeostasis – two important regulators of muscle excitability. Several studies have shown that changing the taurine concentration within muscle leads to changes in muscle function including reduced force production, action potential speed, and exercise capacity (148, 149). Ito *et al.* recently showed that taurine depletion, via a taurine transporter knockout, accelerates aging-associated skeletal muscle tissue damage in mice (150). Knockout mice showed significant skeletal muscle defects, elevated aging biomarker mRNA levels, and increased activation of the unfolded protein response (150). These mice also had shorter lifespans, decreased mitochondrial complex 1 activity, and centralized nuclei distribution in muscle cells, a histological change indicative of muscle injury (150). Together these defects indicate severe muscle injury in taurine-deficient mice.

Aged muscle develops a phenotype with many of the same functional and histological characteristics as younger taurine-depleted muscles. An age-associated reduction in taurine content has been reported in serum and tissues, including skeletal muscle (151-153).

Because many of the defects observed in muscles of aged animals are also observed in young taurine-depleted animals, several studies have evaluated the effect of taurine supplementation on taurine restoration and muscle function in aged animals. Treating aged rats with 2% taurine in their drinking water (water was administered until rats drank a daily dose of 1g taurine · kg⁻¹ bodyweight) for 3 months significantly increased muscle taurine content and resting chloride conductance (152). Similarly, feeding aged rats normal chow plus water supplemented with 0.12 mol · L⁻¹ of taurine for 8 months increased serum and tissue taurine concentrations to the levels found in younger rats and reduced age-related markers of oxidative damage (154).

As previously noted, bedrest further exacerbates the aging muscle phenotype. Several studies suggest that taurine supplementation may help alleviate the harmful physiologic effects of bedrest. Supplementing rats with 5 g taurine · kg⁻¹ bodyweight after 14 days of hindlimb unloading (which induces muscle atrophy in rats) restored cytosolic calcium concentration and resting chloride conductance (155). However, although taurine supplementation helped restore muscle function, it did not prevent muscle atrophy in this model (155). Alternatively, pretreatment of C2C12 mouse myotubes with taurine counteracted muscle atrophy and restored mitochondrial function when the myotubes were exposed to cisplatin, a pharmacological inducer of muscle weakness and cachexia (156).

Understanding the effect of taurine on muscle atrophy is also important for cardiovascular health interventions. Human cross-sectional and case control studies suggest that increased taurine is negatively associated with cardiovascular disease (157, 158). Taurine transporter knockout mice show that taurine deficiency alters the structure of ventricular cardiomyocytes, induces cardiomyopathy, and increases expression of heart failure genes (159, 160). Although the role of taurine in cardiac and skeletal muscle atrophy remains controversial, evidence suggests that taurine plays a role in preventing age related muscle deterioration in animal and cell models. Furthermore, supplementing with taurine may alleviate the age-associated decline in taurine content and restore several muscle functions.

Taurine's antioxidant and anti-inflammatory properties may also have beneficial roles in the treatment of type II diabetes. Taurine supplementation may improve diabetic outcomes through alleviating the negative effects of reactive oxygen species on insulin release and resistance. In animal models, taurine supplements counteract oxidative stress, improve insulin sensitivity by inhibiting c-Jun N-terminal kinase activation, and improve beta-cell stimulus-secretion coupling (161, 162). Similarly, in overweight, non-diabetic male patients with heparin and intravenous intralipid induced insulin resistance,

taurine supplementation ($3 \text{ g} \cdot \text{day}^{-1}$ for 2 weeks) improved insulin sensitivity and beta cell function (163).

Collectively, an abundance of experimental evidence suggests that taurine plays a role in the prevention and/or treatment of many chronic conditions. These conditions include, but are not limited to, age-related skeletal muscle decline, cardiovascular disease, and type II diabetes. However, before recommendations for nutritional therapy can be established, large-scale clinical trials are needed to confirm the findings from animal models and cross-sectional and case control observations.

5. ARGININE

Arginine is a conditionally essential amino acid that plays a role in cell signaling and insulin and growth hormone release (164, 165). An arginine dietary reference intake value has not been established. However, food sources include soy products, seeds, crustaceans, meats, and poultry. Within the body, arginine's biosynthesis depends on the availability of its carbon and nitrogen precursors: glutamate, glutamine, and proline (166). Available arginine can be converted to L-ornithine, an important intermediate in the urea cycle, or to phosphocreatine, an important source of readily available and quickly used ATP within skeletal muscle cells (167, 168). Additionally, arginine is a precursor for nitric oxide, a neurotransmitter, mediator of the immune response, and potent vasodilator. Functionally, arginine plays important roles in cardiovascular function, blood pressure control, and wound healing (169, 170).

5.1. Arginine, growth hormone, and insulin-like growth factor 1

Insulin and growth hormone both play important roles in cell growth and other anabolic processes within the body. Growth hormone also stimulates the production of insulin like growth factor-1 (IGF-1), a hormone involved in muscle mass maintenance and accretion (171). Aging is associated with decreased circulating levels of growth hormone and IGF-1 (22). Growth hormone therapy has shown positive effects on body composition, strength, and circulating IGF-1 levels in adults with confirmed growth hormone deficiencies (172, 173). However, growth hormone interventions have a high incidence of adverse effects including fluid retention, joint pain, and muscle pain (174). These side effects may further limit mobility in older adults. Additionally, growth hormone interventions are expensive – another important limitation for their use in the elderly (175). Therefore, alternative therapies, such as arginine supplementation, that induce growth hormone release and promote muscle mass accretion are being investigated.

Recent evidence suggests that arginine positively affects skeletal muscle growth through actions

dependent and independent of the growth hormone axis in animal and cell culture models (176-180). He *et al.* found that 120-day-old pigs supplemented for 46 days with 1.0 % arginine in their corn and soybean meal-based diet had significantly greater skeletal muscle mass and lower fat mass (176). Metabolomic analysis of serum from treated pigs showed that dietary arginine enhanced skeletal muscle protein synthesis (176). Creatinine, lysine, tyrosine, and tricarboxylic acid cycle metabolites were all higher in arginine-supplemented pigs. Although growth hormone and insulin levels were not evaluated in this study, they were measured in a similar study conducted by the same group (177). In this latter study, 60 days of arginine supplementation did not significantly alter growth hormone or insulin-like growth factor-1 levels measured 12 hours after feeding. Similarly, mice supplemented with 1.51% L-arginine-HCl in their drinking water increased muscle weight by 12% although no increase in growth hormone was observed 5 hours after feeding (178).

In 2005, Collier *et al.* first reported the oral doses of arginine required to elicit the growth hormone response (181). Compared to placebo, both 5 and 9 grams of ingested arginine induced a transient increase in circulating growth hormone concentrations that were apparent by 30 minutes post-ingestion with peak levels around 60 minutes post-ingestion. Intriguingly, when compared to placebo, 13 grams of arginine did not result in a significant growth hormone response. Most subjects reported gastrointestinal distress after the 13 gram dose; therefore, much of the arginine may have been excreted before it was absorbed. This work produced a new understanding of the time dependency of the growth hormone response following arginine ingestion. Previously, this time-dependent response was not always considered when evaluating arginine's effect on the growth hormone response. Therefore, the effect of arginine on the growth hormone response may have been missed by sampling at an inappropriate time.

Collectively, these results suggest that arginine or dietary protein sources rich in arginine can improve skeletal muscle health in animals. However, whether similar effects occur in humans is unknown but is an important area for future investigation.

5.2. Arginine and muscle protein synthesis

Aside from the effect of arginine on the growth hormone axis, arginine supplementation may also affect other signaling cascades, including the mTORC1 pathway. In 2008, Yao *et al.* first analyzed the effect of dietary arginine supplementation on the mTORC1 pathway in neonatal pigs (179). Treated pigs were supplemented with 0.6 % L-arginine in a milk-based diet for 7 days. Control pigs were fed the same diet supplemented with an isonitrogenous amount of alanine. After their last meal, the abundance and

phosphorylation state of mTORC1, S6K1, and several eukaryotic initiation factors were assessed in the skeletal muscle. Although arginine did not alter phosphorylated levels of S6K1, it did increase levels of phosphorylated mTORC1 and 4EBP1. Protein fractional synthetic rate and formation of the active eIF4G-eIF4E complex also increased in the skeletal muscle. It is important to note, however, that arginine supplementation also increased levels of circulating insulin, an independent stimulator of mTORC1. Other growth factors, such as growth hormone, were not measured. Collectively, these results suggest that arginine supplementation potentiates mTORC1 signaling compared to controls fed a milk-based diet only. However, it was not clear whether these results occurred independently of growth factor release.

Ham *et al.* investigated the direct role of arginine on skeletal muscle protein synthesis in growth factor and nutrient deprived C2C12 cells (180). C2C12 myotubes were incubated in HEPES buffered saline with arginine or equimolar concentrations of alanine for up to 4 hours. Compared to alanine, arginine supplementation increased protein synthesis, myotube diameter, and phosphorylation of mTORC1 and its downstream targets rS6 (a phosphorylation target of S6K1) and 4EBP1 during the first hour of treatment (180). Furthermore, experiments using identical methodology with the inclusion of rapamycin, an inhibitor of mTORC1, did not show any effect of arginine on protein synthesis or myotube diameter (180). This was the first study to show the direct effect of arginine in protection against muscle wasting during catabolic conditions *in vitro*. Because cells were deprived of growth factors and nutrients, the results of this study support a direct role for arginine in the regulation of mTORC1 in skeletal muscle. Collectively, these results suggest that arginine positively influences cell growth and muscle protein synthesis through both growth factor-independent and growth factor-dependent stimulation.

5.3. Arginine in aging

Both cell culture and animal model experiments report positive effects of arginine on muscle protein synthesis. However, limited evidence exists on the role of arginine alone on muscle mass maintenance in aging adults. Studies have shown that a mixture of β -hydroxy- β -methylbutyrate (HMB, a metabolite from leucine catabolism), arginine, and glutamine increases fat free mass in adults with various wasting diseases, including AIDS and cancer-related cachexia (182, 183). Additionally, supplementing elderly women with a combination of 2 g HBM, 5 g arginine, and 1.5 g lysine for 12 weeks improved protein synthesis, strength, and functionality (184). Although these studies support the beneficial effects of arginine mixed with HMB, lysine, and/or glutamine, there is limited evidence for the long-term effects of arginine alone on the age-related decline in skeletal muscle mass and/or function.

Arginine has also been investigated for its use in the treatment of coronary endothelial dysfunction. Arginine is the precursor of nitric oxide, the endothelium-derived vasodilator. Experimental studies have shown that oral L-arginine supplementation improves coronary endothelial function in patients with coronary artery disease, hypercholesterolemia, and hypertension (185-187). Bode-Boger *et al.* also reported the positive effects of L-arginine supplementation in healthy older adults with age-related progressive endothelial dysfunction (188). Although several reports and initial clinical trials show the beneficial effects of arginine on endothelial function, the direct mechanism remains unclear. In the normal physiologic state, intracellular concentrations of arginine far exceed those needed by endothelial nitric oxide synthase (NOS), the enzyme that converts arginine into nitric oxide. The effect of supplemental arginine on nitric oxide production may be explained, in part, by the enzyme arginase. Like NOS, arginase also requires arginine as a substrate, and it is upregulated in several disease conditions including hypercholesterolemia and hypertension (189, 190). Increased arginase activity and expression may decrease the amount of arginine available for nitric oxide synthesis (191). When additional arginine is supplemented to those with upregulated arginase, arginine's availability for nitric oxide production may be restored.

6. EFFECTS OF EXCESS AMINO ACID INTAKE

There is no evidence that amino acids derived from usual or even high intakes of protein from food present any risk. For individuals consuming typical foods, intake of protein is unlikely to exceed 25% of energy, and these levels are well within safe intakes of protein. However, it does not follow that a high intake of a purified protein supplement or of an individual L-amino acid is safe. At this time, no tolerable upper intake levels have been established by the Food and Nutrition Board of the Institute of Medicine; a detailed summary of the available data related to risk of excess intakes of individual L-amino acids is included in their publication "Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acids, Cholesterol, Protein, and Amino Acids (31)."

The majority of the research on adverse effects of excess intakes of individual L-amino acids has been conducted in animals, with reductions in feed intake and growth rate being the most apparent outcomes. Excess amino acids are most toxic when they are given in amounts that are disproportionate to the normal amino acid composition of the diet (192-194). Consistent with this association of adverse effects with the degree of imbalance of the overall amino acid pattern, Imamura *et al.* found adverse effects of 3% (w/w) L-leucine supplementation on growth of rats fed a low protein diet (6% casein), whereas there was no effect of up

to 8% L-leucine supplementation on growth of rats fed a moderate (12% casein) or high (40% casein) diet (195). This relationship of adversity to degree of imbalance can be seen as related to the degree to which protein synthesis can serve as a sink for the excess amino acid and the degree of competition among amino acids for shared transporters or shared metabolic enzymes. In addition to effects on feed intake and growth rate, excess intake of individual amino acids by animals have been reported to have a variety of effects, including alterations in hormone levels, one-carbon metabolism, mineral bioavailability and blood lipid profiles.

There is little dose-response data for the effects of excess amino acid intake, either acute or chronic, by humans. Pencharz *et al.* studied the effect of acute ingestion of excess L-leucine by young men on leucine catabolism and a panel of markers for adverse effects (196). They estimated that the metabolic limit for leucine oxidation was between 550 and 700 mg. kg⁻¹.d⁻¹ (~39 g/d), a level that was associated with increased plasma leucine and ammonia concentrations. Hiratsuka *et al.* studied the effect of intake of various doses of L-tryptophan (1 to 5 g/d) for 3-week periods in young women and found no adverse effects (197). The urinary excretion of nicotinamide and other tryptophan catabolites increased in proportion to the ingested amounts of tryptophan, demonstrating that the metabolic capacity was not saturated. In rare cases, excess intake of tryptophan combined with use of serotonin drugs has resulted in "serotonin syndrome" due to toxic effects of excess serotonin on the nervous system.

Although almost all studies suggest that supplementation with an L-amino acid at several times its requirement level is not associated with adverse effects in humans, little data is available that allows evaluation of possible adverse effects associated with long-term intake of excess L-amino acid. Those using or promoting the use of supplemental amino acids should also be aware of potential hazards related to use of any supplement, in particular errors in dosage calculation and the possible presence of contaminants. Two examples serve to underscore the importance of these potential, though rare, risks of supplement use. Although oral doses of L-methionine (100 mg/kg body wt) are widely used to test for a tendency to develop hyperhomocysteinemia and have been regarded as very safe, one subject died after a methionine loading test in which the subject is believed to have been accidentally given 10-times the specified amount (198). A condition called eosinophilia myalgia syndrome appeared in 1989 that was linked to supplemental L-tryptophan use and subsequently shown to be due to a contaminant in certain production batches (199). Future research and/or clinical case studies of observed effects are necessary to determine adverse effects of amino acid supplementation.

7. CONCLUSION AND FUTURE DIRECTION

As evidenced in this review, amino acids play a role in healthy aging. Knowledge of the interactions between specific amino acids and age-related physiological and metabolic impairment has advanced through discovery research. Research using animal and cell culture models has led to a better understanding of amino acid signaling cascades and potential impacts of amino acids on health outcomes. However, there are several important differences between animal and cell culture models and humans. Of the amino acids presented in this paper, only leucine has been well-studied in humans. Human data support increased protein intakes in older adults and bolus doses of leucine and/or essential amino acids between meals for skeletal muscle health. Although discovery research has shown positive effects for cysteine, taurine, and arginine in mammalian physiology, the effects of supplemental doses of each of these amino acids in humans remains unclear. Future research to characterize the role of these and other amino acids (not discussed) in healthy human aging is also necessary.

8. REFERENCES

1. C. f. D. Control: Deaths: Final Data for 2011, Table 7. In: Ed M. M. C. M.-d. Files. (2011)
2. G. K. a. V. A. V. Vincent: THE NEXT FOUR DECADES, The Older Population in the United States 2010 to 2050. In: *Current Population Reports*. U.S. Census Bureau, Washington, DC. (2010)
3. C. W. Bales and C. S. Ritchie: Sarcopenia, weight loss, and nutritional frailty in the elderly. *Annu Rev Nutr*, 22, 309-23 (2002)
DOI: 10.1146/annurev.nutr.22.010402.102715
4. T. J. Doherty: Invited review: Aging and sarcopenia. *J Appl Physiol*, 95(4), 1717-27 (2003)
DOI: 10.1152/jappphysiol.00347.2003
5. T. Rantanen, J. M. Guralnik, R. Sakari-Rantala, S. Leveille, E. M. Simonsick, S. Ling and L. P. Fried: Disability, physical activity, and muscle strength in older women: the Women's Health and Aging Study. *Arch Phys Med Rehabil*, 80(2), 130-5 (1999)
DOI: 10.1016/S0003-9993(99)90109-0
6. L. A. West, S. Cole, D. Goodkind and W. He: 65+ in the United States: 2010. In: *Current Population Reports*. United States Census Bureau, (2014)
7. T. P. Wycherley, M. Noakes, P. M. Clifton, X.

- Cleanthous, J. B. Keogh and G. D. Brinkworth: A high-protein diet with resistance exercise training improves weight loss and body composition in overweight and obese patients with type 2 diabetes. *Diabetes Care*, 33(5), 969-76 (2010)
DOI: 10.2337/dc09-1974
8. D. K. Layman, E. Evans, J. I. Baum, J. Seyler, D. J. Erickson and R. A. Boileau: Dietary protein and exercise have additive effects on body composition during weight loss in adult women. *J Nutr*, 135(8), 1903-10 (2005)
 9. R. N. Baumgartner, K. M. Koehler, D. Gallagher, L. Romero, S. B. Heymsfield, R. R. Ross, P. J. Garry and R. D. Lindeman: Epidemiology of sarcopenia among the elderly in New Mexico. *Am J Epidemiol*, 147(8), 755-63 (1998)
DOI: 10.1093/oxfordjournals.aje.a009520
 10. W. R. Frontera, V. A. Hughes, R. A. Fielding, M. A. Fiatarone, W. J. Evans and R. Roubenoff: Aging of skeletal muscle: a 12-yr longitudinal study. *J Appl Physiol (1985)*, 88(4), 1321-6 (2000)
 11. L. Ferrucci, J. M. Guralnik, D. Buchner, J. Kasper, S. E. Lamb, E. M. Simonsick, M. C. Corti, K. Bandeen-Roche and L. P. Fried: Departures from linearity in the relationship between measures of muscular strength and physical performance of the lower extremities: the Women's Health and Aging Study. *J Gerontol A Biol Sci Med Sci*, 52(5), M275-85 (1997)
DOI: 10.1093/gerona/52A.5.M275
 12. B. H. Goodpaster, S. W. Park, T. B. Harris, S. B. Kritchevsky, M. Nevitt, A. V. Schwartz, E. M. Simonsick, F. A. Tyllavsky, M. Visser and A. B. Newman: The loss of skeletal muscle strength, mass, and quality in older adults: the health, aging and body composition study. *J Gerontol A Biol Sci Med Sci*, 61(10), 1059-64 (2006)
DOI: 10.1093/gerona/61.10.1059
 13. E. Volpi, M. Sheffield-Moore, B. B. Rasmussen and R. R. Wolfe: Basal muscle amino acid kinetics and protein synthesis in healthy young and older men. *JAMA*, 286(10), 1206-12 (2001)
DOI: 10.1001/jama.286.10.1206
 14. C. S. Katsanos, H. Kobayashi, M. Sheffield-Moore, A. Aarsland and R. R. Wolfe: Aging is associated with diminished accretion of muscle proteins after the ingestion of a small bolus of essential amino acids. *Am J Clin Nutr*, 82(5), 1065-73 (2005)
 15. P. G. Giresi, E. J. Stevenson, J. Theilhaber, A. Koncarevic, J. Parkington, R. A. Fielding and S. C. Kandarian: Identification of a molecular signature of sarcopenia. *Physiol Genomics*, 21(2), 253-63 (2005)
DOI: 10.1152/physiolgenomics.00249.2004
 16. S. Welle, A. I. Brooks, J. M. Delehanty, N. Needler, K. Bhatt, B. Shah and C. A. Thornton: Skeletal muscle gene expression profiles in 20-29 year old and 65-71 year old women. *Exp Gerontol*, 39(3), 369-77 (2004)
DOI: 10.1016/j.exger.2003.11.011
 17. S. Welle, A. I. Brooks, J. M. Delehanty, N. Needler and C. A. Thornton: Gene expression profile of aging in human muscle. *Physiol Genomics*, 14(2), 149-59 (2003)
DOI: 10.1152/physiolgenomics.00049.2003
 18. A. Zykovich, A. Hubbard, J. M. Flynn, M. Tarnopolsky, M. F. Fraga, C. Kerksick, D. Ogborn, L. MacNeil, S. D. Mooney and S. Melov: Genome-wide DNA methylation changes with age in disease-free human skeletal muscle. *Aging Cell*, 13(2), 360-6 (2014)
DOI: 10.1111/accel.12180
 19. J. Lexell: Evidence for nervous system degeneration with advancing age. *J Nutr*, 127(5 Suppl), 1011S-1013S (1997)
 20. K. R. Short, M. L. Bigelow, J. Kahl, R. Singh, J. Coenen-Schimke, S. Raghavakaimal and K. S. Nair: Decline in skeletal muscle mitochondrial function with aging in humans. *Proc Natl Acad Sci U S A*, 102(15), 5618-23 (2005)
DOI: 10.1073/pnas.0501559102
 21. O. E. Rooyackers, D. B. Adey, P. A. Ades and K. S. Nair: Effect of age on *in vivo* rates of mitochondrial protein synthesis in human skeletal muscle. *Proc Natl Acad Sci U S A*, 93(26), 15364-9 (1996)
DOI: 10.1073/pnas.93.26.15364
 22. S. W. Lamberts, A. W. van den Beld and A. J. van der Lely: The endocrinology of aging. *Science*, 278(5337), 419-24 (1997)
DOI: 10.1126/science.278.5337.419
 23. T. J. Doherty: Invited review: Aging and sarcopenia. *J Appl Physiol (1985)*, 95(4), 1717-27 (2003)
DOI: 10.1152/jappphysiol.00347.2003

24. M. R. Deschenes: Effects of aging on muscle fibre type and size. *Sports Med*, 34(12), 809-24 (2004)
DOI: 10.2165/00007256-200434120-00002
25. C. Dutta and E. C. Hadley: The significance of sarcopenia in old age. *J Gerontol A Biol Sci Med Sci*, 50 Spec No, 1-4 (1995)
DOI: 10.1093/gerona/50A.Special_Issue.1
26. L. Larsson, G. Grimby and J. Karlsson: Muscle strength and speed of movement in relation to age and muscle morphology. *J Appl Physiol Respir Environ Exerc Physiol*, 46(3), 451-6 (1979)
27. M. Braga, A. P. Sinha Hikim, S. Datta, M. G. Ferrini, D. Brown, E. L. Kovacheva, N. F. Gonzalez-Cadavid and I. Sinha-Hikim: Involvement of oxidative stress and caspase 2-mediated intrinsic pathway signaling in age-related increase in muscle cell apoptosis in mice. *Apoptosis*, 13(6), 822-32 (2008)
DOI: 10.1007/s10495-008-0216-7
28. B. Chabi, V. Ljubcic, K. J. Menzies, J. H. Huang, A. Saleem and D. A. Hood: Mitochondrial function and apoptotic susceptibility in aging skeletal muscle. *Aging Cell*, 7(1), 2-12 (2008)
DOI: 10.1111/j.1474-9726.2007.00347.x
29. N. A. o. Sciences: Dietary Reference Intakes: The Essential Guide to Nutrient Requirements. In: Ed J. J. Otten, J. P. Hellwig&L. D. Meyers. Institute of Medicine, (2006)
30. G. Wu, F. W. Bazer, Z. Dai, D. Li, J. Wang and Z. Wu: Amino acid nutrition in animals: protein synthesis and beyond. *Annu Rev Anim Biosci*, 2, 387-417 (2014)
DOI: 10.1146/annurev-animal-022513-114113
31. N. A. o. S. Institute of Medicine, Food and Nutrition Board.: Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acids, Cholesterol, Protein, and Amino Acids (Macronutrients). In, Washington, D.C. (2005)
32. W. W. Campbell, T. A. Trappe, R. R. Wolfe and W. J. Evans: The recommended dietary allowance for protein may not be adequate for older people to maintain skeletal muscle. *J Gerontol A Biol Sci Med Sci*, 56(6), M373-80 (2001)
DOI: 10.1093/gerona/56.6.M373
33. A. E. Thalacker-Mercer, J. C. Fleet, B. A. Craig, N. S. Carnell and W. W. Campbell: Inadequate protein intake affects skeletal muscle transcript profiles in older humans. *Am J Clin Nutr*, 85(5), 1344-52 (2007)
34. M. Tang, G. P. McCabe, R. Elango, P. B. Pencharz, R. O. Ball and W. W. Campbell: Assessment of protein requirement in octogenarian women with use of the indicator amino acid oxidation technique. *Am J Clin Nutr*, 99(4), 891-8 (2014)
DOI: 10.3945/ajcn.112.042325
35. M. Rafii, K. Chapman, J. Owens, R. Elango, W. W. Campbell, R. O. Ball, P. B. Pencharz and G. Courtney-Martin: Dietary protein requirement of female adults >65 years determined by the indicator amino Acid oxidation technique is higher than current recommendations. *J Nutr*, 145(1), 18-24 (2015)
DOI: 10.3945/jn.114.197517
36. D. K. Houston, B. J. Nicklas, J. Ding, T. B. Harris, F. A. Tylavsky, A. B. Newman, J. S. Lee, N. R. Sahyoun, M. Visser and S. B. Kritchevsky: Dietary protein intake is associated with lean mass change in older, community-dwelling adults: the Health, Aging, and Body Composition (Health ABC) Study. *Am J Clin Nutr*, 87(1), 150-5 (2008)
37. D. Misra, S. D. Berry, K. E. Broe, R. R. McLean, L. A. Cupples, K. L. Tucker, D. P. Kiel and M. T. Hannan: Does dietary protein reduce hip fracture risk in elders? The Framingham Osteoporosis Study. *Osteoporos Int*, 22(1), 345-9 (2011)
DOI: 10.1007/s00198-010-1179-4
38. E. S. Zoltick, S. Sahni, R. R. McLean, L. Quach, V. A. Casey and M. T. Hannan: Dietary protein intake and subsequent falls in older men and women: the Framingham Study. *J Nutr Health Aging*, 15(2), 147-52 (2011)
DOI: 10.1007/s12603-011-0028-2
39. D. J. Millward and A. A. Jackson: Protein/energy ratios of current diets in developed and developing countries compared with a safe protein/energy ratio: implications for recommended protein and amino acid intakes. *Public Health Nutr*, 7(3), 387-405 (2004)
DOI: 10.1079/PHN2003545
40. D. Paddon-Jones and B. B. Rasmussen: Dietary protein recommendations and the prevention of sarcopenia. *Curr Opin Clin Nutr Metab Care*, 12(1), 86-90 (2009)
DOI: 10.1097/MCO.0b013e32831cef8b

41. D. Cuthbertson, K. Smith, J. Babraj, G. Leese, T. Waddell, P. Atherton, H. Wackerhage, P. M. Taylor and M. J. Rennie: Anabolic signaling deficits underlie amino acid resistance of wasting, aging muscle. *FASEB J*, 19(3), 422-4 (2005)
42. J. Rosenthal, A. Angel and J. Farkas: Metabolic fate of leucine: a significant sterol precursor in adipose tissue and muscle. *Am J Physiol*, 226(2), 411-8 (1974)
43. N. A. o. S. Institute of Medicine, Food and Nutrition Board.: Dietary Reference Intakes: Macronutrients. In, (2012)
44. J. C. Anthony, T. G. Anthony, S. R. Kimball and L. S. Jefferson: Signaling pathways involved in translational control of protein synthesis in skeletal muscle by leucine. *J Nutr*, 131(3), 856S-860S (2001)
45. T. Yoshimasa, K. Nakao, H. Ohtsuki, S. Li and H. Imura: Methionine-enkephalin and leucine-enkephalin in human sympathoadrenal system and pheochromocytoma. *J Clin Invest*, 69(3), 643-50 (1982)
DOI: 10.1172/JCI110491
46. W. H. Landschulz, P. F. Johnson and S. L. McKnight: The leucine zipper: a hypothetical structure common to a new class of DNA binding proteins. *Science*, 240(4860), 1759-64 (1988)
DOI: 10.1126/science.3289117
47. E. Volpi, H. Kobayashi, M. Sheffield-Moore, B. Mittendorfer and R. R. Wolfe: Essential amino acids are primarily responsible for the amino acid stimulation of muscle protein anabolism in healthy elderly adults. *Am J Clin Nutr*, 78(2), 250-8 (2003)
48. M. K. Holz, B. A. Ballif, S. P. Gygi and J. Blenis: mTOR and S6K1 mediate assembly of the translation preinitiation complex through dynamic protein interchange and ordered phosphorylation events. *Cell*, 123(4), 569-80 (2005)
DOI: 10.1016/j.cell.2005.10.024
49. A. Pause, G. J. Belsham, A. C. Gingras, O. Donze, T. A. Lin, J. C. Lawrence, Jr. and N. Sonenberg: Insulin-dependent stimulation of protein synthesis by phosphorylation of a regulator of 5'-cap function. *Nature*, 371(6500), 762-7 (1994)
DOI: 10.1038/371762a0
50. A. Haghghat and N. Sonenberg: eIF4G dramatically enhances the binding of eIF4E to the mRNA 5'-cap structure. *J Biol Chem*, 272(35), 21677-80 (1997)
51. J. C. Anthony, F. Yoshizawa, T. G. Anthony, T. C. Vary, L. S. Jefferson and S. R. Kimball: Leucine stimulates translation initiation in skeletal muscle of postabsorptive rats via a rapamycin-sensitive pathway. *J Nutr*, 130(10), 2413-9 (2000)
52. G. J. Wilson, D. K. Layman, C. J. Moulton, L. E. Norton, T. G. Anthony, C. G. Proud, S. I. Rupassara and P. J. Garlick: Leucine or carbohydrate supplementation reduces AMPK and eEF2 phosphorylation and extends postprandial muscle protein synthesis in rats. *Am J Physiol Endocrinol Metab*, 301(6), E1236-42 (2011)
DOI: 10.1152/ajpendo.00242.2011
53. C. S. Katsanos, H. Kobayashi, M. Sheffield-Moore, A. Aarsland and R. R. Wolfe: A high proportion of leucine is required for optimal stimulation of the rate of muscle protein synthesis by essential amino acids in the elderly. *Am J Physiol Endocrinol Metab*, 291(2), E381-7 (2006)
DOI: 10.1152/ajpendo.00488.2005
54. Y. Yang, L. Breen, N. A. Burd, A. J. Hector, T. A. Churchward-Venne, A. R. Josse, M. A. Tarnopolsky and S. M. Phillips: Resistance exercise enhances myofibrillar protein synthesis with graded intakes of whey protein in older men. *Br J Nutr*, 108(10), 1780-8 (2012)
DOI: 10.1017/S0007114511007422
55. Z. R. Xu, Z. J. Tan, Q. Zhang, Q. F. Gui and Y. M. Yang: The effectiveness of leucine on muscle protein synthesis, lean body mass and leg lean mass accretion in older people: a systematic review and meta-analysis. *Br J Nutr*, 1-10 (2014)
56. E. L. Glynn, C. S. Fry, M. J. Drummond, K. L. Timmerman, S. Dhanani, E. Volpi and B. B. Rasmussen: Excess leucine intake enhances muscle anabolic signaling but not net protein anabolism in young men and women. *J Nutr*, 140(11), 1970-6 (2010)
DOI: 10.3945/jn.110.127647
57. E. Borsheim, Q. U. Bui, S. Tissier, H. Kobayashi, A. A. Ferrando and R. R. Wolfe: Effect of amino acid supplementation on muscle mass, strength and physical function

- in elderly. *Clin Nutr*, 27(2), 189-95 (2008)
DOI: 10.1016/j.clnu.2008.01.001
58. S. B. Solerte, C. Gazzaruso, R. Bonacasa, M. Rondanelli, M. Zamboni, C. Basso, E. Locatelli, N. Schifino, A. Giustina and M. Fioravanti: Nutritional supplements with oral amino acid mixtures increases whole-body lean mass and insulin sensitivity in elderly subjects with sarcopenia. *Am J Cardiol*, 101(11A), 69E-77E (2008)
DOI: 10.1016/j.amjcard.2008.03.004
 59. E. L. Dillon, M. Sheffield-Moore, D. Paddon-Jones, C. Gilkison, A. P. Sanford, S. L. Casperson, J. Jiang, D. L. Chinkes and R. J. Urban: Amino acid supplementation increases lean body mass, basal muscle protein synthesis, and insulin-like growth factor-I expression in older women. *J Clin Endocrinol Metab*, 94(5), 1630-7 (2009)
DOI: 10.1210/jc.2008-1564
 60. D. R. Moore, T. A. Churchward-Venne, O. Witard, L. Breen, N. A. Burd, K. D. Tipton and S. M. Phillips: Protein ingestion to stimulate myofibrillar protein synthesis requires greater relative protein intakes in healthy older versus younger men. *J Gerontol A Biol Sci Med Sci*, 70(1), 57-62 (2015)
DOI: 10.1093/gerona/glu103
 61. T. B. Symons, S. E. Schutzler, T. L. Cocke, D. L. Chinkes, R. R. Wolfe and D. Paddon-Jones: Aging does not impair the anabolic response to a protein-rich meal. *Am J Clin Nutr*, 86(2), 451-6 (2007)
 62. M. L. Dirks, B. T. Wall, R. Nilwik, D. H. Weerts, L. B. Verdijk and L. J. van Loon: Skeletal muscle disuse atrophy is not attenuated by dietary protein supplementation in healthy older men. *J Nutr*, 144(8), 1196-203 (2014)
DOI: 10.3945/jn.114.194217
 63. G. Q. Fryar CD, Ogden CL.: Anthropometric Reference Data for Children and Adults: United States, 2007-2010. In: *Vital Health Statistics*. National Center for Health Statistics, (2012)
 64. S. L. Casperson, M. Sheffield-Moore, S. J. Hewlings and D. Paddon-Jones: Leucine supplementation chronically improves muscle protein synthesis in older adults consuming the RDA for protein. *Clin Nutr*, 31(4), 512-9 (2012)
DOI: 10.1016/j.clnu.2012.01.005
 65. S. Verhoeven, K. Vanschoonbeek, L. B. Verdijk, R. Koopman, W. K. Wodzig, P. Dendale and L. J. van Loon: Long-term leucine supplementation does not increase muscle mass or strength in healthy elderly men. *Am J Clin Nutr*, 89(5), 1468-75 (2009)
DOI: 10.3945/ajcn.2008.26668
 66. M. Leenders, L. B. Verdijk, L. van der Hoeven, J. van Kranenburg, F. Hartgens, W. K. Wodzig, W. H. Saris and L. J. van Loon: Prolonged leucine supplementation does not augment muscle mass or affect glycemic control in elderly type 2 diabetic men. *J Nutr*, 141(6), 1070-6 (2011)
DOI: 10.3945/jn.111.138495
 67. P. Wakimoto and G. Block: Dietary intake, dietary patterns, and changes with age: an epidemiological perspective. *J Gerontol A Biol Sci Med Sci*, 56 Spec No 2, 65-80 (2001)
DOI: 10.1093/gerona/56.suppl_2.65
 68. J. E. Morley: Anorexia of aging: physiologic and pathologic. *Am J Clin Nutr*, 66(4), 760-73 (1997)
 69. M. A. Fiatarone Singh, M. A. Bernstein, A. D. Ryan, E. F. O'Neill, K. M. Clements and W. J. Evans: The effect of oral nutritional supplements on habitual dietary quality and quantity in frail elders. *J Nutr Health Aging*, 4(1), 5-12 (2000)
 70. D. Paddon-Jones, M. Sheffield-Moore, A. Aarsland, R. R. Wolfe and A. A. Ferrando: Exogenous amino acids stimulate human muscle anabolism without interfering with the response to mixed meal ingestion. *Am J Physiol Endocrinol Metab*, 288(4), E761-7 (2005)
DOI: 10.1152/ajpendo.00291.2004
 71. A. A. Ferrando, D. Paddon-Jones, N. P. Hays, P. Kortebein, O. Ronsen, R. H. Williams, A. McComb, T. B. Symons, R. R. Wolfe and W. Evans: EAA supplementation to increase nitrogen intake improves muscle function during bed rest in the elderly. *Clin Nutr*, 29(1), 18-23 (2010)
DOI: 10.1016/j.clnu.2009.03.009
 72. M. J. Drummond, J. M. Dickinson, C. S. Fry, D. K. Walker, D. M. Gundermann, P. T. Reidy, K. L. Timmerman, M. M. Markofski, D. Paddon-Jones, B. B. Rasmussen and E. Volpi: Bed rest impairs skeletal muscle amino acid transporter expression, mTORC1 signaling,

- and protein synthesis in response to essential amino acids in older adults. *Am J Physiol Endocrinol Metab*, 302(9), E1113-22 (2012)
DOI: 10.1152/ajpendo.00603.2011
73. S. R. Fisher, Y. F. Kuo, J. E. Graham, K. J. Ottenbacher and G. V. Ostir: Early ambulation and length of stay in older adults hospitalized for acute illness. *Arch Intern Med*, 170(21), 1942-3 (2010)
DOI: 10.1001/archinternmed.2010.422
 74. P. Kortebein, T. B. Symons, A. Ferrando, D. Paddon-Jones, O. Ronsen, E. Protas, S. Conger, J. Lombeida, R. Wolfe and W. J. Evans: Functional impact of 10 days of bed rest in healthy older adults. *J Gerontol A Biol Sci Med Sci*, 63(10), 1076-81 (2008)
DOI: 10.1093/gerona/63.10.1076
 75. M. G. Cree, D. Paddon-Jones, B. R. Newcomer, O. Ronsen, A. Aarsland, R. R. Wolfe and A. Ferrando: Twenty-eight-day bed rest with hypercortisolemia induces peripheral insulin resistance and increases intramuscular triglycerides. *Metabolism*, 59(5), 703-10 (2010)
DOI: 10.1016/j.metabol.2009.09.014
 76. C. Suetta, L. G. Hvid, L. Justesen, U. Christensen, K. Neergaard, L. Simonsen, N. Ortenblad, S. P. Magnusson, M. Kjaer and P. Aagaard: Effects of aging on human skeletal muscle after immobilization and retraining. *J Appl Physiol* (1985), 107(4), 1172-80 (2009)
DOI: 10.1152/jappphysiol.00290.2009
 77. C. Suetta, U. Frandsen, A. L. Mackey, L. Jensen, L. G. Hvid, M. L. Bayer, S. J. Petersson, H. D. Schroder, J. L. Andersen, P. Aagaard, P. Schjerling and M. Kjaer: Ageing is associated with diminished muscle re-growth and myogenic precursor cell expansion early after immobility-induced atrophy in human skeletal muscle. *J Physiol*, 591(Pt 15), 3789-804 (2013)
DOI: 10.1113/jphysiol.2013.257121
 78. S. V. Amely M Verreijen, Marielle F Engberink, Sophie Swinkels, Johan de Vogel-van den Bosch, and Peter JM Weijs: A high whey protein-, leucine-, and vitamin D-enriched supplement preserves muscle mass during intentional weight loss in obese older adults: a double-blind randomized controlled trial. *Am J Clin Nutr* (2015)
 79. P. Srikanthan, A. L. Hevener and A. S. Karlamangla: Sarcopenia exacerbates obesity-associated insulin resistance and dysglycemia: findings from the National Health and Nutrition Examination Survey III. *PLoS One*, 5(5), e10805 (2010)
DOI: 10.1371/journal.pone.0010805
 80. E. Zoico, V. Di Francesco, J. M. Guralnik, G. Mazzali, A. Bortolani, S. Guariento, G. Sergi, O. Bosello and M. Zamboni: Physical disability and muscular strength in relation to obesity and different body composition indexes in a sample of healthy elderly women. *Int J Obes Relat Metab Disord*, 28(2), 234-41 (2004)
DOI: 10.1038/sj.ijo.0802552
 81. J. Filipovsky, P. Ducimetiere, B. Darne and J. L. Richard: Abdominal body mass distribution and elevated blood pressure are associated with increased risk of death from cardiovascular diseases and cancer in middle-aged men. The results of a 15- to 20-year follow-up in the Paris prospective study I. *Int J Obes Relat Metab Disord*, 17(4), 197-203 (1993)
 82. G. R. Hunter, T. Kekes-Szabo, S. W. Snyder, C. Nicholson, I. Nyikos and L. Berland: Fat distribution, physical activity, and cardiovascular risk factors. *Med Sci Sports Exerc*, 29(3), 362-9 (1997)
DOI: 10.1097/00005768-199703000-00011
 83. M. J. Williams, G. R. Hunter, T. Kekes-Szabo, S. Snyder and M. S. Treuth: Regional fat distribution in women and risk of cardiovascular disease. *Am J Clin Nutr*, 65(3), 855-60 (1997)
 84. C. L. Ogden, M. D. Carroll, B. K. Kit and K. M. Flegal: Prevalence of obesity in the United States, 2009-2010. *NCHS Data Brief*(82), 1-8 (2012)
 85. D. K. Layman, R. A. Boileau, D. J. Erickson, J. E. Painter, H. Shiue, C. Sather and D. D. Christou: A reduced ratio of dietary carbohydrate to protein improves body composition and blood lipid profiles during weight loss in adult women. *J Nutr*, 133(2), 411-7 (2003)
 86. T. L. Halton and F. B. Hu: The effects of high protein diets on thermogenesis, satiety and weight loss: a critical review. *J Am Coll Nutr*, 23(5), 373-85 (2004)
DOI: 10.1080/07315724.2004.10719381
 87. E. Farnsworth, N. D. Luscombe, M. Noakes,

- G. Wittert, E. Argyiou and P. M. Clifton: Effect of a high-protein, energy-restricted diet on body composition, glycemic control, and lipid concentrations in overweight and obese hyperinsulinemic men and women. *Am J Clin Nutr*, 78(1), 31-9 (2003)
88. S. M. Pasiakos, J. J. Cao, L. M. Margolis, E. R. Sauter, L. D. Whigham, J. P. McClung, J. C. Rood, J. W. Carbone, G. F. Combs, Jr. and A. J. Young: Effects of high-protein diets on fat-free mass and muscle protein synthesis following weight loss: a randomized controlled trial. *FASEB J*, 27(9), 3837-47 (2013)
DOI: 10.1096/fj.13-230227
 89. S. O. Hong and D. K. Layman: Effects of leucine on *in vitro* protein synthesis and degradation in rat skeletal muscles. *J Nutr*, 114(7), 1204-12 (1984)
 90. D. K. Layman and D. A. Walker: Potential importance of leucine in treatment of obesity and the metabolic syndrome. *J Nutr*, 136(1 Suppl), 319S-23S (2006)
 91. S. R. Kimball and L. S. Jefferson: Regulation of protein synthesis by branched-chain amino acids. *Curr Opin Clin Nutr Metab Care*, 4(1), 39-43 (2001)
DOI: 10.1097/00075197-200101000-00008
 92. R. H. Coker, S. Miller, S. Schutzler, N. Deutz and R. R. Wolfe: Whey protein and essential amino acids promote the reduction of adipose tissue and increased muscle protein synthesis during caloric restriction-induced weight loss in elderly, obese individuals. *Nutr J*, 11, 105 (2012)
DOI: 10.1186/1475-2891-11-105
 93. A. R. Josse, S. A. Atkinson, M. A. Tarnopolsky and S. M. Phillips: Increased consumption of dairy foods and protein during diet- and exercise-induced weight loss promotes fat mass loss and lean mass gain in overweight and obese premenopausal women. *J Nutr*, 141(9), 1626-34 (2011)
DOI: 10.3945/jn.111.141028
 94. M. van Nielen, E. J. Feskens, M. Mensink, I. Sluijs, E. Molina, P. Amiano, E. Ardanaz, B. Balkau, J. W. Beulens, H. Boeing, F. Clavel-Chapelon, G. Fagherazzi, P. W. Franks, J. Halkjaer, J. M. Huerta, V. Katzke, T. J. Key, K. T. Khaw, V. Krogh, T. Kuhn, V. V. Menendez, P. Nilsson, K. Overvad, D. Palli, S. Panico, O. Rolandsson, I. Romieu, C. Sacerdote, M. J. Sanchez, M. B. Schulze, A. M. Spijkerman, A. Tjonneland, R. Tumino, A. D. van der, A. M. Wurtz, R. Zamora-Ros, C. Langenberg, S. J. Sharp, N. G. Forouhi, E. Riboli and N. J. Wareham: Dietary protein intake and incidence of type 2 diabetes in Europe: the EPIC-InterAct Case-Cohort Study. *Diabetes Care*, 37(7), 1854-62 (2014)
DOI: 10.2337/dc13-2627
 95. U. Ericson, E. Sonestedt, B. Gullberg, S. Hellstrand, G. Hindy, E. Wirfalt and M. Orho-Melander: High intakes of protein and processed meat associate with increased incidence of type 2 diabetes. *Br J Nutr*, 109(6), 1143-53 (2013)
DOI: 10.1017/S0007114512003017
 96. J. C. Floyd, Jr., S. S. Fajans, J. W. Conn, R. F. Knopf and J. Rull: Stimulation of insulin secretion by amino acids. *J Clin Invest*, 45(9), 1487-502 (1966)
DOI: 10.1172/JCI1105456
 97. B. Parker, M. Noakes, N. Luscombe and P. Clifton: Effect of a high-protein, high-monounsaturated fat weight loss diet on glycemic control and lipid levels in type 2 diabetes. *Diabetes Care*, 25(3), 425-30 (2002)
DOI: 10.2337/diacare.25.3.425
 98. G. I. Shulman, D. L. Rothman, T. Jue, P. Stein, R. A. DeFronzo and R. G. Shulman: Quantitation of muscle glycogen synthesis in normal subjects and subjects with non-insulin-dependent diabetes by ¹³C nuclear magnetic resonance spectroscopy. *N Engl J Med*, 322(4), 223-8 (1990)
DOI: 10.1056/NEJM199001253220403
 99. R. A. DeFronzo, E. Jacot, E. Jequier, E. Maeder, J. Wahren and J. P. Felber: The effect of insulin on the disposal of intravenous glucose. Results from indirect calorimetry and hepatic and femoral venous catheterization. *Diabetes*, 30(12), 1000-7 (1981)
DOI: 10.2337/diab.30.12.1000
 100. C. B. Newgard, J. An, J. R. Bain, M. J. Muehlbauer, R. D. Stevens, L. F. Lien, A. M. Haqq, S. H. Shah, M. Arlotto, C. A. Slentz, J. Rochon, D. Gallup, O. Ilkayeva, B. R. Wenner, W. S. Yancy, Jr., H. Eisenson, G. Musante, R. S. Surwit, D. S. Millington, M. D. Butler and L. P. Svetkey: A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and

- contributes to insulin resistance. *Cell Metab*, 9(4), 311-26 (2009)
DOI: 10.1016/j.cmet.2009.02.002
101. T. J. Wang, M. G. Larson, R. S. Vasan, S. Cheng, E. P. Rhee, E. McCabe, G. D. Lewis, C. S. Fox, P. F. Jacques, C. Fernandez, C. J. O'Donnell, S. A. Carr, V. K. Mootha, J. C. Florez, A. Souza, O. Melander, C. B. Clish and R. E. Gerszten: Metabolite profiles and the risk of developing diabetes. *Nat Med*, 17(4), 448-53 (2011)
DOI: 10.1038/nm.2307
 102. S. E. McCormack, O. Shaham, M.A. McCarthy, A. A. Deik, T. J. Wang, R. E. Gerszten, C. B. Clish, V. K. Mootha, S. K. Grinspoon and A. Fleischman: Circulating branched-chain amino acid concentrations are associated with obesity and future insulin resistance in children and adolescents. *Pediatr Obes*, 8(1), 52-61 (2013)
DOI: 10.1111/j.2047-6310.2012.00087.x
 103. C. B. Newgard: Interplay between lipids and branched-chain amino acids in development of insulin resistance. *Cell Metab*, 15(5), 606-14 (2012)
DOI: 10.1016/j.cmet.2012.01.024
 104. A. E. Thalacker-Mercer, K. H. Ingram, F. Guo, O. Ilkayeva, C. B. Newgard and W. T. Garvey: BMI, RQ, diabetes, and sex affect the relationships between amino acids and clamp measures of insulin action in humans. *Diabetes*, 63(2), 791-800 (2014)
DOI: 10.2337/db13-0396
 105. B. Laferrere, D. Reilly, S. Arias, N. Swerdlow, P. Gorroochurn, B. Bawa, M. Bose, J. Teixeira, R. D. Stevens, B. R. Wenner, J. R. Bain, M. J. Muehlbauer, A. Haqq, L. Lien, S. H. Shah, L. P. Svetkey and C. B. Newgard: Differential metabolic impact of gastric bypass surgery versus dietary intervention in obese diabetic subjects despite identical weight loss. *Sci Transl Med*, 3(80), 80re2 (2011)
DOI: 10.1126/scitranslmed.3002043
 106. M. E. Levine, J. A. Suarez, S. Brandhorst, P. Balasubramanian, C. W. Cheng, F. Madia, L. Fontana, M. G. Mirisola, J. Guevara-Aguirre, J. Wan, G. Passarino, B. K. Kennedy, M. Wei, P. Cohen, E. M. Crimmins and V. D. Longo: Low protein intake is associated with a major reduction in IGF-1, cancer, and overall mortality in the 65 and younger but not older population. *Cell Metab*, 19(3), 407-17 (2014)
DOI: 10.1016/j.cmet.2014.02.006
 107. J. M. Olefsky and C. K. Glass: Macrophages, inflammation, and insulin resistance. *Annu Rev Physiol*, 72, 219-46 (2010)
DOI: 10.1146/annurev-physiol-021909-135846
DOI: 10.1146/annurev-physiol-021909-135846
 108. J. S. Burrill, E. K. Long, B. Reilly, Y. Deng, I. M. Armitage, P. E. Scherer and D. A. Bernlohr: Inflammation and ER Stress Regulate Branched-Chain Amino Acid Uptake and Metabolism in Adipocytes. *Mol Endocrinol*, 29(3), 411-20 (2015)
DOI: 10.1210/me.2014-1275
 109. M. M. Nellis, C. B. Doering, A. Kasinski and D. J. Danner: Insulin increases branched-chain alpha-ketoacid dehydrogenase kinase expression in Clone 9 rat cells. *Am J Physiol Endocrinol Metab*, 283(4), E853-60 (2002)
DOI: 10.1152/ajpendo.00133.2002
 110. H. Weiss, T. Friedrich, G. Hofhaus and D. Preis: The respiratory-chain NADH dehydrogenase (complex I) of mitochondria. *Eur J Biochem*, 197(3), 563-76 (1991)
DOI: 10.1111/j.1432-1033.1991.tb15945.x
 111. W. Droge: Oxidative stress and ageing: is ageing a cysteine deficiency syndrome? *Philos Trans R Soc Lond B Biol Sci*, 360(1464), 2355-72 (2005)
DOI: 10.1098/rstb.2005.1770
 112. N. K. Fukagawa, A. M. Ajami and V. R. Young: Plasma methionine and cysteine kinetics in response to an intravenous glutathione infusion in adult humans. *Am J Physiol*, 270(2 Pt 1), E209-14 (1996)
 113. N. K. Fukagawa, Y. M. Yu and V. R. Young: Methionine and cysteine kinetics at different intakes of methionine and cysteine in elderly men and women. *Am J Clin Nutr*, 68(2), 380-8 (1998)
 114. J. M. BH Lauterberg: Therapeutic doses of acetaminophen stimulate the turnover of cysteine and glutathione in man. *J Hepatol*(4), 206-211 (1987)
 115. K. J. Storch, D. A. Wagner, J. F. Burke and V. R. Young: Quantitative study *in vivo* of methionine cycle in humans using (methyl-2H3)- and (1-13C)methionine. *Am J Physiol*, 255(3 Pt 1), E322-31 (1988)
 116. J. I. Lee, M. Londono, L. L. Hirschberger

- and M. H. Stipanuk: Regulation of cysteine dioxygenase and gamma-glutamylcysteine synthetase is associated with hepatic cysteine level. *J Nutr Biochem*, 15(2), 112-22 (2004)
DOI: 10.1016/j.jnutbio.2003.10.005
117. M. H. Stipanuk, R. M. Coloso, R. A. Garcia and M. F. Banks: Cysteine concentration regulates cysteine metabolism to glutathione, sulfate and taurine in rat hepatocytes. *J Nutr*, 122(3), 420-7 (1992)
118. M. H. Stipanuk, J. E. Dominy, Jr., J. I. Lee and R. M. Coloso: Mammalian cysteine metabolism: new insights into regulation of cysteine metabolism. *J Nutr*, 136(6 Suppl), 1652S-1659S (2006)
119. M. H. Stipanuk, M. Londono, J. I. Lee, M. Hu and A. F. Yu: Enzymes and metabolites of cysteine metabolism in nonhepatic tissues of rats show little response to changes in dietary protein or sulfur amino acid levels. *J Nutr*, 132(11), 3369-78 (2002)
120. R. S. Balaban, S. Nemoto and T. Finkel: Mitochondria, oxidants, and aging. *Cell*, 120(4), 483-95 (2005)
DOI: 10.1016/j.cell.2005.02.001
121. C. Martin, H. Dubouchaud, L. Mosoni, J. M. Chardigny, A. Oudot, E. Fontaine, C. Vergely, C. Keriel, L. Rochette, X. Leverve and L. Demaison: Abnormalities of mitochondrial functioning can partly explain the metabolic disorders encountered in sarcopenic gastrocnemius. *Aging Cell*, 6(2), 165-77 (2007)
DOI: 10.1111/j.1474-9726.2007.00271.x
122. S. J. Stohs, T. Lawson and W. A. Al-Turk: Changes in glutathione and glutathione metabolizing enzymes in erythrocytes and lymphocytes of mice as a function of age. *Gen Pharmacol*, 15(3), 267-70 (1984)
DOI: 10.1016/0306-3623(84)90173-3
123. M. Y. Farooqui, W. W. Day and D. M. Zamorano: Glutathione and lipid peroxidation in the aging rat. *Comp Biochem Physiol B*, 88(1), 177-80 (1987)
DOI: 10.1016/0305-0491(87)90097-6
124. L. Mosoni, D. Breuille, C. Buffiere, C. Obled and P. P. Mirand: Age-related changes in glutathione availability and skeletal muscle carbonyl content in healthy rats. *Exp Gerontol*, 39(2), 203-10 (2004)
DOI: 10.1016/j.exger.2003.10.014
125. P. S. Samiec, C. Drews-Botsch, E. W. Flagg, J. C. Kurtz, P. Sternberg, Jr., R. L. Reed and D. P. Jones: Glutathione in human plasma: decline in association with aging, age-related macular degeneration, and diabetes. *Free Radic Biol Med*, 24(5), 699-704 (1998)
DOI: 10.1016/S0891-5849(97)00286-4
126. W. A. Al-Turk, S. J. Stohs, F. H. el-Rashidy and S. Othman: Changes in glutathione and its metabolizing enzymes in human erythrocytes and lymphocytes with age. *J Pharm Pharmacol*, 39(1), 13-6 (1987)
DOI: 10.1111/j.2042-7158.1987.tb07154.x
127. R. V. Sekhar, S. G. Patel, A. P. Guthikonda, M. Reid, A. Balasubramanyam, G. E. Taffet and F. Jahoor: Deficient synthesis of glutathione underlies oxidative stress in aging and can be corrected by dietary cysteine and glycine supplementation. *Am J Clin Nutr*, 94(3), 847-53 (2011)
DOI: 10.3945/ajcn.110.003483
128. A. D. Dam, A. S. Mitchell, J. W. Rush and J. Quadriatero: Elevated skeletal muscle apoptotic signaling following glutathione depletion. *Apoptosis*, 17(1), 48-60 (2012)
DOI: 10.1007/s10495-011-0654-5
129. I. Sinha-Hikim, A. P. Sinha-Hikim, M. Parveen, R. Shen, R. Goswami, P. Tran, A. Crum and K. C. Norris: Long-term supplementation with a cystine-based antioxidant delays loss of muscle mass in aging. *J Gerontol A Biol Sci Med Sci*, 68(7), 749-59 (2013)
DOI: 10.1093/gerona/gls334
130. A. Badaloo, M. Reid, T. Forrester, W. C. Heird and F. Jahoor: Cysteine supplementation improves the erythrocyte glutathione synthesis rate in children with severe edematous malnutrition. *Am J Clin Nutr*, 76(3), 646-52 (2002)
131. R. V. Sekhar, S. V. McKay, S. G. Patel, A. P. Guthikonda, V. T. Reddy, A. Balasubramanyam and F. Jahoor: Glutathione synthesis is diminished in patients with uncontrolled diabetes and restored by dietary supplementation with cysteine and glycine. *Diabetes Care*, 34(1), 162-7 (2011)
DOI: 10.2337/dc10-1006
132. D. Nguyen, J. W. Hsu, F. Jahoor and R. V. Sekhar: Effect of increasing glutathione with

- cysteine and glycine supplementation on mitochondrial fuel oxidation, insulin sensitivity, and body composition in older HIV-infected patients. *J Clin Endocrinol Metab*, 99(1), 169-77 (2014)
DOI: 10.1210/jc.2013-2376
133. K. Vidal, D. Breuille, P. Serrant, P. Denis, F. Glomot, F. Bechereau and I. Papet: Long-term cysteine fortification impacts cysteine/glutathione homeostasis and food intake in ageing rats. *Eur J Nutr*, 53(3), 963-71 (2014)
DOI: 10.1007/s00394-013-0600-0
134. N. Hernadfalvi, W. Langhans, C. von Meyenburg, B. Onteniente, D. Richard and D. Arsenijevic: Role for glutathione in the hyposensitivity of LPS-pretreated mice to LPS anorexia. *Eur Cytokine Netw*, 18(2), 86-92 (2007)
135. G. B. Frisoni, S. Franzoni, R. Rozzini, L. Ferrucci, S. Boffelli and M. Trabucchi: Food intake and mortality in the frail elderly. *J Gerontol A Biol Sci Med Sci*, 50(4), M203-10 (1995)
DOI: 10.1093/gerona/50A.4.M203
136. V. Grey, S. R. Mohammed, A. A. Smountas, R. Bahloul and L. C. Lands: Improved glutathione status in young adult patients with cystic fibrosis supplemented with whey protein. *J Cyst Fibros*, 2(4), 195-8 (2003)
DOI: 10.1016/S1569-1993(03)00097-3
137. T. Chitapanarux, P. Tienboon, S. Pojchamarnwiputh and D. Leelarungrayub: Open-labeled pilot study of cysteine-rich whey protein isolate supplementation for nonalcoholic steatohepatitis patients. *J Gastroenterol Hepatol*, 24(6), 1045-50 (2009)
DOI: 10.1111/j.1440-1746.2009.05865.x
138. P. Micke, K. M. Beeh, J. F. Schlaak and R. Buhl: Oral supplementation with whey proteins increases plasma glutathione levels of HIV-infected patients. *Eur J Clin Invest*, 31(2), 171-8 (2001)
DOI: 10.1046/j.1365-2362.2001.00781.x
139. P. G. Reeves, F. H. Nielsen and G. C. Fahey, Jr.: AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76A rodent diet. *J Nutr*, 123(11), 1939-51 (1993)
140. G. F. Rushworth and I. L. Megson: Existing and potential therapeutic uses for N-acetylcysteine: the need for conversion to intracellular glutathione for antioxidant benefits. *Pharmacol Ther*, 141(2), 150-9 (2014)
DOI: 10.1016/j.pharmthera.2013.09.006
141. P. Porta, S. Aebi, K. Summer and B. H. Lauterburg: L-2-oxothiazolidine-4-carboxylic acid, a cysteine prodrug: pharmacokinetics and effects on thiols in plasma and lymphocytes in human. *J Pharmacol Exp Ther*, 257(1), 331-4 (1991)
142. M. F. Banks and M. H. Stipanuk: The utilization of N-acetylcysteine and 2-oxothiazolidine-4-carboxylate by rat hepatocytes is limited by their rate of uptake and conversion to cysteine. *J Nutr*, 124(3), 378-87 (1994)
143. J. L. Ensunsa, L. L. Hirschberger and M. H. Stipanuk: Catabolism of cysteine, cystine, cysteinesulfinate, and OTC by isolated perfused rat hindquarter. *Am J Physiol*, 264(5 Pt 1), E782-9 (1993)
144. R. M. Coloso, L. L. Hirschberger and M. H. Stipanuk: Uptake and metabolism of L-2-oxo-(35S)thiazolidine-4-carboxylate by rat cells is slower than that of L-(35S)cysteine or L-(35S) methionine. *J Nutr*, 121(9), 1341-8 (1991)
145. R. M. Coloso, M. R. Drake and M. H. Stipanuk: Effect of bathocuproine disulfonate, a copper chelator, on cyst(e)ine metabolism by freshly isolated rat hepatocytes. *Am J Physiol*, 259(3 Pt 1), E443-50 (1990)
146. R. J. Huxtable: Physiological actions of taurine. *Physiol Rev*, 72(1), 101-63 (1992)
147. R. A. Chapman, M. S. Suleiman and Y. E. Earm: Taurine and the heart. *Cardiovasc Res*, 27(3), 358-63 (1993)
DOI: 10.1093/cvr/27.3.358
148. U. Warskulat, U. Flogel, C. Jacoby, H. G. Hartwig, M. Thewissen, M. W. Merx, A. Molojavyi, B. Heller-Stilb, J. Schrader and D. Haussinger: Taurine transporter knockout depletes muscle taurine levels and results in severe skeletal muscle impairment but leaves cardiac function uncompromised. *FASEB J*, 18(3), 577-9 (2004)
DOI: 10.1096/fj.03-0496fje
149. E. J. Hamilton, H. M. Berg, C. J. Easton and A. J. Bakker: The effect of taurine depletion on the contractile properties and fatigue in

- fast-twitch skeletal muscle of the mouse. *Amino Acids*, 31(3), 273-8 (2006)
DOI: 10.1007/s00726-006-0291-4
150. T. Ito, N. Yoshikawa, T. Inui, N. Miyazaki, S. W. Schaffer and J. Azuma: Tissue depletion of taurine accelerates skeletal muscle senescence and leads to early death in mice. *PLoS One*, 9(9), e107409 (2014)
DOI: 10.1371/journal.pone.0107409
 151. D. R. W. Ralph Dawson Jr.: Taurine Content in Tissues From Aged Fischer 344 RAts. *Age* 15, 73-81 (1992)
DOI: 10.1007/BF02435005
 152. S. Pierno, A. De Luca, C. Camerino, R. J. Huxtable and D. C. Camerino: Chronic administration of taurine to aged rats improves the electrical and contractile properties of skeletal muscle fibers. *J Pharmacol Exp Ther*, 286(3), 1183-90 (1998)
 153. M. Jeevanandam, D. H. Young, L. Ramias and W. R. Schiller: Effect of major trauma on plasma free amino acid concentrations in geriatric patients. *Am J Clin Nutr*, 51(6), 1040-5 (1990)
 154. B. Eppler and R. Dawson, Jr.: Dietary taurine manipulations in aged male Fischer 344 rat tissue: taurine concentration, taurine biosynthesis, and oxidative markers. *Biochem Pharmacol*, 62(1), 29-39 (2001)
DOI: 10.1016/S0006-2952(01)00647-5
 155. S. Pierno, A. Liantonio, G. M. Camerino, M. De Bellis, M. Cannone, G. Gramegna, A. Scaramuzzi, S. Simonetti, G. P. Nicchia, D. Basco, M. Svelto, J. F. Desaphy and D. C. Camerino: Potential benefits of taurine in the prevention of skeletal muscle impairment induced by disuse in the hindlimb-unloaded rat. *Amino Acids*, 43(1), 431-45 (2012) DOI: 10.1007/s00726-011-1099-4
 156. A. Stacchiotti, F. Rovetta, M. Ferroni, G. Corsetti, A. Lavazza, G. Sberveglieri and M. F. Aleo: Taurine rescues cisplatin-induced muscle atrophy *in vitro*: a morphological study. *Oxid Med Cell Longev*, 2014, 840951 (2014)
DOI: 10.1155/2014/840951
 157. Y. Yamori, L. Liu, K. Ikeda, A. Miura, S. Mizushima, T. Miki and Y. Nara: Distribution of twenty-four hour urinary taurine excretion and association with ischemic heart disease mortality in 24 populations of 16 countries: results from the WHO-CARDIAC study. *Hypertens Res*, 24(4), 453-7 (2001)
DOI: 10.1291/hypres.24.453
 158. O. P. Wojcik, K. L. Koenig, A. Zeleniuch-Jacquotte, C. Pearte, M. Costa and Y. Chen: Serum taurine and risk of coronary heart disease: a prospective, nested case-control study. *Eur J Nutr*, 52(1), 169-78 (2013)
DOI: 10.1007/s00394-011-0300-6
 159. T. Ito, Y. Kimura, Y. Uozumi, M. Takai, S. Muraoka, T. Matsuda, K. Ueki, M. Yoshiyama, M. Ikawa, M. Okabe, S. W. Schaffer, Y. Fujio and J. Azuma: Taurine depletion caused by knocking out the taurine transporter gene leads to cardiomyopathy with cardiac atrophy. *J Mol Cell Cardiol*, 44(5), 927-37 (2008)
DOI: 10.1016/j.yjmcc.2008.03.001
 160. T. Ito, S. Oishi, M. Takai, Y. Kimura, Y. Uozumi, Y. Fujio, S. W. Schaffer and J. Azuma: Cardiac and skeletal muscle abnormality in taurine transporter-knockout mice. *J Biomed Sci*, 17 Suppl 1, S20 (2010)
DOI: 10.1186/1423-0127-17-S1-S20
 161. E. M. Carneiro, M. Q. Latorraca, E. Araujo, M. Beltra, M. J. Oliveras, M. Navarro, G. Berna, F. J. Bedoya, L. A. Velloso, B. Soria and F. Martin: Taurine supplementation modulates glucose homeostasis and islet function. *J Nutr Biochem*, 20(7), 503-11 (2009)
DOI: 10.1016/j.jnutbio.2008.05.008
 162. N. Wu, Y. Lu, B. He, Y. Zhang, J. Lin, S. Zhao, W. Zhang, Y. Li and P. Han: Taurine prevents free fatty acid-induced hepatic insulin resistance in association with inhibiting JNK1 activation and improving insulin signaling *in vivo*. *Diabetes Res Clin Pract*, 90(3), 288-96 (2010)
DOI: 10.1016/j.diabres.2010.08.020
 163. C. Xiao, A. Giacca and G. F. Lewis: Oral taurine but not N-acetylcysteine ameliorates NEFA-induced impairment in insulin sensitivity and beta cell function in obese and overweight, non-diabetic men. *Diabetologia*, 51(1), 139-46 (2008)
DOI: 10.1007/s00125-007-0859-x
 164. E. Ghigo, S. Goffi, M. Nicolosi, E. Arvat, F. Valente, E. Mazza, M. C. Ghigo and F. Camanni: Growth hormone (GH) responsiveness to combined administration of arginine and GH-releasing hormone does not vary with age in man. *J Clin Endocrinol*

- Metab*, 71(6), 1481-5 (1990)
DOI: 10.1210/jcem-71-6-1481
165. F. Blachier, V. Leclercq-Meyer, J. Marchand, M. C. Woussen-Colle, P. C. Mathias, A. Sener and W. J. Malaisse: Stimulus-secretion coupling of arginine-induced insulin release. Functional response of islets to L-arginine and L-ornithine. *Biochim Biophys Acta*, 1013(2), 144-51 (1989)
DOI: 10.1016/0167-4889(89)90042-6
166. P. J. Reeds: Dispensable and indispensable amino acids for humans. *J Nutr*, 130(7), 1835S-40S (2000)
167. A. Casey and P. L. Greenhaff: Does dietary creatine supplementation play a role in skeletal muscle metabolism and performance? *Am J Clin Nutr*, 72(2 Suppl), 607S-17S (2000)
168. L. Castillo, M. Sanchez, J. Vogt, T. E. Chapman, T. C. DeRoja-Walker, S. R. Tannenbaum, A. M. Ajami and V. R. Young: Plasma arginine, citrulline, and ornithine kinetics in adults, with observations on nitric oxide synthesis. *Am J Physiol*, 268(2 Pt 1), E360-7 (1995)
DOI: 10.1097/00003246-199501001-00445
169. J. K. Stechmiller, B. Childress and L. Cowan: Arginine supplementation and wound healing. *Nutr Clin Pract*, 20(1), 52-61 (2005)
DOI: 10.1177/011542650502000152
170. A. Palloshi, G. Fragasso, P. Piatti, L. D. Monti, E. Setola, G. Valsecchi, E. Galluccio, S. L. Chierchia and A. Margonato: Effect of oral L-arginine on blood pressure and symptoms and endothelial function in patients with systemic hypertension, positive exercise tests, and normal coronary arteries. *Am J Cardiol*, 93(7), 933-5 (2004)
DOI: 10.1016/j.amjcard.2003.12.040
171. H. Kim, E. Barton, N. Muja, S. Yakar, P. Pennisi and D. Leroith: Intact insulin and insulin-like growth factor-I receptor signaling is required for growth hormone effects on skeletal muscle growth and function *in vivo*. *Endocrinology*, 146(4), 1772-9 (2005)
DOI: 10.1210/en.2004-0906
172. J. O. Jorgensen, S. A. Pedersen, L. Thuesen, J. Jorgensen, T. Ingemann-Hansen, N. E. Skakkebaek and J. S. Christiansen: Beneficial effects of growth hormone treatment in GH-deficient adults. *Lancet*, 1(8649), 1221-5 (1989)
DOI: 10.1016/S0140-6736(89)92328-3
173. F. Salomon, R. C. Cuneo, R. Hesp and P. H. Sonksen: The effects of treatment with recombinant human growth hormone on body composition and metabolism in adults with growth hormone deficiency. *N Engl J Med*, 321(26), 1797-803 (1989)
DOI: 10.1056/NEJM198912283212605
174. W. M. Drake, D. Coyte, C. Camacho-Hubner, N. M. Jivanji, G. Kaltsas, D. F. Wood, P. J. Trainer, A. B. Grossman, G. M. Besser and J. P. Monson: Optimizing growth hormone replacement therapy by dose titration in hypopituitary adults. *J Clin Endocrinol Metab*, 83(11), 3913-9 (1998)
DOI: 10.1210/jcem.83.11.5223
175. J. Bryant, E. Loveman, D. Chase, B. Mihaylova, C. Cave, K. Gerard and R. Milne: Clinical effectiveness and cost-effectiveness of growth hormone in adults in relation to impact on quality of life: a systematic review and economic evaluation. *Health Technol Assess*, 6(19), 1-106 (2002)
176. Q. He, X. Kong, G. Wu, P. Ren, H. Tang, F. Hao, R. Huang, T. Li, B. Tan, P. Li, Z. Tang, Y. Yin and Y. Wu: Metabolomic analysis of the response of growing pigs to dietary L-arginine supplementation. *Amino Acids*, 37(1), 199-208 (2009)
DOI: 10.1007/s00726-008-0192-9
177. B. Tan, Y. Yin, Z. Liu, X. Li, H. Xu, X. Kong, R. Huang, W. Tang, I. Shinzato, S. B. Smith and G. Wu: Dietary L-arginine supplementation increases muscle gain and reduces body fat mass in growing-finishing pigs. *Amino Acids*, 37(1), 169-75 (2009)
DOI: 10.1007/s00726-008-0148-0
178. W. Jobgen, C. J. Meininger, S. C. Jobgen, P. Li, M. J. Lee, S. B. Smith, T. E. Spencer, S. K. Fried and G. Wu: Dietary L-arginine supplementation reduces white fat gain and enhances skeletal muscle and brown fat masses in diet-induced obese rats. *J Nutr*, 139(2), 230-7 (2009)
DOI: 10.3945/jn.108.096362
179. K. Yao, Y. L. Yin, W. Chu, Z. Liu, D. Deng, T. Li, R. Huang, J. Zhang, B. Tan, W. Wang and G. Wu: Dietary arginine supplementation increases mTOR signaling activity in skeletal muscle of neonatal pigs. *J Nutr*, 138(5), 867-72 (2008)
180. D. J. Ham, M. K. Caldwell, G. S. Lynch and R.

- Koopman: Arginine protects muscle cells from wasting *in vitro* in an mTORC1-dependent and NO-independent manner. *Amino Acids*, 46(12), 2643-52 (2014)
DOI: 10.1007/s00726-014-1815-y
181. S. R. Collier, D. P. Casey and J. A. Kanaley: Growth hormone responses to varying doses of oral arginine. *Growth Horm IGF Res*, 15(2), 136-9 (2005)
DOI: 10.1016/j.ghir.2004.12.004
182. P. E. May, A. Barber, J. T. D'Olimpio, A. Hourihane and N. N. Abumrad: Reversal of cancer-related wasting using oral supplementation with a combination of beta-hydroxy-beta-methylbutyrate, arginine, and glutamine. *Am J Surg*, 183(4), 471-9 (2002)
DOI: 10.1016/S0002-9610(02)00823-1
183. R. H. Clark, G. Feleke, M. Din, T. Yasmin, G. Singh, F. A. Khan and J. A. Rathmacher: Nutritional treatment for acquired immunodeficiency virus-associated wasting using beta-hydroxy beta-methylbutyrate, glutamine, and arginine: a randomized, double-blind, placebo-controlled study. *JPEN J Parenter Enteral Nutr*, 24(3), 133-9 (2000)
DOI: 10.1177/0148607100024003133
184. P. Flakoll, R. Sharp, S. Baier, D. Levenhagen, C. Carr and S. Nissen: Effect of beta-hydroxy-beta-methylbutyrate, arginine, and lysine supplementation on strength, functionality, body composition, and protein metabolism in elderly women. *Nutrition*, 20(5), 445-51 (2004)
DOI: 10.1016/j.nut.2004.01.009
185. J. P. Lekakis, S. Papathanassiou, T. G. Papaioannou, C. M. Papamichael, N. Zakopoulos, V. Kotsis, A. G. Dagle, K. Stamatelopoulos, A. Protogerou and S. F. Stamatelopoulos: Oral L-arginine improves endothelial dysfunction in patients with essential hypertension. *Int J Cardiol*, 86(2-3), 317-23 (2002)
DOI: 10.1016/S0167-5273(02)00413-8
186. A. Lerman, J. C. Burnett, Jr., S. T. Higano, L. J. McKinley and D. R. Holmes, Jr.: Long-term L-arginine supplementation improves small-vessel coronary endothelial function in humans. *Circulation*, 97(21), 2123-8 (1998)
DOI: 10.1161/01.CIR.97.21.2123
187. P. Clarkson, M. R. Adams, A. J. Powe, A. E. Donald, R. McCredie, J. Robinson, S. N. McCarthy, A. Keech, D. S. Celermajer and J. E. Deanfield: Oral L-arginine improves endothelium-dependent dilation in hypercholesterolemic young adults. *J Clin Invest*, 97(8), 1989-94 (1996)
DOI: 10.1172/JCI118632
188. S. M. Bode-Boger, J. Muke, A. Surdacki, G. Brabant, R. H. Boger and J. C. Frolich: Oral L-arginine improves endothelial function in healthy individuals older than 70 years. *Vasc Med*, 8(2), 77-81 (2003)
DOI: 10.1191/1358863x03vm474oa
189. C. Zhang, T. W. Hein, W. Wang, M. W. Miller, T. W. Fossum, M. M. McDonald, J. D. Humphrey and L. Kuo: Upregulation of vascular arginase in hypertension decreases nitric oxide-mediated dilation of coronary arterioles. *Hypertension*, 44(6), 935-43 (2004)
DOI: 10.1161/01.HYP.0000146907.82869.f2
190. A. Erdely, D. Kepka-Lenhart, R. Salmen-Muniz, R. Chapman, T. Hulderman, M. Kashon, P. P. Simeonova and S. M. Morris, Jr.: Arginase activities and global arginine bioavailability in wild-type and ApoE-deficient mice: responses to high fat and high cholesterol diets. *PLoS One*, 5(12), e15253 (2010)
DOI: 10.1371/journal.pone.0015253
191. S. Elms, F. Chen, Y. Wang, J. Qian, B. Askari, Y. Yu, D. Pandey, J. Iddings, R. B. Caldwell and D. J. Fulton: Insights into the arginine paradox: evidence against the importance of subcellular location of arginase and eNOS. *Am J Physiol Heart Circ Physiol*, 305(5), H651-66 (2013)
DOI: 10.1152/ajpheart.00755.2012
192. D. H. Baker: Tolerance for branched-chain amino acids in experimental animals and humans. *J Nutr*, 135(6 Suppl), 1585S-90S (2005)
193. S. Moehn, P. B. Pencharz and R. O. Ball: Lessons learned regarding symptoms of tryptophan deficiency and excess from animal requirement studies. *J Nutr*, 142(12), 2231S-2235S (2012)
DOI: 10.3945/jn.112.159061
194. A. Morales, N. Arce, M. Cota, L. Buenabad, E. Avelar, J. K. Htoo and M. Cervantes: Effect of dietary excess of branched-chain amino acids on performance and serum concentrations of amino acids in growing pigs. *J Anim Physiol Anim Nutr (Berl)* (2015)
DOI: 10.1111/jpn.12267

195. W. Imamura, R. Yoshimura, M. Takai, J. Yamamura, R. Kanamoto and H. Kato: Adverse effects of excessive leucine intake depend on dietary protein intake: a transcriptomic analysis to identify useful biomarkers. *J Nutr Sci Vitaminol (Tokyo)*, 59(1), 45-55 (2013)
DOI: 10.3177/jnsv.59.45
196. P. B. Pencharz, R. Elango and R. O. Ball: Determination of the tolerable upper intake level of leucine in adult men. *J Nutr*, 142(12), 2220S-2224S (2012)
DOI: 10.3945/jn.112.160259
197. C. Hiratsuka, T. Fukuwatari, M. Sano, K. Saito, S. Sasaki and K. Shibata: Supplementing healthy women with up to 5.0 g/d of L-tryptophan has no adverse effects. *J Nutr*, 143(6), 859-66 (2013)
DOI: 10.3945/jn.112.173823
198. E. M. Cottington, C. LaMantia, S. P. Stabler, R. H. Allen, A. Tangerman, C. Wagner, S. H. Zeisel and S. H. Mudd: Adverse event associated with methionine loading test: a case report. *Arterioscler Thromb Vasc Biol*, 22(6), 1046-50 (2002)
DOI: 10.1161/01.ATV.0000020400.25088.A7
199. J. D. Fernstrom: Effects and side effects associated with the non-nutritional use of tryptophan by humans. *J Nutr*, 142(12), 2236S-2244S (2012)
DOI: 10.3945/jn.111.157065

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