

## p53 and P-glycoprotein influence chemoresistance in hepatocellular carcinoma

ShengLei Li<sup>1</sup>, Ming Gao<sup>2</sup>, ZongMing Li<sup>3</sup>, LiJie Song<sup>2</sup>, XianZheng Gao<sup>1</sup>, Jing Han<sup>1</sup>, Feng Wang<sup>2</sup>, YongFang Chen<sup>1</sup>, WenCai Li<sup>1</sup>, JianPing Yang<sup>1</sup>

<sup>1</sup>Department of Pathology, The First Affiliated Hospital of ZhengZhou University, ZhengZhou, 450000, China, <sup>2</sup>Department of Oncology, The First Affiliated Hospital of ZhengZhou University, ZhengZhou, 450000, China, <sup>3</sup>Department of Interventional Therapy, The First Affiliated Hospital of ZhengZhou University, ZhengZhou, 450000, China

### TABLE OF CONTENTS

1. Abstract
2. Introduction
3. p53 gene in chemoresistance of hepatocellular carcinoma
4. P-gp in chemoresistance of hepatocellular carcinoma
5. Conclusion
6. Acknowledgements
7. References

Ms 12996.

### 1. ABSTRACT

Chemoresistance is a critical obstacle to the treatment of hepatocellular carcinoma (HCC). The mechanisms underlying resistance to doxorubicin, cisplatin, and 5-fluorouracil involve p53 and P-glycoprotein (P-gp). p53 plays a role in cell growth; therefore, resistance mechanisms involve chemotherapy-induced apoptosis and p53 mutation and inactivation. P-gp is an energy-dependent drug efflux pump regulated by p53. Its role in drug resistance has provided new insights into the mechanisms underlying the involvement of p53 and P-gp in chemoresistance and may alter our traditional understanding of p53 and P-gp function. This review outlines the roles and principal mechanisms of p53 and P-gp mediated chemoresistance in HCC.

### 2. INTRODUCTION

Hepatocellular carcinoma (HCC) is the fifth most common cancer and the third leading cause of cancer-related death worldwide (1,2). Chemotherapy is the best option for patients who are ineligible for surgical treatment. However, HCC is often resistant to chemotherapeutic drugs (3), with only a few drugs eliciting a therapeutic effect in more than 20% of patients with HCC (4). The treatment results obtained with chemotherapeutic agents in advanced HCC have been disappointing. Because the successful long-term use of chemotherapy for HCC is often hampered by resistance to chemotherapeutic agents, the reversal of

drug resistance has become a critical issue in HCC therapy.

Drug resistance is a multifactorial phenomenon involving mechanisms such as gene mutation, DNA methylation, alterations in drug metabolism and processing, and changes in the expression or activity of target proteins (5–9). Among these mechanisms, the presence of gene mutations in drug targets is a major cause of acquired resistance in cancer (10). Loss of p53 function, which in a subset of tumors is caused by mutation, is a common feature in human cancers (11). The TP53 mutation is present in almost every type of human cancer, including HCC (11). The multidrug resistance 1 (MDR1) gene product P-glycoprotein (P-gp), an energy-dependent drug efflux pump involved in oncogene activation and tumor aggressiveness, is a predictor of chemoresistance in cancer cells (12–14). P-gp potentially regulates apoptosis, immune cell function, and cellular differentiation, proliferation, and survival (15). The MDR1 gene may be activated during tumor progression in association with mutations in p53 (16). Herein, we discuss the roles of p53 and P-gp in chemoresistance in HCC.

### 3. p53 GENE IN CHEMORESISTANCE IN HCC

The accumulation of mutant p53 with gain of function can result in drug resistance in cancers via direct suppression of apoptotic pathways



**Figure 1.** The p53 mediateds chemoresistance in hepatocellular carcinoma (HCC). In HCC, the action of p53 can be inhibited by dominant-negative p73 (Delta Np73), resulting in resistance to chemotherapy. DeltaNp63alpha can directly interfere with the transcriptional activation function of p53 family target genes, resulting in chemoresistance.

(17,18). In HCC, p53 is associated with resistance to several chemotherapeutic drugs. Cisplatin (cis-diamminedichloroplatinum; CDDP) is a fundamental component of standard treatment regimens for cancers of the respiratory, digestive, and genitourinary systems (19). In systemic monotherapy for HCC, the response rate to CDDP is 15% (20), whereas multidrug regimens containing this agent yield higher response rates. The response rates to arterial infusion regimens containing CDDP range from 41% to 61% (21,22) because they achieve higher concentrations of the drug inside the tumor, thereby exerting a more robust antitumor effect. A previous study suggested that CDDP has a synergistic effect with tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)-induced apoptosis in most HCC cell lines, regardless of p53 status (23). However, an *in vitro* study of the Bel-7402 (p53 wild type), Huh 7 (p53 mutant), and Hep3B (p53 defective) cell lines demonstrated that CDDP reversal of TRAIL resistance in HCC cells is partially dependent on p53 status (24). Thus, the efficiency of CDDP in apoptosis induction is associated with p53. Defective p53 was later shown to contribute to CDDP resistance (25). Downregulation of p53 expression by long interspersed nuclear element-1 ORF-1 protein (LINE-1 ORF-1p) promotes CDDP and epirubicin resistance in HepG2 cells (26). Therefore, decreased expression of p53 promotes CDDP resistance in HCC cells.

Doxorubicin (Adriamycin, ADA) is the cornerstone of chemotherapy for HCC; however, doxorubicin resistance is an obstacle to successful treatment in these patients. Doxorubicin induces apoptosis in human HCC cells via the p53 pathway (27). Mutant p53 promotes doxorubicin resistance in HCC cell lines (28), whereas wild-type p53 increases doxorubicin chemosensitivity in the drug-resistant human HCC cell line Bel7402/5-FU (29). Inhibition of p53 activation reduces sensitivity to doxorubicin (30). The downstream target of p53, N-myc downstream-regulated gene-1, is also involved in doxorubicin resistance in HCC cells (31). In addition to resistance

to doxorubicin, 5-fluorouracil (5-FU) and vincristine resistance in HCC cells is related to the mutation and inactivation of p53 (29,30).

Combination treatment with doxorubicin and sorafenib yields better outcomes than treatment with doxorubicin alone and results in greater median time to progression, overall survival, and progression-free survival in HCC patients (32). These results indicate that sorafenib has a synergistic effect with doxorubicin in the treatment of HCC. The direct target of p53, microRNA-34a, increases the sensitivity of human HCC cells to the antitumor effect of sorafenib, thus potentiating sorafenib-induced apoptosis and toxicity by inhibiting Bcl-2 expression (33).

In HCC cells, p53 mutation and inactivation are key events leading to resistance to CDDP, doxorubicin, 5-FU, and other agents. The p53 mutant p53 (G245D) is associated with resistance to histone deacetylase inhibitors, which decreases the chemosensitivity of HCC cells (34). On the contrary, p53 activity can be inhibited by dominant-negative p73 (Delta Np73), which results in resistance to chemotherapy (35). DeltaNp63alpha directly interferes with the transcriptional activation function of p53 family target genes, which results in chemoresistance (Figure 1) (36). Because chemotherapy-induced apoptosis involving p53 in HCC is mediated by extrinsic and intrinsic pathways (37), the mechanism underlying p53-related chemoresistance may be initiated or regulated by both extrinsic and intrinsic factors.

#### 4. P-GP IN CHEMORESISTANCE IN HCC

Although numerous mechanisms underlying chemoresistance have been described, a large body of evidence strongly supports the involvement of energy-dependent efflux systems (e.g., P-gp) that pump anticancer agents out of cells (38). P-gp, which is encoded by the MDR1 gene, is a 170-kDa protein belonging to the ATP-binding cassette superfamily of membrane transporter proteins (39,40). It reportedly

operates as an ATP-powered drug efflux pump. Several studies have provided functional insight into the role of P-gp in HCC chemotherapy. P-gp is highly expressed in the HCC QGY-TR 50 cell line, which is resistant to actinomycin D, doxorubicin, vinblastine, and vincristine (41). P-gp is also involved in the chemoresistance of human HCC cells to other anticancer drugs such as taxol, 5-FU, and methotrexate (42,43). On the contrary, a P-gp interacting agent has shown anticancer activity in HCC (44).

P-gp overexpression is also associated with doxorubicin resistance in tumor tissues (45–50). In an *in vivo* study, tumor tissues from nude mice implanted with the doxorubicin-resistant HepG2 cell line showed increased P-gp expression (51). Inhibition of P-gp expression increases the cytotoxicity of doxorubicin in the human HCC cell line HepG2 and its drug-resistant subline R-HepG2 (52). Compared with HepG2 cells, R-HepG2 cells show decreased intracellular accumulation of doxorubicin and increased P-gp expression (53). Among the proposed mechanisms of doxorubicin resistance is the mitogen-activated protein kinase-dependent upregulation of P-gp (53). Moreover, the multikinase inhibitor sorafenib tosylate inhibits the expression of P-gp (53). Because the non-steroidal anti-inflammatory drug indomethacin and the cyclooxygenase-2-selective inhibitor SC236 have been found to enhance doxorubicin cytotoxicity in HepG2 and R-HepG2 cells by reversing the upregulation of P-gp expression (52), the COX-2 gene has been proposed to participate in multidrug resistance in HCC by regulating P-gp (54). These studies demonstrate that P-gp promotes doxorubicin resistance in HCC; however, the underlying mechanism remains to be elucidated.

Increased P-gp expression is associated with CDDP resistance in the HepG2 cell line (55), and the CDDP- and doxorubicin-resistant Bel-7402 cell line shows significantly increased P-gp expression (56). However, in the HCC cell line QGY/CDDP, which is established via stepwise increases in CDDP exposure, the mechanism underlying resistance may not be associated with P-gp expression (57). These conflicting findings regarding the resistance of HCC cells to CDDP may be associated with the types of cells studied. Although these *in vitro* findings have not been verified by *in vivo* data, they may provide key contributions to our understanding of the role of P-gp in CDDP resistance in human HCC.

The response to 5-FU treatment can be modulated by CDDP. In 5-FU-resistant BEL-7402/5-FU cells, the inhibition of P-gp function contributes to the reversal of chemoresistance (58,59). However, a recent study has shown that, unlike the roles of multidrug resistance protein 1, Bcl-xl, TS, and E-cad, the role of P-gp in drug resistance may be limited in

BEL-7402/5-FU cells (60). These *in vitro* studies used human cell lines, and *in vivo* studies in humans may provide more definite findings.

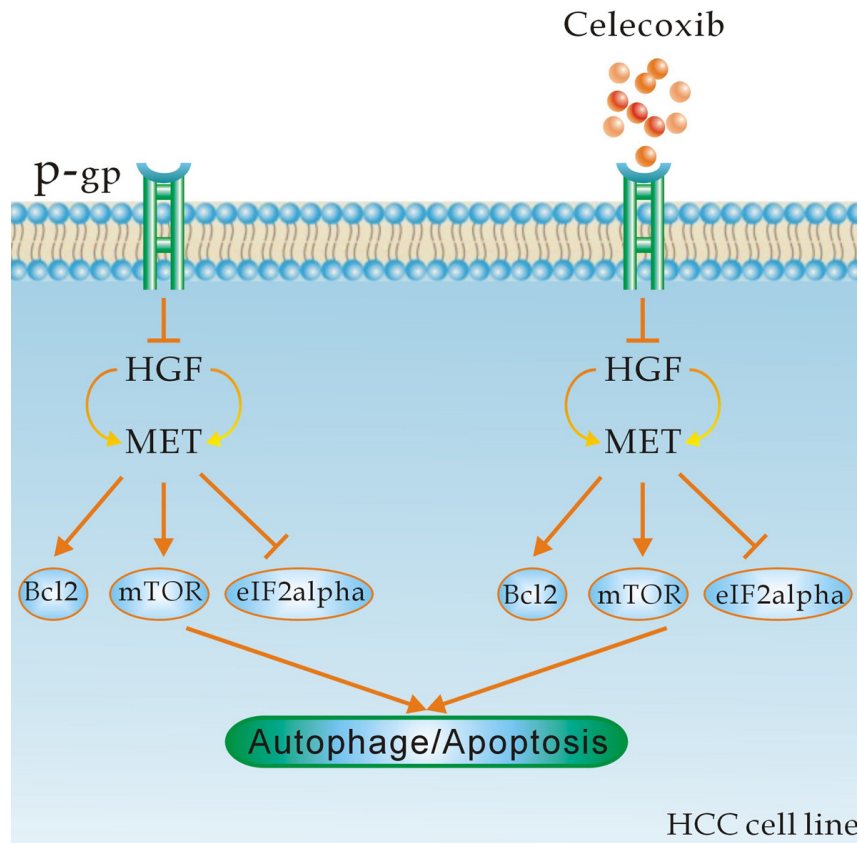
P-gp appears to play a key role in HCC chemoresistance. The mechanism underlying this resistance in multidrug-resistant overexpressing human HCC cell lines involves P-gp mediation of celecoxib-induced cell-cycle arrest and autophagy via downregulation of the HGF/MET autocrine loop and Bcl-2 expression (Figure 2) (61). Moreover, the CDDP-resistant HCC cell line SK-Hep1 shows increased mitochondrial translocation and functional activation of P-gp (62).

In a study examining the role of P-gp in chemoresistance, Feng *et al.* (63) showed that LINE-1 ORF-1p upregulates P-gp gene expression, which results in increased sensitivity of HepG2 cells to CDDP and epirubicin. A common mechanism involving increased P-gp ATPase activity in doxorubicin, CDDP, and 5-FU resistance has been described (64). Thus, despite the limited role of P-gp in CDDP and 5-FU resistance suggested by *in vitro* studies, the involvement of P-gp in HCC chemotherapy with these drugs warrants further investigation *in vivo*.

## 5. CONCLUSION

In osteosarcoma and colon carcinoma, mutant p53 activates MDR-1 promoter activity and wild-type p53 represses this activity (65). In HCC, p53 has a similar regulatory effect on P-gp. Although an association between p53 and P-gp expression in advanced HCC was not detected in studies of tumor samples from HCC patients (50), P-gp expression was found to be modulated by downregulation of p53 gene expression (66). Furthermore, transfection of wild-type p53 into Bel-7402 cells resulted in significant downregulation of P-gp expression and increased vincristine chemosensitivity in these cells (67). These findings further demonstrate that p53 mutation, but not p53 protein expression, regulates P-gp expression.

The p53 gene functions in cell cycle control and apoptosis. The involvement of p53 in resistance to CDDP, doxorubicin, and 5-FU among others is mediated by chemotherapy-induced apoptosis and p53 mutation and inactivation. These mechanisms are associated with the role of p53 in cell growth. However, elucidating the role of P-gp, an energy-dependent drug efflux pump regulated by p53, in the resistance to these drugs may provide new insight into chemoresistance mechanisms involving p53 and P-gp and alter our traditional understanding of p53 and P-gp function therein. Therefore, future studies should focus on exploring the mechanisms underlying the regulation of P-gp by p53, which could provide critical information for the development of treatment strategies aimed at



**Figure 2.** P-glycoprotein (P-gp) can mediate cell-cycle arrest and autophagy in multidrug-resistant hepatocellular carcinoma (HCC). The P-gp can mediate cell-cycle arrest and autophagy induced by celecoxib in human multidrug-resistant overexpressing HCC cell line by down-regulation of the HGF/MET autocrine loop and Bcl-2 expression.

reducing chemoresistance and improving the survival of cancer patients.

## 6. ACKNOWLEDGEMENT

This work was supported by the National Science Foundation of China (NSFC NO. 81372677). Henan Province foundation and front engineering research project(NO. 132300410073)

## 7. REFERENCES

1. Jemal A, Thomas A, Murray T, Thun M: Cancer statistics, 2002. *CA Cancer J Clin* 52,23-47 (2002)  
DOI: 10.3322/canjclin.52.1.23
2. Parkin DM, Bray F, Ferlay J, Pisani P: Global cancer statistics, 2002. *CA Cancer J Clin* 55,74-108 (2005)  
DOI: 10.3322/canjclin.55.2.74
3. Bruix J, Sherman M, Llovet JM: Clinical management of hepatocellular carcinoma. Conclusions of the Barcelona-EASL conference. European Association for the Study of the Liver. *J Hepatol* 10,421-430 (2001)  
DOI: 10.1016/S0168-8278(01)00130-1
4. Nowak AK, Chow PK, Findlay M: Systemic therapy for advanced hepatocellular carcinoma: a review. *Eur J Cancer* 10,1474-1484 (2004)  
DOI: 10.1016/j.ejca.2004.02.027
5. Zheng T, Wang J, Chen X, Liu L: Role of microRNA in anticancer drug resistance. *Int J Cancer* 126, 2-10 (1995)  
DOI: 10.1002/ijc.24782
6. Harrison DJ: Molecular mechanisms of drug resistance in tumours. *J Pathol* 175, 7-12 (2010)  
DOI: 10.1002/path.1711750103
7. Ma J, Dong C, Ji C: MicroRNA and drug resistance. *Cancer Gene Ther* 17, 523-531 (2010)  
DOI: 10.1038/cgt.2010.18

8. Roberti A, La Sala D, Cinti C: Multiple genetic and epigenetic interacting mechanisms contribute to clonally selection of drug-resistant tumors: Current views and new therapeutic prospective. *J Cell Physiol* 207, 571-581 (2006)  
DOI: 10.1002/jcp.20515
9. Allen KE, Weiss GJ: Resistance may not be futile: MicroRNA biomarkers for chemoresistance and potential therapeutics. *Mol Cancer Ther* 9, 3126-3136 (2010)  
DOI: 10.1158/1535-7163.MCT-10-0397
10. Kumar R, Chaudhary K, Gupta S, Singh H, Kumar S, Gautam A, Kapoor P, Raghava GP: CancerDR: cancer drug resistance database. *Sci Rep* 3, 1445 (2013)  
DOI: 10.1038/srep01445
11. Muller PA, Vousden KH: p53 mutations in cancer. *Nat Cell Biol* 15(1), 2-8 (2013)  
DOI: 10.1038/ncb2641
12. Pinedo HM, Giaccone G: P-glycoprotein-a marker of cancer-cell behavior. *N Engl J Med*, 333, 1417-9 (1995)  
DOI: 10.1056/NEJM199511233332111
13. Ralhan R, Narayan M, Salotra P, Shukla NK, Chauhan SS: Evaluation of P-glycoprotein expression in human oral oncogenesis: correlation with clinicopathological features. *Int J Cancer* 72, 728-34 (1997)  
DOI: 10.1002/(SICI)1097-0215(19970904)72:5<728::AID-IJC4>3.0.CO;2-U
14. Yakirevich E, Sabo E, Naroditsky I, Sova Y, Lavie O, Resnick MB: Multidrug resistance-related phenotype and apoptosis-related protein expression in ovarian serous carcinomas. *Gynecol Oncol* 100, 152-9 (2006)  
DOI: 10.1016/j.ygyno.2005.08.050
15. Johnstone RW, Ruefli AA, Tainton KM, Smyth MJ: A role for P-glycoprotein in regulating cell death. *Leuk Lymphoma* 38, 1-11 (2000)
16. Chin KV, Ueda K, Pastan I, Gottesman MM: Modulation of activity of the promoter of the human MDR1 gene by Ras and p53. *Science* 255, 459-62 (1992)  
DOI: 10.1126/science.1346476
17. Lowe SW, Ruley HE, Jacks T, Housman DE: p53-dependent apoptosis modulates the cytotoxicity of anticancer agents. *Cell* 74(6), 957-67 (1993)  
DOI: 10.1016/0092-8674(93)90719-7
18. Kamesaki S, Kamesaki H, Jorgensen TJ, Tanizawa A, Pommier Y, Cossman J: bcl-2 protein inhibits etoposide-induced apoptosis through its effects on events subsequent to topoisomerase II-induced DNA strand breaks and their repair. *Cancer Res* 53(18), 4251-6 (1993)
19. Go RS, Adjei AA: Review of the comparative pharmacology and clinical activity of cisplatin and carboplatin. *J Clin Oncol* 17, 409-422 (1999)
20. Okada S, Okazaki N, Nose H, Shimada Y, Yoshimori M, Aoki K: A phase 2 study of cisplatin in patients with hepatocellular carcinoma. *Oncology* 50, 22-26 (1993)  
DOI: 10.1159/000227142
21. Ikeda M, Maeda S, Shibata J, Muta R, Ashihara H, Tanaka M, Fujiyama S, Tomita K: Transcatheter arterial chemotherapy with and without embolization in patients with hepatocellular carcinoma. *Oncology* 66, 24-31 (2004)  
DOI: 10.1159/000076331
22. Ikeda M, Maeda S, Ashihara H, Nagahama H, Tanaka M, Sasaki Y: Transcatheter arterial infusion chemotherapy with cisplatin-lipiodol suspension in patients with hepatocellular carcinoma. *J Gastroenterol* 45(1), 60-7 (2010)  
DOI: 10.1007/s00535-009-0109-8
23. Shin EC, Seong YR, Kim CH, Kim H, Ahn YS, Kim K, Kim SJ, Hong SS, Park JH: Human hepatocellular carcinoma cells resist to TRAIL-induced apoptosis, and the resistance is abolished by cisplatin. *Exp Mol Med* 34(2), 114-22 (2002)  
DOI: 10.1038/emmm.2002.17
24. Zhang B, Shan H, Li D, Li ZR, Zhu KS, Jiang ZB, Huang MS: Cisplatin sensitizes human hepatocellular carcinoma cells, but not hepatocytes and mesenchymal stem cells, to TRAIL within a therapeutic window partially depending on the upregulation of DR5. *Oncol Rep* 25(2), 461-8 (2011)
25. Kim Y, Jang M, Lim S, Won H, Yoon KS, Park JH, Kim HJ, Kim BH, Park WS, Ha J, Kim SS: Role of cyclophilin B in tumorigenesis and cisplatin resistance in hepatocellular carcinoma in humans. *Hepatology* 54(5), 1661-78 (2011)  
DOI: 10.1002/hep.24539

26. Feng F, Lu YY, Zhang F, Gao XD, Zhang CF, Meredith A, Xu ZX, Yang YT, Chang XJ, Wang H, Qu JH, Zeng Z, Yang JL, Wang CP, Zhu YF, Cui JJ, Yang YP: Long interspersed nuclear element ORF-1 protein promotes proliferation and resistance to chemotherapy in hepatocellular carcinoma. *World J Gastroenterol* 19(7),1068-78 (2013)  
DOI: 10.3748/wjg.v19.i7.1068
27. Zheng T, Wang J, Song X, Meng X, Pan S, Jiang H, Liu L: Nutlin-3 cooperates with doxorubicin to induce apoptosis of human hepatocellular carcinoma cells through p53 or p73 signaling pathways. *J Cancer Res Clin Oncol* 136(10),1597-604 (2010)  
DOI: 10.1007/s00432-010-0817-8
28. Chan KT, Lung ML: Mutant p53 expression enhances drug resistance in a hepatocellular carcinoma cell line. *Cancer Chemother Pharmacol* 53(6),519-26 (2004)  
DOI: 10.1007/s00280-004-0767-4
29. Li YX, Lin ZB, Tan HR: Wild type p53 increased chemosensitivity of drug-resistant human hepatocellular carcinoma Bel7402/5-FU cells. *Acta Pharmacol Sin* 25(1),76-82 (2004)
30. Fang F, Yang L, Tao Y, Qin W: FBI-1 promotes cell proliferation and enhances resistance to chemotherapy of hepatocellular carcinoma *in vitro* and *in vivo*. *Cancer* 118(1),134-46 (2012)  
DOI: 10.1002/cncr.26251
31. Jung EU, Yoon JH, Lee YJ, Lee JH, Kim BH, Yu SJ, Myung SJ, Kim YJ, Lee HS: Hypoxia and retinoic acid-inducible NDRG1 expression is responsible for doxorubicin and retinoic acid resistance in hepatocellular carcinoma cells. *Cancer Lett* 298(1),9-15 (2010)  
DOI: 10.1016/j.canlet.2010.05.020
32. Abou-Alfa GK, Johnson P, Knox JJ, Capanu M, Davidenko I, Lacava J, Leung T, Gansukh B, Saltz LB: Doxorubicin plus sorafenib vs doxorubicin alone in patients with advanced hepatocellular carcinoma: a randomized trial. *JAMA* 304(19),2154-60 (2010)  
DOI: 10.1001/jama.2010.1672
33. Yang F, Li QJ, Gong ZB, Zhou L, You N, Wang S, Li XL, Li JJ, An JZ, Wang DS, He Y, Dou KF: MicroRNA-34a Targets Bcl-2 and Sensitizes Human Hepatocellular Carcinoma Cells to Sorafenib Treatment. *Technol Cancer Res Treat.* (Epub ahead of print) (2013)  
DOI: 10.7785/tcrt.2012.500364
34. Zhang CZ, Chen GG, Merchant JL, Lai PB: Interaction between ZBP-89 and p53 mutants and its contribution to effects of HDACi on hepatocellular carcinoma. *Cell Cycle* 11(2),322-34 (2012)  
DOI: 10.4161/cc.11.2.18758
35. Müller M, Schilling T, Sayan AE, Kairat A, Lorenz K, Schulze-Bergkamen H, Oren M, Koch A, Tannapel A, Stremmel W, Melino G, Krammer PH: TAp73/Delta Np73 influences apoptotic response, chemosensitivity and prognosis in hepatocellular carcinoma. *Cell Death Differ* 12(12),1564-77 (2005)  
DOI: 10.1038/sj.cdd.4401774
36. Mundt HM, Stremmel W, Melino G, Krammer PH, Schilling T, Müller M: Dominant negative (DeltaN) p63alpha induces drug resistance in hepatocellular carcinoma by interference with apoptosis signaling pathways. *Biochem Biophys Res Commun* 396(2),335-41 (2010)  
DOI: 10.1016/j.bbrc.2010.04.093
37. Seitz SJ, Schleithoff ES, Koch A, Schuster A, Teufel A, Staib F, Stremmel W, Melino G, Krammer PH, Schilling T, Müller M: Chemotherapy-induced apoptosis in hepatocellular carcinoma involves the p53 family and is mediated via the extrinsic and the intrinsic pathway. *Int J Cancer* 126(9),2049-66 (2010)
38. Gottesman MM, Pastan I: Biochemistry of multidrug resistance mediated by the multidrug transporter. *Annu Rev Biochem* 62,385-427 (1993)  
DOI: 10.1146/annurev.bi.62.070193.002125
39. Ambudkar SV, Dey S, Hrycyna CA, Ramachandra M, Pastan I, Gottesman MM: Biochemical, cellular, and pharmacological aspects of the multidrug transporter. *Annu Rev Pharmacol Toxicol* 39,361-98 (1999)  
DOI: 10.1146/annurev.pharmtox.39.1.361
40. Ling V, Kartner N, Sudo T, Siminovitch L, Riordan JR: Multidrug-resistance phenotype in Chinese hamster ovary cells. *Cancer Treat Rep* 67(10),869-74 (1983)
41. Zhou J, Cheng SC, Luo D, Xie Y: Study of multi-drug resistant mechanisms in a taxol-resistant hepatocellular carcinoma QGY-TR

- 50 cell line. *Biochem Biophys Res Commun* 280(5),1237-42 (2001)  
DOI: 10.1006/bbrc.2001.4268
42. Cheng SC, Zhou J, Xie Y: P-glycoprotein expression induced by glucose depletion enhanced the chemosensitivity in human hepatocellular carcinoma cell-lines. *Cell Biol Int* 29(4),269-75 (2005)  
DOI: 10.1016/j.cellbi.2004.12.010
  43. Meena AS, Sharma A, Kumari R, Mohammad N, Singh SV, Bhat MK: Inherent and acquired resistance to paclitaxel in hepatocellular carcinoma: molecular events involved. *PLoS One* 8(4),e61524 (2013)  
DOI: 10.1371/journal.pone.0061524
  44. Kuo TC, Chiang PC, Yu CC, Nakagawa-Goto K, Bastow KF, Lee KH, Guh JH: A unique P-glycoprotein interacting agent displays anticancer activity against hepatocellular carcinoma through inhibition of GRP78 and mTOR pathways. *Biochem Pharmacol* 81(9),1136-44 (2011)  
DOI: 10.1016/j.bcp.2011.02.013
  45. Ishimura H: Chemosensitivity test on hepatocellular carcinoma (HCC) and drug resistance. *Hokkaido Igaku Zasshi* 71(6),689-98 (1996)
  46. Huang M, Liu G: The study of innate drug resistance of human hepatocellular carcinoma Bel7402 cell line. *Cancer Lett* 135(1),97-105 (1999)  
DOI: 10.1016/S0304-3835(98)00280-8
  47. Nakajima A, Yamamoto Y, Taura K, Hata K, Fukumoto M, Uchinami H, Yonezawa K, Yamaoka Y: Beneficial effect of cepharanthine on overcoming drug-resistance of hepatocellular carcinoma. *Int J Oncol* 24(3),635-45 (2004)
  48. Zhai BJ, Wu F, Shao ZY, Hu K, Wang ZB: Establishment of human multidrug-resistant hepatocellular carcinoma cell line (HepG2/Adm) and biological characteristics evaluation. *Ai Zheng* 23(4),391-5 (2004)
  49. Yan F, Wang XM, Liu ZC, Pan C, Yuan SB, Ma QM: JNK1, JNK2, and JNK3 are involved in P-glycoprotein-mediated multidrug resistance of hepatocellular carcinoma cells. *Hepatobiliary Pancreat Dis Int* 9(3),287-95 (2010)
  50. Chou YY, Cheng AL, Hsu HC: Expression of P-glycoprotein and p53 in advanced hepatocellular carcinoma treated by single agent chemotherapy: clinical correlation. *J Gastroenterol Hepatol* 12(8),569-75 (1997)  
DOI: 10.1111/j.1440-1746.1997.tb00487.x
  51. Ding L, Chen XP, Zhang ZW, Jing K, Zhang WG: Human multi-drug resistant hepatocellular carcinoma induced in nude mice by B-ultrasonographically-directed orthotopic implantation: a new experimental model. *Hepatobiliary Pancreat Dis Int* 6(4),393-8 (2007)
  52. Ye CG, Wu WK, Yeung JH, Li HT, Li ZJ, Wong CC, Ren SX, Zhang L, Fung KP, Cho CH: Indomethacin and SC236 enhance the cytotoxicity of doxorubicin in human hepatocellular carcinoma cells via inhibiting P-glycoprotein and MRP1 expression. *Cancer Lett* 304(2),90-6 (2011)  
DOI: 10.1016/j.canlet.2011.01.025
  53. Ye CG, Yeung JH, Huang GL, Cui P, Wang J, Zou Y, Zhang XN, He ZW, Cho CH: Increased glutathione and mitogen-activated protein kinase phosphorylation are involved in the induction of doxorubicin resistance in hepatocellular carcinoma cells. *Hepatol Res* 43(3),289-99 (2013)  
DOI: 10.1111/j.1872-034X.2012.01067.x
  54. Li B, Liu Y, Su S, Zhang MY, Yuan Q, Chen C, Xia XM, Liu CA, Gong JP: Expressions and significance of COX-2 and P-gp in human hepatocellular carcinoma tissues. *Zhonghua Gan Zang Bing Za Zhi* 19(10),755-8 (2011)
  55. Fang D, Guo Y, Zhu Z, Chen W: Silence of p15 expression by RNAi enhances cisplatin resistance in hepatocellular carcinoma cells. *Bosn J Basic Med Sci* 12(1),4-9 (2012)
  56. Zhong X, Xiong M, Meng X, Gong R: Comparison of the multi-drug resistant human hepatocellular carcinoma cell line Bel-7402/ADM model established by three methods. *J Exp Clin Cancer Res* 29,115 (2010)  
DOI: 10.1186/1756-9966-29-115
  57. Yang JX, Luo Y, Qiu HM, Tang WX: Characterization and resistance mechanisms of cisplatin-resistant human hepatocellular carcinoma cell line. *Saudi Med J* 30(1),35-40 (2009)
  58. Shi LX, Ma R, Lu R, Xu Q, Zhu ZF, Wang L, Zhou CL, Li XL, Zhang HL, Yao Z: Reversal effect of tyroservatide (YSV) tripeptide on multi-drug resistance in resistant human

- hepatocellular carcinoma cell line BEL-7402/5-FU. *Cancer Lett* 269(1),101-10 (2008)  
DOI: 10.1016/j.canlet.2008.04.033
59. Lu F, Hou YQ, Song Y, Yuan ZJ: TFPI-2 downregulates multidrug resistance protein in 5-FU-resistant human hepatocellular carcinoma BEL-7402/5-FU cells. *Anat Rec (Hoboken)* 296(1),56-63 (2013)  
DOI: 10.1002/ar.22611
60. Gu W, Fang FF, Li B, Cheng BB, Ling CQ: Characterization and resistance mechanisms of a 5-fluorouracil-resistant hepatocellular carcinoma cell line. *Asian Pac J Cancer Prev* 13(9),4807-14 (2012)  
DOI: 10.7314/APJCP.2012.13.9.4807
61. Mazzanti R, Platini F, Bottini C, Fantappiè O, Solazzo M, Tessitore L: Down-regulation of the HGF/MET autocrine loop induced by celecoxib and mediated by P-gp in MDR-positive human hepatocellular carcinoma cell line. *Biochem Pharmacol* 78(1),21-32 (2009)  
DOI: 10.1016/j.bcp.2009.03.013
62. Ling X, Zhou Y, Li SW, Yan B, Wen L: Modulation of mitochondrial permeability transition pore affects multidrug resistance in human hepatocellular carcinoma cells. *Int J Biol Sci* 6(7),773-83 (2010)  
DOI: 10.7150/ijbs.6.773
63. Feng F, Lu YY, Zhang F, Gao XD, Zhang CF, Meredith A, Xu ZX, Yang YT, Chang XJ, Wang H, Qu JH, Zeng Z, Yang JL, Wang CP, Zhu YF, Cui JJ, Yang YP: Long interspersed nuclear element ORF-1 protein promotes proliferation and resistance to chemotherapy in hepatocellular carcinoma. *World J Gastroenterol* 19(7),1068-78 (2013)  
DOI: 10.3748/wjg.v19.i7.1068
64. Wang CF, Wang YQ, Huang FZ, Nie WP, Liu XY, Jiang XZ: Association between reversal of multidrug resistance by methyl jasmonate and P-glycoprotein ATPase activity in hepatocellular carcinoma. *J Int Med Res* 41(4),964-74 (2013)  
DOI: 10.1177/0300060513483401
65. Sampath J, Sun D, Kidd VJ, Grenet J, Gandhi A, Shapiro LH, Wang Q, Zambetti GP, Schuetz JD: Mutant p53 cooperates with ETS and selectively up-regulates human MDR1 not MRP1. *J Biol Chem* 276, 39359-67(2001)  
DOI: 10.1074/jbc.M103429200
66. Takeba Y, Sekine S, Kumai T, Matsumoto N, Nakaya S, Tsuzuki Y, Yanagida Y, Nakano H, Asakura T, Ohtsubo T, Kobayashi S: Irinotecan-induced apoptosis is inhibited by increased P-glycoprotein expression and decreased p53 in human hepatocellular carcinoma cells. *Biol Pharm Bull* 30(8),1400-6 (2007)  
DOI: 10.1248/bpb.30.1400
67. Gai XD, Li GL, Huang JZ, Xue HJ, Wang D: Reversal of multidrug resistance of human hepatocellular carcinoma cells by wild-type p53 gene and related mechanisms. *Ai Zheng* 25(8),954-9 (2006)

**Abbreviations:** HCC, Hepatocellular carcinoma; P-gp, P-glycoprotein; MDR1, multidrug resistance 1; TRAIL, TNF-related apoptosis-inducing ligand; 5-FU, 5-fluorouracil; ABC, ATP-binding cassette

**Key Words:** p53, P-glycoprotein, Chemoresistance, Hepatocellular Carcinoma, Review

**Send correspondence to:** JianPing Yang, Department of Pathology, The First Affiliated Hospital of ZhengZhou University, Zheng Zhou, 450000, China, Tel: 86-13598423254, Fax: 86-0371-66913114, E-mail: yjping1111@yeah.net