

## Molecular diagnostics in gastric cancer

Jan Bornschein<sup>1,2</sup>, Marcis Leja<sup>3</sup>, Juozas Kupcinskas<sup>4</sup>, Alexander Link<sup>1</sup>, Jamie Weaver<sup>2</sup>, Massimo Rugge<sup>5</sup>, Peter Malferttheiner<sup>1</sup>

<sup>1</sup>Dept. of Gastroenterology, Hepatology and Infectious Diseases, Otto-von-Guericke University, Magdeburg, Germany, <sup>2</sup>MRC Cancer Cell Unit, Hutchison-MRC Research Centre, Cambridge, UK, <sup>3</sup>Faculty of Medicine, University of Latvia, Riga, Latvia, <sup>4</sup>Department of Gastroenterology, Lithuanian University of Health Sciences, Kaunas, Lithuania, <sup>5</sup>Department of Medicine - DIMED, Surgical Pathology and Cytopathology Unit, University of Padova, Padova, Italy

### TABLE OF CONTENTS

1. Abstract
2. Introduction and epidemiological background
3. Developments in endoscopy
4. Histological assessment
  - 4.1. Histopathology of gastritis and attributable risk
  - 4.2. SPEM - spasmodic polypeptide expressing metaplasia
  - 4.3. Subtyping of mucins
  - 4.4. Expression of Her2
5. Serological biomarkers
  - 5.1. Pepsinogens and the "serological biopsy"
  - 5.2. Cancer autoantibodies
  - 5.3. Circulating cancer cells
6. Volatile organic components in the breath
7. Molecular diagnostics
  - 7.1. MicroRNA
  - 7.2. Epigenetic changes
  - 7.3. Genetic alterations
8. Biostatistical assessment
9. View to the future and conclusion
10. Acknowledgements
11. References

## 1. ABSTRACT

Despite recent advances in individualised targeted therapy, gastric cancer remains one of the most challenging diseases in gastrointestinal oncology. Modern imaging techniques using endoscopic filter devices and *in vivo* molecular imaging are designed to enable early detection of the cancer and surveillance of patients at risk. Molecular characterisation of the tumour itself as well as of the surrounding inflammatory environment is more sophisticated in the view of tailored therapies and individual prognostic assessment. The broad application of high throughput techniques for the description of genome wide patterns of structural (copy number aberrations, single nucleotide polymorphisms, methylation pattern) and functional (gene expression profiling, proteomics, miRNA) alterations in the cancer tissue lead not only to a better understanding of the tumour biology but also to a description of gastric cancer subtypes independent from classical stratification systems. Biostatistical means are required for the interpretation of the massive amount of data generated by these approaches. In this review we give an overview on the current knowledge of diagnostic methods for detection, description and understanding of gastric cancer disease.

## 2. INTRODUCTION AND EPIDEMIOLOGICAL BACKGROUND

Taken into consideration the predicted growth of the world population and the increase of average life expectancies in many countries, the absolute number of gastric cancer cases is likely to be stable or will even increase in the future despite a declining incidence (1). Almost one million cases are newly diagnosed each year, and about 740,000 deaths are caused by this disease resulting in 8% of all cancer cases and 10% of all cancer-related deaths annually (2). There is a regional variation of the incidence rates by the factor 10, and more than 70% of gastric cancers occur in developing countries due to higher *H. pylori* prevalence rates (2).

In contrast to a decline in the incidence of distal gastric cancer there has been an increase of adenocarcinomas at the oesophagogastric junction including gastric cardia cancer, mainly in North America and Europe(3-6). In Asia, distal gastric cancer still remains the main entity.

At the time of diagnosis of gastric cancer, there is usually only a short period of symptoms such as

## Modern approaches in gastric cancer assessment

unintentional weight loss, anaemia, epigastric pain, nausea and vomiting, or dyspeptic symptoms. About 40% patients don't complain about any dyspeptic symptoms at any time (7). Therefore, diagnosis for the majority of patients is made at an advanced stage when only limited treatment options can be offered. Even though there has been some improvement in the clinical management of gastric cancer, the 5 year survival rate is <30% in most countries and the reported mortality rates mirror the incidence of the disease. Therefore, tools to enable an early diagnose are of critical importance.

### 3. DEVELOPMENTS IN ENDOSCOPY

In the absence of appropriate blood-based diagnostic tests for gastric cancer, upper gastrointestinal endoscopy with biopsy sampling for histopathological evaluation remains the gold-standard for determination of a definite diagnosis. The diagnostic yield of traditional white light endoscopy (WLE) can be enhanced by chromoendoscopic techniques (e.g. staining with methylene blue or indigo carmine) or application of acidic dye (8, 9).

Technical advances that allow virtual chromoendoscopy by the use of optical filter systems in combination with optical magnification of the mucosal surface pattern have nearly replaced the classical application of intraluminal dye (10).

Narrow band imaging (NBI) uses special wavelength filters for red coloured structures (e.g. blood vessels or inflamed areas) by which the detection of even early neoplastic lesions in the stomach can be dramatically increased compared to traditional WLE resulting in sensitivity and specificity above 90% (11-14).

The evaluation of the microvascular pattern as well as of the superficial structures allows the differentiation of small elevated lesions between adenomatous and cancerous tissue formations with higher accuracy, sensitivity and specificity for NBI compared to WLE(14-16). This approach can also be used for surveillance after endoscopic resection to detect residual or recurrent disease(17).

The mucosal pattern in magnifying NBI-enhanced endoscopy can give some indication for the depth of invasion in case of a malignant lesion (18). By evaluation of surface and microvessel structure it has even been possible to distinguish between Sm1 (cancers with intramucosal and minute submucosal invasion <500 µm in depth) and Sm2 (deeper submucosal invasion ≥500 µm in depth) early gastric cancer, (19). Similar criteria have been applied as surrogate parameters for the degree of differentiation of an early gastric cancer (20).

During the past years there has been strong effort to develop a general classification system for NBI-documented characteristics mainly focusing on the surface appearance including the gastric pit pattern, the microvascular structure as well as colour and shape of the respective lesion (21). The diagnostic quality of these

patterns were satisfying although interobserver variation was still high (13, 22). Compared with adenomas, carcinomas present at bigger size, depressed morphology, red colour and positive findings (irregularities) in surface and vessel structure (23-25).

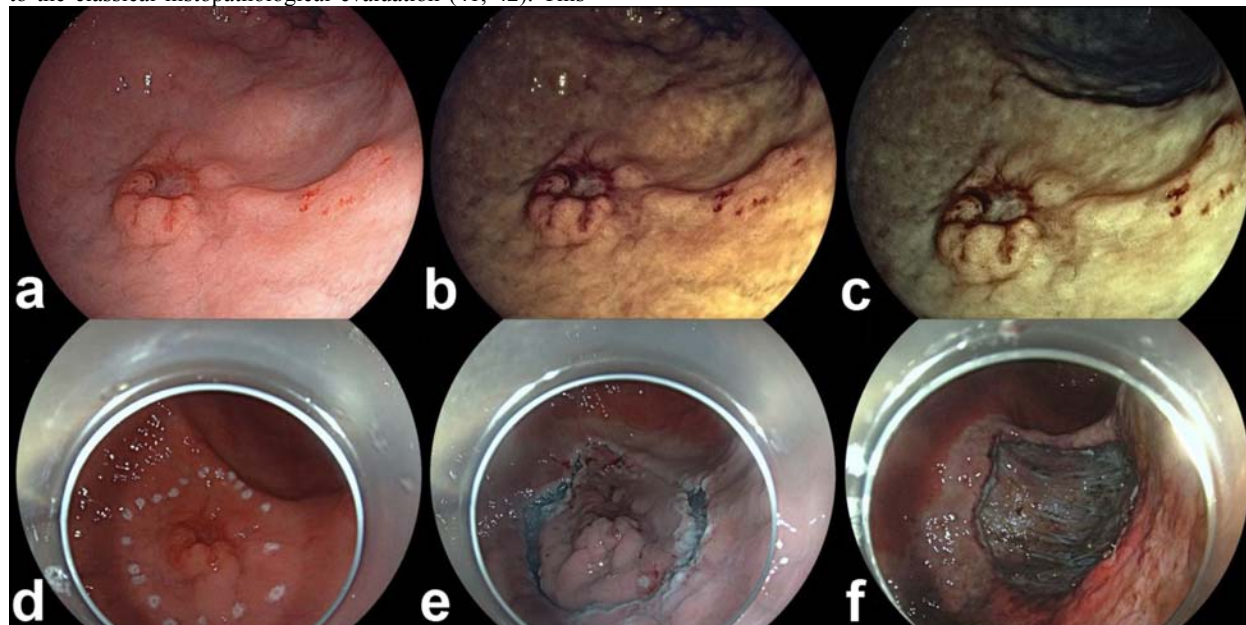
One approach to improve the diagnostic quality of NBI-based endoscopic imaging is the combination with other filter systems. Autofluorescence imaging (AFI) has been introduced more than a decade ago. Despite a generally lower resolution compared to WLE, the different autofluorescent pattern in case of metaplastic or dysplastic lesions increased the diagnostic yield of targeted biopsies for dysplastic or neoplastic changes (26-28). However, it has been stated early, that AFI is of limited quality if used as single technique (29). Thus, the combination of a high resolution magnifying endoscope with AFI and NBI filter systems was introduced as so-called Trimodal Imaging Endoscopy (TME) for a direct assessment of AFI-positive lesions NBI imaging including a magnifying zoom technique to increase the sensitivity, specificity and accuracy of endoscopic diagnostic procedures (30). However, the final diagnostic value of the increased rate of targeted biopsies still needs to be evaluated (31, 32).

A similar approach to NBI represents the flexible spectral imaging colour enhancement endoscopy (FICE), which offers more than one optical filter in different digital channels that can be easily switched during the investigation. Each mucosal structure can be assessed best by certain channels resulting in high interobserver agreement (33). Furthermore, the high contrast enhancement allows a better definition of the lateral demarcation line of gastric lesions compared with other approaches, especially for depressed lesions (34, 35)(Figure 1).

The most recent technique is confocal laser endoscopy (CLE) that enables an *in vivo* real-time assessment of the histopathological alterations present in the gastric mucosa. First classifications of the related pit patterns have been developed on surgical specimens from gastric cancer patients and then validated in healthy volunteers (36). *In vivo* differentiation between physiological tissue from different regions of the stomach was possible as well as to distinguish between IM, non-metaplastic glandular atrophy and neoplastic lesions (36). This high-end magnification technique allows the detection of goblet cells and of a respective brush border in an absorptive intestinalised epithelium in case of intestinal metaplasia (IM) with a sensitivity about 90% and a specificity even higher (37, 38). Even the assessment of epithelial barrier function in response to *H. pylori* eradication can be assessed (39). However, the diagnostic accuracy and gastric cancer detection rate for CLE is significantly higher for experienced compared to inexperienced examiners since the investigation is time-consuming and demands high concentration and patience (40). In experienced hands – or better with experienced eyes - the diagnostic accuracy for the discrimination of cancerous lesions or even high grade intraepithelial neoplasia can be as high as 98.8% and by this comparable

## Modern approaches in gastric cancer assessment

to the classical histopathological evaluation (41, 42). This



**Figure 1.** FICE imaging of an early gastric cancer before and after endoscopic resection. a-c) The images show an early gastric cancer of the intestinal type with central ulceration. The different FICE filter channels are used for better demarcation of the lateral tumour margins. d) Demarcation of the planned resection zone with an argon plasma beamer. e) Circumferential incision of the dissection zone by a hook-knife down to the level of the *submucosa*. f) Complete histopathological confirmed R0 resection of the intramucosal cancer. In view are the muscle layers of the *muscularis propria*.

approach has further value in the assessment of resection margins after endoscopic treatment of early gastric cancer, there being even superior to biopsy sampling and histopathological assessment (43, 44). Another approach for the future could be the *in vivo* microscopy after labelling of the gastric mucosa with cancer specific fluorescent agents (45).

These technical advances can also be applied for diagnosis and risk stratification of premalignant conditions of the gastric mucosa like intestinal metaplasia or atrophic gastritis. For NBI, the appearance of a light blue crest sign is a valuable surrogate for the presence of IM (46, 47). The combination of an optical filter system with CLE can even enable "real time histology" that could reduce the number of biopsies that are necessary for the diagnosis of IM or atrophic changes (48).

However, despite the exciting development in the field of diagnostic gastrointestinal endoscopy, this does not allow so far to replace routine standard biopsy work-up and thorough inspection by standard white light endoscopy (49, 50). So far, new optical systems are only of practical value in specialised high volume centres in Asia, i.e. in regions with high incidence of gastric cancer.

## 4. HISTOLOGICAL ASSESSMENT

### 4.1. Histopathology of gastritis and attributable risk

The up-dated Sydney classification of gastritis with its updates is still the most widely used system for characterizing the status of the gastric mucosa either for

research purposes or clinical practice. It combines topographic, morphological, and etiological information (51). The analysis of five biopsies – two from the antral part, one from the angulus, and two from the gastric body - is required to characterise gastritis in the absence of any visually detectable lesions. In addition to glandular atrophy and IM, each graded according to a visual analogue scale (0: no changes, 1: minor changes, 2: moderate changes, 3: severe changes), the classification requires also the reporting on the presence of *H. pylori*, active and chronic inflammation, the latter by the degree of mucosal infiltration by neutrophil granulocytes and lymphocytes, respectively.

In the past decade new classification systems have been developed to enable more consistent stratification of the individual gastric cancer risk attributable to the present preneoplastic changes of the gastric mucosa. OLGA and OLGIM staging systems for gastric premalignant lesions aim to simplify the clinical approach while using the same biopsy work-up as for the Sydney system. The abbreviation OLGA stands for "Operative Link on Gastritis Assessment" and is based on the assessment of glandular atrophy, while OLGIM emphasizes the importance of Intestinal Metaplasia.

The initially proposed OLGA system is based on pooling the atrophy stages in each part of the stomach into a simple scoring system ranging from "0" to "IV" (52). The stage by itself does not allow to judge the topography of the lesion revealed (in particular for the lower stages), but it indicates the individual likelihood to develop malignant

## Modern approaches in gastric cancer assessment

neoplasia since most of the cancer cases are expected to develop in patients who present with stages III and IV (53). In addition, the stage distribution is convenient also for research purposes, like the correlation of the histopathological OLGA score with serological biomarkers (54).

Considering that the interobserver agreement is better for assessment of IM than of non-metaplastic glandular atrophy, the OLGIM system is using the same approach, but classifying the degree of IM instead of assessing atrophic changes with or without metaplastic transformation of the gastric mucosa (55). It is still under debate if one of these approaches is superior to the other, resulting in a higher sensitivity for gastric cancer risk assessment (55-57). OLGA based stratification of premalignant lesions in the stomach can also be associated with non-atrophic mucosal alterations and certain inflammatory conditions (58, 59).

### 4.2. SPEM – spasmolytic polypeptide-expressing metaplasia

Spasmolytic polypeptide-expressing metaplasia (SPEM) is markedly characterised by an induction of the gene expression of the spasmolytic polypeptide which has been identified as the trefoil factor 2 (TFF2) (60). SPEM is mainly associated with corpus predominant gastritis and has been shown to be associated with gastric cancer development (61, 62). In surgical specimens from patients with early gastric cancer there was positive evidence of SPEM in the tumour surrounding mucosa if the tumour was located in the gastric body or at the body-antrum junction (63). In three quarters of the cases there was also SPEM in the tumour-distant body mucosa. TFF2 could be detected in 76% of dysplastic cells. In samples from a control cohort with gastritis and without neoplastic lesions, 82% of patients who developed gastric cancer during follow-up were positive for SPEM compared to 37% in cases without malignant transformation (63). Similar results are reported for the mucosa of patients with remnant adenocarcinoma after limited resection of gastric cancer (64).

Data from animal experiments suggested that the metaplastic cells derive from gastric Chief cells or alternatively develop by activation of basal crypt progenitor cells (65). However, recent data has shown, that these leucine-rich repeat containing G-protein-coupled receptor 5 (Lgr5) positive gastric stem cells are not the origin of SPEM (66, 67). Signalling cascades leading to the induction of these metaplastic changes seem to involve PGE2-related pathways and even Wnt-dependent signals (68).

### 4.3. Subtyping of mucins

The differential expression of mucin (MUC) subtypes marks the phenotype of gastrointestinal epithelium and allows the differentiation of physiological or metaplastic mucosa. There is physiological expression of MUC5AC in the superficial epithelium and the upper part of the gastric pits, as well as MUC6 in the lower part of the gastric glands (69). Further gastric-type mucins are MUC1 and the human gastric mucin (HGM). MUC2 is mainly

expressed by goblet cells and characterises intestinal epithelium (69). Induction of the intestinal transcription factor CDX2 in the gastric mucosa leads to an upregulation of MUC2 gene expression and is mostly accompanied by a downregulation of the gastric transcription factor SOX2 mirrored by decreased MUC5AC secretion (70, 71).

These processes can be a response to *H. pylori* induced chronic inflammation since the degree of the lymphocellular infiltration is concordant with the intramucosal level of MUC2 expression and the pro-inflammatory cytokines *TNF $\alpha$*  and *IL1 $\beta$*  are capable of MUC2 gene induction (72). In contrast, gastric type mucins are significantly lower expressed in the gastric body of *H. pylori* positive patients, but show increasing levels after eradication therapy (73, 74). Lower expression is maintained in case of present mucosal transformation towards atrophic changes, dysplasia and gastric cancer.

According to the mucin expression profile, adenocarcinomas in the stomach can be classified into a gastric, an intestinal, and a gastrointestinal (mixed) phenotype (75, 76). The related mucin phenotype can alter in relation to the status of *H. pylori* infection. After eradication of *H. pylori* the mucin expression profile in the tumour tissue shows more often the gastric predominant type, whereas in non-eradicated patients the intestinal type is more present being associated with a less favourable prognosis (76-78).

Atypical mucins like MUC13 can be detected in 90% of IM, and MUC13 also upregulated in gastric cancer, mainly in intestinal type adenocarcinomas (79, 80).

Interestingly, the phenotype of present IM has not to be related to the histological type of the cancer, and intestinal type gastric cancer shows even often a gastric mucin profile (81, 82). There are still conflicting results concerning the correlation of mucin expression with clinicopathological parameters (83-87). MUC1 expression in the tumour centre correlates with advanced TNM-stage and lymph node involvement, and expression at the invasion front of the cancer represents an independent predictor for worse prognosis in multivariate analysis (88). MUC1 expression is lost during dedifferentiation of gastric tumours and most studies confirm a positive association to distant metastases, accompanied in most cases also by a decrease of MUC5AC gene expression (82, 89).

A recent genome-wide association study revealed single nucleotide polymorphisms (SNPs) of the MUC1 promotor to be involved in differential regulation of this mucin and therefore to be related to gastric carcinogenesis, mainly of the diffuse type (90). SNPs in the MUC1 gene have already been reported to be related to gastric cancer, whereas SNPs in the MUC5AC gene have only a minor impact (91). In contrast, polymorphisms of the MUC2 gene are associated with a decreased risk of progression of premalignant changes in the gastric mucosa, also interfering with the probability of regression after *H. pylori* eradication therapy (92). MUC2 gene expression can

## Modern approaches in gastric cancer assessment

also be epigenetically regulated by hypermethylation of its promoter region (93).

In a minor proportion of gastric cancer patients mucins can be detected in the peripheral blood as indicator for an induced expression. Surrogates like anti-MUC1 IgG in the peripheral blood are not constantly present and have therefore no value for clinical practice (94-96).

### 4.4. Expression of HER2

In the era of individualised targeted therapy of malignant diseases there has been a recent breakthrough for the treatment of gastric cancer when the results of the ToGA trial have been published, a prospective, randomized, placebo-controlled phase III trial on almost 600 patients with cancer of the stomach or at the oesophagogastric junction (97). In these patients with positive expression of the HER2 molecule, trastuzumab, a monoclonal antibody against HER2, has been administered in combination with standard cisplatin/5-fluorouracil based systemic chemotherapy. Mean overall survival of patients receiving the trastuzumab combination has been 13.8 months compared to 11.1 months in the placebo group (cisplatin/5-FU alone). Since this difference was not statistically significant, the major impact of the study was given, when a subgroup analysis revealed that in patients with strong expression of HER2 (IHC 3+ or IHC2+/FISH+) survival was as high as 17.9 months being highly significant compared to the placebo group (97).

HER2 can be detected in 10.1% to 20.6% of patients with gastric adenocarcinoma with higher positivity in intestinal type tumours and in patients with distant metastases (97-102). Results concerning HER2 expression as prognostic indicator are still conflicting with studies showing partly poorer and partly improved survival for HER2 positive patients (99, 100, 103-105). A systematic review on 42 publications including 12,749 patients reported in 71% of the trials an association of positive HER2 status with poor survival, serosal invasion, positive lymph node status, distant metastases, and more advanced stage of the disease (106). In another comprehensive review on 35 studies assessing survival, 20 could not demonstrate a survival benefit, two reported longer and 13 even shorter survival in HER2 positive patients. Overall five-year survival was 42% vs. 52% in favour of HER2 negative patients. However, the introduction of trastuzumab-based treatment regimens results in a clear survival advantage for HER2 positive patients (103).

HER2 expression is increasing with severity of mucosal alterations from lowest in low grade IEN and highest in adenocarcinoma of the stomach, even showing further increase with dedifferentiation of neoplastic lesions (107). Since there is high intratumour heterogeneity (Figure 2), there is an on-going debate whether biopsy sampling during endoscopy is adequate for assessment of HER2 status, and if tissue from metastatic sites (e.g. liver) can be used for estimation of the primary. As of today, it can be stated that there is high concordance between HER2 expression in the primary tumour and distant metastases as well as affected lymph nodes (101, 108-110). There is also

no significant difference in the diagnostic quality if tissue from surgical resection specimens or endoscopic biopsies were used (101, 110, 111). However, these data need still further validation in clinical practice.

Since molecular testing and quantification of a positive test result has a significant value for the treatment decision there is still debate which test method presents the best diagnostic yield. Fluorescence *in situ* hybridisation (FISH) is superior to classical immune-histochemical staining (IHC) although it is more complicated and more expensive (112). Another option is dual-colour silver chromogenic ISH (CISH/SISH) that reveals even more precise test results and a higher detection rate than FISH (98, 99, 112). Tissue microarrays have been discussed to further enhance the diagnostic yield (100, 104, 113, 114).

HER2 expression in gastric cancer might even correlate with HER2 detection levels of the serum of the patients, but it is doubtful if these marginal changes render a benefit as a clinical biomarker for gastric cancer screening (115).

## 5. SEROLOGICAL BIOMARKERS

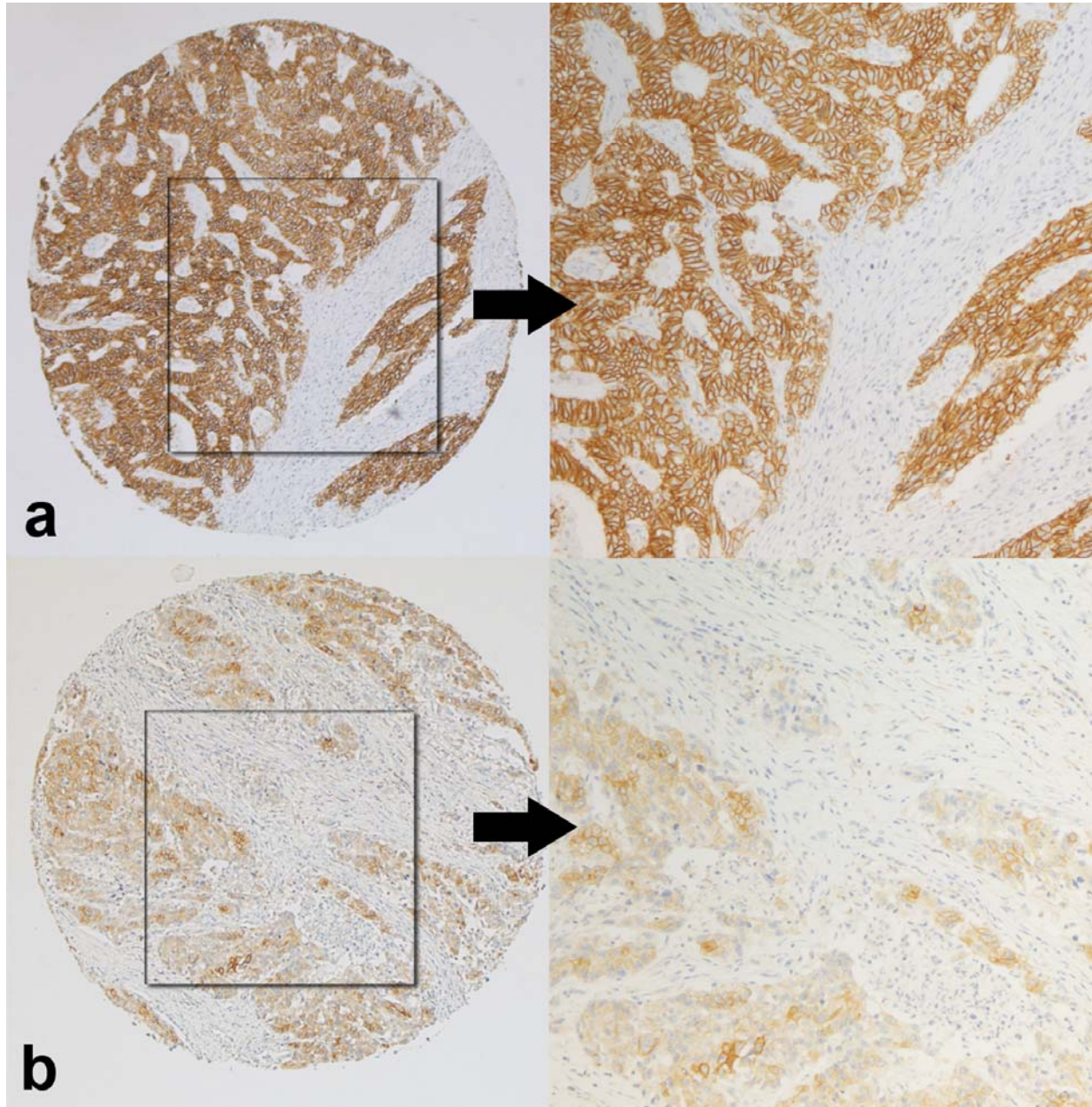
### 5.1. Pepsinogens and the „serological biopsy“

Pepsinogens are pro-enzymes of pepsin. Pepsinogen I (Pgl) is exclusively produced by the chief and mucous neck cells of the corpus, while pepsinogen II (PglI) is also secreted by cardiac, pyloric and Brunner gland cells (116). Pepsinogen levels are decreased if atrophy occurs in the gastric body, while an increase is observed during inflammation. To reduce the possibility of false normal results at the occasion when both atrophy and *H. pylori* caused inflammation co-exist, the ratio between Pgl and PglI (Pgl/II) is considered a more reliable marker than Pgl alone (117-119).

A comprehensive meta-analysis published in 2006 on more than 40 studies including about 300,000 individuals suggested the rationale of using pepsinogen testing to identify individuals at high risk to develop gastric cancer who would need further diagnostic work-up (118). Decreased pepsinogen levels yield a sensitivity of 66.7-84.6% and a specificity of 73.5-87.1% for the detection of atrophic gastritis (120-123). However, lower sensitivity (36.8%-62.3%) has been reported for direct gastric cancer screening even with similar cut-off values (124-126).

Considering confounding factors in gastric cancer patients, that could be included in a further stratified analysis, would be a valuable approach to modulate outcome of the pepsinogen test (127). However, neither Laurén type, nor tumor localisation or tumor stage have an influence on the serum values for Pgl, PglI and Pgl/II (127, 128). Only for the Pgl/II a significant difference between intestinal and diffuse type carcinomas is documented in some studies which is related to the higher incidence of intestinal metaplasia and glandular atrophy in case of intestinal type cancers (129, 130).

Amidated gastrin-17 (G-17) has been suggested as additional marker to characterise atrophy in the antral



**Figure 2.** Representative examples of Her2 immunohistochemical staining. a) Strong and complete membranous Her2 staining with homogenous distribution throughout the sample (IHC-score 3+). b) Heterogenous distribution of Her2 staining, IHC-score 2+; FISH analysis is recommended. Original magnifications 100x, 200x.

part of the stomach. G-17 is a sub-fraction of total gastrin consisting of 17 amino-acids that is secreted exclusively by the G-cells in the gastric antrum (131-133). G-17 levels in the serum can indicate two conditions: decreased levels could be indicative for atrophy in the antral part of the stomach, but increased are characteristic for corpus atrophy (further characterised by low pepsinogen levels) in the absence of atrophy in the antral part. G-17 levels in the circulation are increased after food intake; therefore the measurements of G-17 following a provocation with a protein-rich meal are considered the best indicator of the

function of antral G-cells (132, 134). However, due to the limitations of this provocation test (e.g. repeated blood sampling since other markers are usually analyzed at a fasting state, intake of a test-meal, duration of the test), in many studies only single measurement at fasting state is performed (116, 135).

A false positive increase of G-17 could be feedback to drug-induced hypochlorhydria in the stomach or inflammation, and a decrease could be related to acid peptic disease (133, 136-138). Therefore, despite a good

## Modern approaches in gastric cancer assessment

specificity of decreased G-17 for diagnosing antral atrophy in Caucasians (91.5% at a fasting state and 92.6% after stimulation), the sensitivity is unsatisfactory (15.4% at a fasting state and 30.8% after stimulation)(139).

Simultaneous detection of pepsinogens and *H. pylori* antibodies have been suggested by Japanese investigators, this approach is known also as the ABC(D)-method (119, 140). Individuals are classified as group „A“ if pepsinogen levels are normal and *H. pylori* antibodies absent, group „B“ if pepsinogen levels are normal and *H. pylori* antibodies present, group „C“ if pepsinogen levels are decreased and *H. pylori* antibodies present, group „D“ if pepsinogen levels are decreased and *H. pylori* antibodies absent. Based on the results individual risk assessment can be optimised and endoscopic surveillance individually tailored (141, 142). Additionally to Pgl, PglI and *H. pylori* antibodies, assessment of <math>g17</math> can also be included in the test-panel for a higher diagnostic yield (116, 133).

Several studies have demonstrated that decreased pepsinogen levels (alone or in combination with anti-*H. pylori* antibodies and/or G-17) are predictive for gastric cancer development. In the Hisayama study (143) 2,446 subjects aged 40 years and above were followed prospectively for 14 years. The hazard ratio (HR) for developing cancer was substantially higher for the group with decreased pepsinogen levels than in individuals with normal test results (HR 4.6, 95% CI: 2.4 - 8.6 for men; HR 5.8, 95% CI: 2.0 - 17.1 for women). Similarly, 5,706 male employees aged 40-60 years were followed for 10 years in the Wakayama City study (144). The HR for gastric cancer development was 5.2 (95% CI: 2.8-9.5) for patients with significantly decreased Pg levels. Watabe et al. (140) followed 6,983 individuals for the duration of 4.7 years. Hazard ratios for gastric cancer incidence were 6.0 (95% CI: 2.4-14.5) in the group of decreased pepsinogen and positive anti-*H. pylori* antibodies (group C according to the „ABC(D)“ classification), and 8.2 (95% CI: 3.2-21.5) in the group of decreased pepsinogen but negative anti-*H. pylori* antibodies (group D). The study demonstrated that a decrease of anti-*H. pylori* antibodies during advanced stages of atrophy is related to an even higher risk. In contrast to these results, a long-term study in a high-risk area of China, showed a higher risk in individuals with decreased pepsinogens and positive anti-*H. pylori* antibodies. This group demonstrated an increase in the relative risk (RR) of 27.5 (95% CI: 3.4-225.4), while in the group with decreased pepsinogens and negative anti-*H. pylori* antibodies the RR was 23.2 (95% CI: 2.1-260.9) (145). Recently, an initial report from a case-control study in Russia indicated a HR for gastric cancer development of 4.31 (95% CI: 1.5 -12.5) for Pgl/II <math>< 3.0</math>(146). Data from Japan has demonstrated that a serological screening for pepsinogens with further referral for upper endoscopy or photofluorography if the test is positive, has been capable to decrease gastric cancer deaths by 76% (in individuals screened 1 year before the diagnosis) or by 62% (in individuals screened within 2 years) (147).

A large cohort study in a population from Portugal demonstrated the feasibility of pepsinogen testing approach even in a Western population (148). A

total of 13,118 individuals have been followed for five years. Of the 446 individuals (3.4%) with decreased pepsinogen levels, 274 underwent upper gastrointestinal endoscopy; six cancer cases have been detected representing one cancer per 2,200 tests or one incident case per 74 positive tests (148).

Different cut-off values to for determining decrease in the pepsinogen levels have been used in many studies (149), and different test-systems are traditionally used in Asia and Europe. Therefore, the results in absolute values cannot be directly compared between the different studies(150). Based on the above, the current guidelines emphasise the need for regionally validated test-systems (151).

### 5.2. Cancer autoantibodies

Autoantibodies against tumor-associated antigens have been identified in several cancer types (152-154). Due to their specificity and stability in the serum, they represent attractive targets for the development of noninvasive serological tests for the early detection of cancer (155). However, the frequency of antibodies against particular tumor associated antigens is rather low, typically ranging 1-15%, therefore an approach of panel-testing is frequently used to explore cancer-specific antibodies (155). Recently, a 45 cancer-associated autoantibody signature was identified able to discriminate gastric cancer from healthy controls with 58.7% sensitivity and 89.7% specificity by using a T7 phage-displayed SEREX approach combined with the phage-displayed antigen microarray technology and a novel strategy for the analysis of microarray data(155). There is a potential to increase further the sensitivity of this test.

### 5.3. Circulating cancer cells

Identification of circulating tumour cells (CTCs) has been related to the presence of a systemic disease and appearance of peripheral metastasis (156). Detection of CTCs is suggested useful for estimation of the individual prognosis and for monitoring several cancer types, including breast, lung, prostate, skin, colon and gastrointestinal cancers (157-159).

A larger number of CTCs has been found in metastatic gastric cancer than non-metastatic disease with the identification of two or more CTCs being an indicator for more advanced stage disease as well as peritoneal dissemination (160). In a recent study by using a telomerase-specific viral agent to detect CTCs in peripheral blood of gastric cancer patients a significant relationship between the number of CTCs and the prognosis of the disease was revealed. However, not all the CTCs might have equal metastatic potential since here, recurrence of early stage disease was not identified even in the presence of CTCs (159). However, the direct detection of gastric cancer derived CTCs is difficult and requires high methodological effort. Therefore, there have been approaches to use RNA products of these cells as surrogate (94, 161, 162).

An alternative to CTCs is the detection of circulating nucleic acid shed from necrotic or apoptotic

## Modern approaches in gastric cancer assessment

cells (163). Using mutation or methylation specific assays it is possible to quantify the fraction of DNA originating directly from tumour cells as well as the total levels of circulating nucleic acid (164, 165). Quantification of the level of CNA has been demonstrated to be a sensitive early marker of disease response to chemotherapeutic and surgical treatments (164, 166).

Several studies have demonstrated elevated levels of circulating nucleic acid in gastric cancer though as yet the clinical relevance of this remains unclear (167, 168).

### 6. VOLATILE ORGANIC COMPONENTS IN BREATH

The main constituents of human breath are nitrogen, oxygen, carbon dioxide, water vapour and inert gases. In addition, thousands of volatile organic compounds (VOCs) are exhaled at very low concentrations (estimated as parts per trillion (ppt) or parts per billion (ppb) by volume of the exhaled breath) (169, 170). While part of the substances is of endogenous origin and could be characteristic for metabolic processes in the human body (including cancer), others are exogenous, i.e. passing through the human body (169, 171).

Initial results on the potential applications of volatile marker tests for detection of several cancer types have been published, e.g. for lung, breast, colorectal and prostate cancers (172, 173), and hepatocellular carcinoma (174).

A recently published study addressed the potential of volatile markers to diagnose gastric cancer (175). By using novel cross-reactive, highly sensitive gas sensor allowing to identify and separate VOC patterns, the obtained results demonstrated 89% sensitivity and 90% specificity to differentiate gastric cancer cases from non-malignant conditions after cross-validation, irrespective of important confounding factors such as tobacco or alcohol consumption and *H. pylori* infection (176). Further validation and reproducibility studies are required, including different populations.

### 7. MOLECULAR DIAGNOSTICS

#### 7.1. MicroRNA

MicroRNAs (miRNA) are short RNA molecules of approximately 22 nucleotides that are involved in posttranslational regulation of gene expression (177, 178). Dependent on the sequence homology of miRNAs and targeted RNA the effect may vary from partial inhibition of the mRNA translation to cytoplasmic degradation of the mRNA transcripts (179, 180). Thus, each of currently known human miRNAs may control hundreds of mRNA targets and so be involved basically in every cellular process which becomes obviously deregulated during the carcinogenesis. For the detailed introduction into the miRNA biogenesis we refer to recently published excellent reviews (181-183). In this section, we will briefly introduce

the most important aspects of miRNA research in regard to gastric cancer development and biomarker research.

Following the pivotal work of Lu *et al.* demonstrating differential miRNA expression across various tumours by using 217 miRNAs (184), an increasing number of studies have clearly and consistently shown differential expression of miRNAs in gastric cancer compared to normal tissues. At present, Ueda and colleagues have performed the largest study on gastric cancer tumours (185). Using a respectable number of gastric cancer tissues paired with non-tumour samples the authors identified 22 up- and 13 downregulated miRNAs. Additionally, using the pattern of the 19 most significantly deregulated miRNAs it was possible to discriminate gastric tumours according to their histological type. In particular, cluster analyses revealed miR-105, -100, -125b, -199b, -99a, 143, -145 and -133a upregulated in diffuse type gastric cancer, while miR-373-3p, -498, -202-3 and -494 were upregulated in intestinal type lesions. Some miRNAs were related to the stage of disease, and, most importantly, let-7g, miR-214 and -433 were identified as independent prognostic biomarkers predictive for overall survival in gastric cancer patients (185).

There is increasing evidence that certain miRNA patterns are also associated with premalignant stages or even risk conditions like *H. pylori* driven inflammation (186, 187). Interestingly, *H. pylori* eradication can result in at least partial normalization of the deregulated miRNAs, further underlining the clinical importance of miRNAs in the initiation and progression of gastric cancer.

Furthermore, an increasing number of studies provided substantial evidence for applicability of miRNAs as non-invasive biomarkers for gastric cancer (188, 189). MiRNAs are easily and reproducibly detectable in various body specimens including blood (188, 189), gastric fluids (190, 191), faeces (192, 193), saliva (194) and others (195). A remarkable stability of miRNAs from degradation in body fluids is perhaps one of the most exciting characteristic of miRNAs which has been linked to the ability to build protein complexes and be sequestered in exosomes (196, 197). Although the mechanism of miRNA extravasation is not completely understood, it is believed that either active release of exosomes or passive extravasation of the protein complexes during apoptosis or cell death are the main source of circulating miRNAs (196-198). This is further supported by the fact that expression pattern of circulating miRNAs change dramatically following surgical resection of the tumour; however, exosome release is not a tumour-specific event but a rather ubiquitous process, and it becomes clear that not only direct tumour-related release, but also indirect changes in miRNA expression (stroma- or immune cells) may be a cause of measurable deregulation of circulation miRNAs. After the publication of the first data, several groups have confirmed the differential expression of certain miRNAs in the blood of cancer patients, partially using high-throughput techniques (199-201). So far, there has been only minor attention paid to the association between *H. pylori* and gastric cancer associated circulating miRNAs. It has been



## Modern approaches in gastric cancer assessment

shown, that levels of miRNA expression in sera were highly correlated with *H. pylori* status both in gastric cancer patients and *H. pylori* infected controls, and particularly miR-223 was present at a significantly higher level in *H. pylori* infected individuals compared to those without the infection (199).

Another elegant approach is the use of gastric juice for miRNA expression analysis. Guo *et al.* assessed miR-421, -129, -21 and -106b expression in the gastric juice of patients with mild superficial gastritis, atrophic gastritis, gastric ulcer or gastric cancer. Although all tested miRNAs we concluded to be promising for the screening of gastric cancer, prospective validation in independent patients cohort is still needed (190, 191, 202).

Mutation or specific variations in the DNA sequence of miRNA genes may be associated with alterations in miRNA processing, if the base pair changes are located in precursor sequence, or it may influence the functional interaction of miRNA with a target molecule, if located in a miRNA coding sequence. Two miRNA-SNPs have been in primary focus of several studies, rs2910164 G>C of the pre-miR-146a and rs11614913 of pre-miR-196a-2 (203-205). Although the results are not homogenous, it seems that at least in Asian gastric cancers, these SNPs may be associated with susceptibility for atrophic gastritis or an increased risk for gastric cancer. Large unbiased genome-wide multicentre studies will be needed to estimate the exact meaning of those changes.

In similar fashion as coding mRNA, transcription of miRNA genes is regulated by epigenetic status of the promoter regions. DNA methylation together with histone modifications are associated with differential miRNA expression by regulation of the accessibility of certain transcription factors. Ando *et al.* observed an up to 13-fold increase in the methylation level of miR-124a in gastric biopsy samples from patients with *H. pylori* infection (206). Several other miRNAs miR-137, miR-34b or -129-3p have been also suggested, but systematic quantitative methylation analyses are still missing (207, 208).

Taken together, miRNA research has been truly a fast break event. At present, the unique features of miRNA with a rapidly increasing amount of data stands next to a lack of understanding in regard of multistep process of carcinogenesis, and unanswered methodological challenges need to be solved prior the broad clinical implementation of miRNAs as biomarkers. As for other biological markers assessed by high throughput techniques, biostatistical evaluation of miRNA patterns facilitate the identification of signature that can be uniquely attributed to certain diseases. It has been shown recently, that the differential expression of miRNA signatures can be assessed for the distinction of gastric and oesophageal adenocarcinoma (209). These approaches can support a better understanding of the different pathobiological pathways leading to cancer of the oesophagogastric junction of either gastric or oesophageal origin.

## 7.2. Epigenetic changes

Epigenetic mechanisms refer to different modifications of the chromatin structure that affect gene expression without altering the primary DNA sequence. The two major epigenetic mechanisms that lead to activation or silencing of the gene function are methylation and histone modification, which are related to the development of different cancers, including gastric cancer (210).

Global hypomethylation and promoter hypermethylation are common features in various gastrointestinal malignancies. The first feature refers to the loss of DNA methylation and is linked with genomic instability and tumor formation, while promotor hypermethylation causes transcriptional silencing of tumor suppressor genes and may affect important molecular pathways (211).

Methylation of CpG islands has been assessed in promoters of different genes related to gastric carcinogenesis. A growing number of genes related to cell cycle, apoptosis, tumour invasion, cell adhesion, cell signaling, and transcription have been shown to be silenced by hypermethylation in gastric cancer. Genes encoding *CDHI* (212), *FOXD3* (213), *RUNX3* (214), *TPEF* (215), and other well-known tumor suppressor genes have been shown to be affected by methylation. As mentioned above, recent studies on small non-coding miRNAs have also revealed modifications of epigenetic regulation that occur in gastric cancer (182). The genes that are effected by methylation in gastric cancer are extensively discussed in several reviews (216-218).

Studies on *H. pylori* and *Epstein-Barr virus* (EBV) infection show that the carcinogenic effect of these pathogens may be reinforced by inducing methylation changes in the gastric mucosa (214, 219). Interestingly, inflammation induced by *H. pylori* infection was critical for methylation induction of promoters containing CpG islands through the release of reactive oxygen species and nitric oxide and by activation of the DNA methyltransferase (220). Furthermore, epigenetic changes in gastric cancer occur not only at the stage of malignancy, but also at early stages of cancer development including atrophic gastritis and IM (221). There are convincing data showing that methylation profiles are different for intestinal and diffuse type gastric cancer (222).

Pyrosequencing and other technological developments enabled researchers to look not only at methylation status in individual oncogenes, but also allowed to assess global methylation patterns in gastrointestinal malignancies. Global demethylation of the tumour cell genome in gastric cancer occurs in parallel to abnormal hypermethylation of tumour suppressor genes (223). Data on global-methylation status in gastric cancer suggest that this could be used as a marker to detect metastasis and may reflect the malignant potential of gastric cancer (224). Further studies on epigenetic changes indicate that global methylation occurs in *H. pylori* induced gastritis representing an early event in gastric carcinogenesis (225).

## Modern approaches in gastric cancer assessment

As the methylation status in the tumour tissue is resembled by patterns retrieved from serum samples, methylation status has the potential to become a non-invasive marker, which could be used for early diagnostics of gastric cancer and a novel target for cancer prevention(226). Furthermore, different studies showed that epigenetic changes are predictors for response to chemotherapy and patient survival and thereby might influence the decision making process in the treatment of these patients (217). Up to date, methylation based non-invasive testing is not yet available for diagnostic, prognostic or treatment-related decision making in the clinical setting, but ongoing work in the field may reveal significant benefits for the patients in the near future.

### 7.3. Genetic alterations

Genetic variation influences individual susceptibility to different malignancies, including gastric cancer (227). Significant advances in molecular biology, next generation sequencing and bio-banking activities enabled researchers to examine different genetic entities in relation to numerous human diseases. Genetic predisposition to sporadic gastric cancer remains largely not understood (228). The only genetic test which can be applied in today's clinical practice is screening for *e-cadherin* (*CDH1*) mutations in familial cancer cases, however, this approach is applied in very few specialised centers (229).

The most common type of genetic variation in the human genome is single nucleotide polymorphisms (SNPs). A currently accepted hypothesis suggests that individuals with a pro-inflammatory genetic profile are more vulnerable to *H. pylori* infection related damage and gastric cancer development. Chronic inflammation is the major event in the model of gastric carcinogenesis, which has been demonstrated in human and animal studies (230, 231). Based on this paradigm, genes responsible for inflammation and *H. pylori* recognition became a major interest of different gastric cancer research groups. In the year 2000, the association between a pro-inflammatory *interleukin-1 $\beta$*  (*IL1B*) gene polymorphism and an increased risk of *H. pylori*-induced gastric cancer was reported (232). This paper was followed by numerous case-controls studies on genes encoding inflammatory cytokines, Toll-like receptors (TLRs), Nucleotide-binding oligomerisation domain (NOD) receptors and other genes involved in *H. pylori* induced carcinogenesis (233). Initial studies have reported a significant effect of certain cytokine gene alterations for the risk of gastric cancer development (232), while data from more recent studies have showed marginal or no associations (234-237). Interestingly, some research groups described a relation between cytokine SNPs and progression of mucosal inflammation and atrophy(238), while genetic variations of TLR receptors might be linked with reduced risk of *H. pylori* induced diseases in the stomach(239). Polymorphisms of *IL1B* and *interleukin-1 receptor antagonist* gene (*IL1RN*) are best studied in the context of gastric cancer development. Several meta-analyses have been published summarizing the results of the smaller case-control studies on *IL1B* and *IL1RN* SNPs in relation to gastric cancer risks (240-243). Based on the conclusions of these meta-analyses, the overall impact of *IL1B* and *IL1RN* genetic alterations for gastric cancer development appears to be

marginal and questions the applicability of these SNPs as potential screening biomarkers.

The major shortcoming of small hypothesis driven case-control genetic association studies discussed above is related to the small number of individuals within the groups, which are poorly stratified according to individual variables (histological subtype, *H. pylori* status, anatomical site of the tumor, etc.). New high-throughput technologies enabled researchers to perform large genotyping association studies. Genome wide association studies (GWAS) on gastric cancer have identified several interesting candidate genes that could serve as non-invasive markers for early detection. *PSCA*, *PLCE1* and *MUC1* gene polymorphisms have been linked with gastric cancer risk in recent GWAS studies (244, 245). It is worth noticing that for some of the SNPs revealed in gastric cancer GWAS studies have also implicated in the formation of premalignant gastric conditions (246). GWAS studies on gastric cancer may serve not only for identification of genetic susceptibility loci, but may also reveal previously unknown pathways in gastric carcinogenesis.

To date, however, none of genetic alterations can be used in daily clinical practice for stratification of risk of sporadic gastric adenocarcinoma in an individual patient (151).Guidelines on the management of precancerous gastric conditions and lesions (MAPS) draw a similar conclusion stating that despite numerous studies on host genetic variations, no clinical recommendations can be made for targeted management based on these factors with regard to diagnosis and surveillance (49, 247).

## 8. BIOSTATISTICAL ASSESSMENT

The development of tissue-based microarrays and chromatin immuno-precipitation (ChIP-) analysis opened the gate to a system-wide evaluation of differential gene-expression related to a variety of diseases (248). The application of biostatistical algorithms enables the structured assessment of the tremendous amount of data that is delivered by array-based techniques. Principal component analyses and hierarchical clustering enable functional classification of the identified gene expression networks and even individual genes and related signalling pathways (249). By this approach putative target genes with prognostic or therapeutic potential can be identified as well as further interacting partners (250, 251). These target genes don't necessarily have to be deregulated below or above a certain threshold (252), comparison to „normal“ tissue or between different entities of malignant diseases defines central node-genes like transcription factors or cell cycle regulators (253). This approach can be applied for the definition of pathogenetic patterns of non-malignant diseases (254, 255)as well for specific oncological diseases, e.g. neuroendocrine tumours, hepatocellular carcinoma, breast and prostate cancer (256-259).

Computational visualisation of gene-gene-interaction networks and the related gene-clusters enables also a topological assessment of gene-coexpression as well as the integration of multiple datasets for a comprehensive

## Modern approaches in gastric cancer assessment

analysis (251, 260). The results are based on features of the ENSEMBL database and classified by “gene-ontology” terms: a) biological process, b) molecular function, c) cellular component (261). The algorithms in use are under permanent development for further optimisation of the gain of information. By genome-wide association studies (GWAS) of gene aberrations or specific expression profiles, classifiers for an individual prognostic assessment, a treatment susceptibility profile or as diagnostic marker panel can be generated from analyses on human tissue or derived cell lines (244, 262, 263).

A further basic approach is the classification or clustering of copy number aberrations, gene amplifications or deletion. Patients can be stratified in distinct subgroups according to their genetic tumour profile, that can even be associated with clinicopathological characteristics like tumour location, TNM stage including depth of invasion and lymph node involvement, or expression of certain receptor subtypes like Her2 (264-266). However, it is still questioned if the detected alterations define distinct subtypes of gastric adenocarcinomas with common genetic profiles, or if an individual “evolution” of certain tumour subclones is responsible (266).

Recently, there has been a focus on genes that are targetable by medical treatment (EGFR, MET, FGFR, HER2) (267, 268). SNPs or copy number aberrations in the respective genes might enable the prediction of the prognostic outcome after neoadjuvant or even palliative therapy. For validation of the identified gene signature and the biological relevance, assessment on protein level by immunohistochemistry or protein expression arrays is useful (265). By “array comparative genomic hybridisation” different levels of gene expression can be taken into account to demonstrate that basic genomic alterations have an actual influence on the related protein profile (269, 270). In a study on tissue from oesophageal adenocarcinomas a genome-wide SNP array was applied in case of differential gene regulation in the microarray data (271). The thereby identified target genes have then been validated by either PCR or FISH analysis. Traditional immunohistochemistry was used for definitive validation in a different study population, including the clinical data for prognostic relevance of the signature by Kaplan-Meier analysis (271).

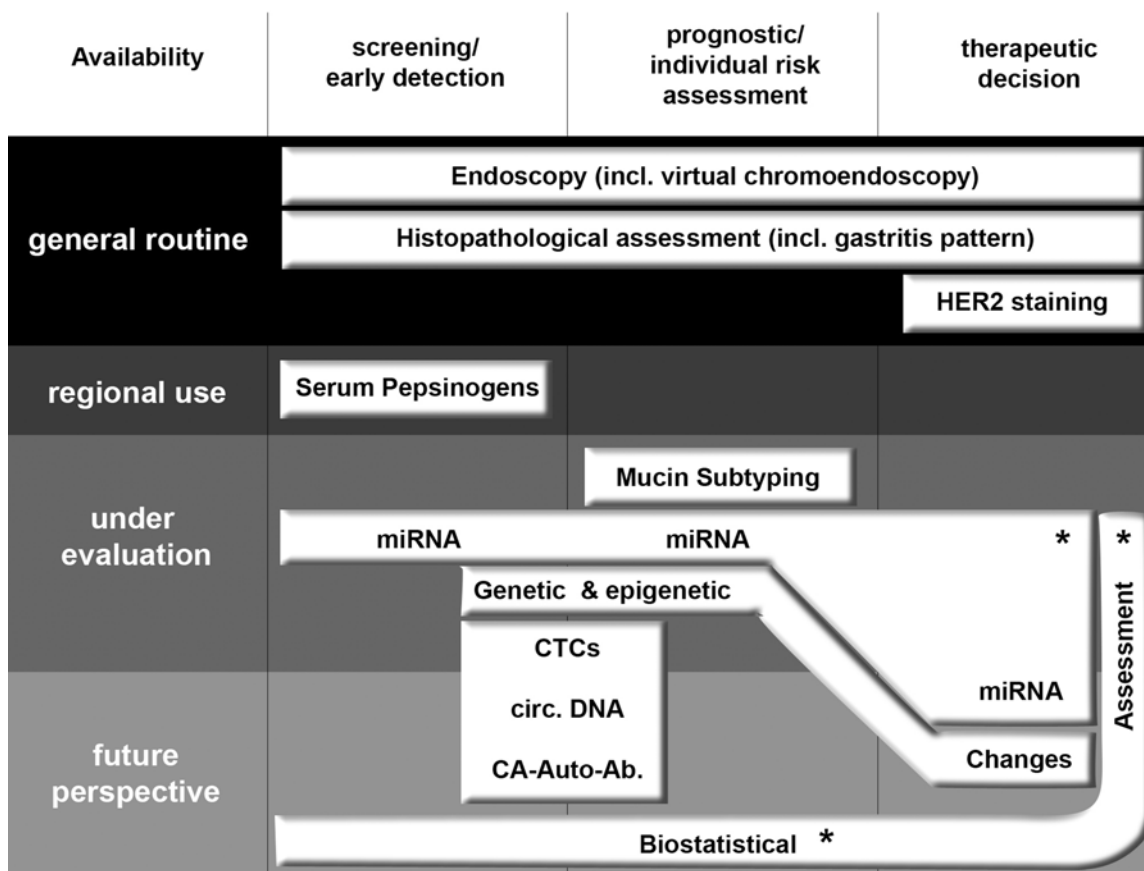
It is not mandatory to generate „de novo“ data for the initial biostatistical evaluation. The computational algorithms can also be applied on publicly available array datasets. Data published in 2006 demonstrated new molecular targets in gastric cancer, derived from formerly published datasets. However, the validation has then been performed in a prospective study population (272). This new field of computational “*in silico*” analysis of data generated by using available array platforms enable the identification of target signatures that can be correlated with any tumour specific feature, like the degree of differentiation, general tumour stage, or survival and outcome after surgical treatment (264, 273). The underlying computational algorithms can be modified according to the clinical question that should be answered.

To enable the implementation of the identified classifiers in clinical practise, it is of high importance to keep the number of involved genes and their products as low as possible (274). Thus, a prognostically relevant signature containing only two genes has been identified for oesophageal adenocarcinomas indicating the clinical response and outcome after radio-chemotherapy (275). Others demonstrated a four-genes signature significantly associated with five-years survival as independent prognostic factor in multivariate analysis(276).

By the analysis of genes that regulate stromal invasion of tumour cells, the hierarchical cluster analysis revealed a stepwise differential regulation of genes that are involved in the transition from metaplastic Barrett’s epithelium to invasive adenocarcinoma of the distal oesophagus and the gastro-oesophageal junction(277). The related genes had mainly immune-modulatory function, regulating the cytokine-cytokine-receptor interaction and TGF- $\beta$ -dependent signalling pathways. This could be shown for different adenocarcinomas of the gastrointestinal tract including gastric cancer (277).

It has been demonstrated that the related gene expression signatures that lead to activation of major oncogenic pathways include regulators of stem cell proliferation and NF $\kappa$ B-, Wnt/ $\beta$ -catenin-related signalling pathways, and are deregulated in more than 70% of the analysed cancer samples (278). A major task is the identification of mechanisms that make the carcinoma prone to local invasion and metastatic spread (279). However, even in the same organ of origin – e.g. the stomach - the involved regulatory processes show different patterns depending from the tumour localisation. Respective differences could be identified for proximal versus distal gastric cancer of the intestinal type, especially, when compared with diffuse type neoplasias (280, 281). Gastric cancer subtypes, identified by their gene expression signature as well as by the patterns of copy number aberrations and epigenetic changes, have been demonstrated to present distinct clinical response to treatment (282). Lei *et al.* presented data on a stratification into three different gastric cancer subtypes showing not only specific pathobiological characteristics but also remarkable differences concerning the response to treatment with either 5-fluorouracil or compounds targeting the PI3K-Akt-mTOR axis (282). These data facilitate the design of clinical trials and the use of small molecule targeting therapeutic agents (283).

Besides genetic alterations there have been numerous attempts to analyse directly the changes on protein level to identify protein signatures that are related to TNM stage and the degree of tumour differentiation (284). This “Proteomics” approach was also successfully used for prediction of response to neoadjuvant therapy or even to specific agents like MET-pathway inhibitors (285, 286). The data generated by high-throughput techniques like matrix-assisted laser desorption/ionisation (MALDI) imaging can be entered in similar computational algorithms that enable hierarchical clustering and general principal component analysis (287), and the related protein



**Figure 3.** Overview on diagnostic tools for gastric cancer assessment. The image gives an overview on the techniques that are either established or under development for gastric cancer diagnostics. These approaches can be either applied for basic diagnostic assessment including population-based screening, for primary staging of the disease to define the optimal treatment strategy, or for prognostic assessment of manifest disease. Indicated are also regional differences concerning the availability of certain techniques. Techniques marked with an asterix (\*), e.g. miRNA and biostatistical assessment, are already in use in a broad perspective, but can still not be regarded for application in clinical routine.

signatures are also capable to differentiate between different tumour types, organ sites or even biological behaviour of metastases (288). An important step in this field is the identification of blood-based parameters. Some groups reported tumour-specific serum biomarkers, but up to today the sensitivity and specificity of these marker panels is far too low to be transferred into clinical practise (96, 289).

### 9. VIEW TO THE FUTURE AND CONCLUSION

Although the incidence of gastric cancer is globally declining, the disease will still represent a major healthcare issue during the decades to come; therefore, identification of individuals at risk and detection of the disease at early stage remains a big challenge. The exponentially rising costs for targeted treatment modalities in case of advanced stage disease require strong and consistent predictors for treatment response to guide the therapeutic decision.

Biomarker testing with pepsinogens for identifying individuals at increased risk for cancer

development is considered by the guidelines in the East and West (49, 151, 290). Although there have been recent significant advances concerning the techniques applied that opened the gates to a variety of blood or tissue based investigations, none of these is at the step to a routine application in a clinical setting yet.

At the moment there are no perfect non-invasive screening tools for gastric cancer available. The most extensive studied of the available tests are serum pepsinogens; however, even there additional data are necessary before these can be recommended for organised screening programs, especially in non-Asian countries.

New and exciting developments are currently under investigation, these include miRNA signatures, cancer autoantibody panels, volatile components in exhaled breath, and other (Figure 3). However, there is still way to go before they could be available to the practice; in addition cost-effectiveness will have to be addressed before such tests could be implemented to population-based screening programs for gastric cancer.

## 10. ACKNOWLEDGEMENTS

We thank Dr. Jochen Weigt, Otto-v.-Guericke University of Magdeburg, Germany, for the endoscopic images (Figure 1).

## 11. REFERENCES

1. D M Parkin: The global health burden of infection-associated cancers in the year 2002. *International Journal of Cancer*, 118(12), 3030-3044 (2006)

2. A Jemal, F Bray, M M Center, J Ferlay, E Ward and D Forman: Global cancer statistics. *CA: A Cancer Journal for Clinicians*, 61(2), 69-90 (2011)

3. S J Crane, G R Locke, 3r, W S Harmsen, N N Diehl, A R Zinsmeister, L J Melton, Y Romero and N J Talley: The changing incidence of oesophageal and gastric adenocarcinoma by anatomic sub-site. *Aliment Pharmacol Ther*, 25(4), 447-53 (2007)

4. W J Blot and J K McLaughlin: The changing epidemiology of esophageal cancer. *Semin Oncol*, 26(5 Suppl 15), 2-8 (1999)

5. S S Devesa, W J Blot and J F Fraumeni: Changing patterns in the incidence of esophageal and gastric carcinoma in the United States. *Cancer*, 83(10), 2049-53 (1998)

6. A A Botterweck, L J Schouten, A Volovics, E Dorant and P A van Den Brandt: Trends in incidence of adenocarcinoma of the oesophagus and gastric cardia in ten European countries. *Int J Epidemiol*, 29(4), 645-54 (2000)

7. N Schmidt, U Peitz, H Lippert and P Malfertheiner: Missing gastric cancer in dyspepsia. *Aliment Pharmacol Ther*, 21(7), 813-20 (2005)

8. U Peitz and P Malfertheiner: Chromoendoscopy: from a research tool to clinical progress. *Dig Dis*, 20(2), 111-9 (2002)

9. G Tao, L Xing-Hua, Y Ai-Ming, Z Wei-Xun, Y Fang, W Xi, W Li-Yin, L Chong-Mei, F Gui-Jun, S Hui-Jun, W Dong-Sheng, L Yue, L Xiao-Qing and Q Jia-Ming: Enhanced magnifying endoscopy for differential diagnosis of superficial gastric lesions identified with white-light endoscopy. *Gastric Cancer* (2013)

10. N Uedo, M Fujishiro, K Goda, D Hirasawa, Y Kawahara, J H Lee, R Miyahara, Y Morita, R Singh, M Takeuchi, S Wang and T Yao: Role of narrow band imaging for diagnosis of early-stage esophagogastric cancer: current consensus of experienced endoscopists in Asia-Pacific region. *Dig Endosc*, 23 Suppl 1, 58-71 (2011)

11. M Kato, M Kaise, J Yonezawa, H Toyoizumi, N Yoshimura, Y Yoshida, M Kawamura and H Tajiri: Magnifying endoscopy with narrow-band imaging achieves superior accuracy in the differential diagnosis of superficial

gastric lesions identified with white-light endoscopy: a prospective study. *Gastrointest Endosc*, 72(3), 523-9 (2010)

12. Y Ezoe, M Muto, N Uedo, H Doyama, K Yao, I Oda, K Kaneko, Y Kawahara, C Yokoi, Y Sugiura, H Ishikawa, Y Takeuchi, Y Kaneko and Y Saito: Magnifying narrowband imaging is more accurate than conventional white-light imaging in diagnosis of gastric mucosal cancer. *Gastroenterology*, 141(6), 2017-2025 e3 (2011)

13. J Zhang, S B Guo and Z J Duan: Application of magnifying narrow-band imaging endoscopy for diagnosis of early gastric cancer and precancerous lesion. *BMC Gastroenterol*, 11, 135 (2011)

14. S Maki, K Yao, T Nagahama, T Beppu, T Hisabe, Y Takaki, F Hirai, T Matsui, H Tanabe and A Iwashita: Magnifying endoscopy with narrow-band imaging is useful in the differential diagnosis between low-grade adenoma and early cancer of superficial elevated gastric lesions. *Gastric Cancer* (2012)

15. M Nakamura, T Shibata, T Tahara, D Yoshioka, M Okubo, Y Mizoguchi, M Kuroda, T Arisawa and I Hirata: The usefulness of magnifying endoscopy with narrow-band imaging to distinguish carcinoma in flat elevated lesions in the stomach diagnosed as adenoma by using biopsy samples. *Gastrointest Endosc*, 71(6), 1070-5 (2010)

16. K Miwa, H Doyama, R Ito, H Nakanishi, K Hirano, S Inagaki, K Tominaga, N Yoshida, K Takemura, S Yamada, Y Kaneko, K Katayanagi, H Kurumaya, T Okada and M Yamagishi: Can magnifying endoscopy with narrow band imaging be useful for low grade adenomas in preoperative biopsy specimens? *Gastric Cancer*, 15(2), 170-8 (2012)

17. R Kosaka, K Tanaka, S Tano, R Takayama, K Nishikawa, Y Hamada, H Toyoda, K Ninomiya, M Katsurahara, H Inoue, N Horiki, N Katayama and Y Takei: Magnifying endoscopy for diagnosis of residual/local recurrent gastric neoplasms after previous endoscopic treatment. *Surg Endosc*, 26(8), 2299-305 (2012)

18. H Y Li, J Dai, H B Xue, Y J Zhao, X Y Chen, Y J Gao, Y Song, Z Z Ge and X B Li: Application of magnifying endoscopy with narrow-band imaging in diagnosing gastric lesions: a prospective study. *Gastrointest Endosc*, 76(6), 1124-32 (2012)

19. H Kobara, H Mori, S Fujihara, M Kobayashi, N Nishiyama, T Nomura, K Kato, S Ishihara, T Morito, K Mizobuchi, H Iwama and T Masaki: Prediction of invasion depth for submucosal differentiated gastric cancer by magnifying endoscopy with narrow-band imaging. *Oncol Rep*, 28(3), 841-7 (2012)

20. A Yokoyama, H Inoue, H Minami, Y Wada, Y Sato, H Satodate, S Hamatani and S E Kudo: Novel narrow-band imaging magnifying endoscopic classification for early gastric cancer. *Dig Liver Dis*, 42(10), 704-8 (2010)

## Modern approaches in gastric cancer assessment

21. P Pimentel-Nunes, M Dinis-Ribeiro, J B Soares, R Marcos-Pinto, C Santos, C Rolanda, R P Bastos, M Areia, L Afonso, J Bergman, P Sharma, T Gotoda, R Henrique and L Moreira-Dias: A multicenter validation of an endoscopic classification with narrow band imaging for gastric precancerous and cancerous lesions. *Endoscopy*, 44(3), 236-46 (2012)
22. M Kaise, M Kato, M Urashima, Y Arai, H Kaneyama, Y Kanzazawa, J Yonezawa, Y Yoshida, N Yoshimura, T Yamasaki, K Goda, H Imazu, H Arakawa, K Mochizuki and H Tajiri: Magnifying endoscopy combined with narrow-band imaging for differential diagnosis of superficial depressed gastric lesions. *Endoscopy*, 41(4), 310-5 (2009)
23. Y Tsuji, K Ohata, M Sekiguchi, A Ohno, T Ito, H Chiba, T Gunji, J Fukushima, N Yamamichi, M Fujishiro, N Matsuhashi and K Koike: Magnifying endoscopy with narrow-band imaging helps determine the management of gastric adenomas. *Gastric Cancer*, 15(4), 414-8 (2012)
24. K Yao: How is the VS (vessel plus surface) classification system applicable to magnifying narrow-band imaging examinations of gastric neoplasias initially diagnosed as low-grade adenomas? *Gastric Cancer*, 15(2), 118-20 (2012)
25. K Yao, G K Anagnostopoulos and K Ragnath: Magnifying endoscopy for diagnosing and delineating early gastric cancer. *Endoscopy*, 41(5), 462-7 (2009)
26. M Kobayashi, H Tajiri, E Seike, M Shitaya, S Tounou, M Mine and K Oba: Detection of early gastric cancer by a real-time autofluorescence imaging system. *Cancer Lett*, 165(2), 155-9 (2001)
27. J Haringsma, G N Tytgat, H Yano, H Iishi, M Tatsuta, T Ogihara, H Watanabe, N Sato, N Marcon, B C Wilson and R W Cline: Autofluorescence endoscopy: feasibility of detection of GI neoplasms unapparent to white light endoscopy with an evolving technology. *Gastrointest Endosc*, 53(6), 642-50 (2001)
28. N Uedo, H Iishi, M Tatsuta, T Yamada, H Ogiyama, K Imanaka, N Sugimoto, K Higashino, R Ishihara, H Narahara and S Ishiguro: A novel videoendoscopy system by using autofluorescence and reflectance imaging for diagnosis of esophagogastric cancers. *Gastrointest Endosc*, 62(4), 521-8 (2005)
29. M Kato, M Kaise, J Yonezawa, Y Yoshida and H Tajiri: Autofluorescence endoscopy versus conventional white light endoscopy for the detection of superficial gastric neoplasia: a prospective comparative study. *Endoscopy*, 39(11), 937-41 (2007)
30. M Kato, M Kaise, J Yonezawa, K Goda, H Toyozumi, N Yoshimura, Y Yoshida, M Kawamura and H Tajiri: Trimodal imaging endoscopy may improve diagnostic accuracy of early gastric neoplasia: a feasibility study. *Gastrointest Endosc*, 70(5), 899-906 (2009)
31. W L Curvers, F G van Vilsteren, L C Baak, C Bohmer, R C Mallant-Hent, A H Naber, A van Oijen, C Y Ponsioen, P Scholten, E Schenk, E Schoon, C A Seldenrijk, G A Meijer, F J ten Kate and J J Bergman: Endoscopic trimodal imaging versus standard video endoscopy for detection of early Barrett's neoplasia: a multicenter, randomized, crossover study in general practice. *Gastrointest Endosc*, 73(2), 195-203 (2011)
32. D F Boerwinkel, M Di Pietro, X Liu, M K Shariff, P Lao-Sirieix, C E Walker, M Visser, M O'Donovan, P Kaye, J J Bergman and R C Fitzgerald: Endoscopic TriModal imaging and biomarkers for neoplasia conjoined: a feasibility study in Barrett's esophagus. *Dis Esophagus* (2012)
33. S W Jung, K S Lim, J U Lim, J W Jeon, H P Shin, S H Kim, E K Lee, J J Park, J M Cha, K R Joo and J I Lee: Flexible spectral imaging color enhancement (FICE) is useful to discriminate among non-neoplastic lesion, adenoma, and cancer of stomach. *Dig Dis Sci*, 56(10), 2879-86 (2011)
34. H Osawa, H Yamamoto, Y Miura, M Yoshizawa, K Sunada, K Satoh and K Sugano: Diagnosis of extent of early gastric cancer using flexible spectral imaging color enhancement. *World J Gastrointest Endosc*, 4(8), 356-61 (2012)
35. H Osawa, H Yamamoto, Y Miura, H Ajibe, H Shinhata, M Yoshizawa, K Sunada, S Toma, K Satoh and K Sugano: Diagnosis of depressed-type early gastric cancer using small-caliber endoscopy with flexible spectral imaging color enhancement. *Dig Endosc*, 24(4), 231-6 (2012)
36. J N Zhang, Y Q Li, Y A Zhao, T Yu, J P Zhang, Y T Guo and H Liu: Classification of gastric pit patterns by confocal endomicroscopy. *Gastrointest Endosc*, 67(6), 843-53 (2008)
37. Y T Guo, Y Q Li, T Yu, T G Zhang, J N Zhang, H Liu, F G Liu, X J Xie, Q Zhu and Y A Zhao: Diagnosis of gastric intestinal metaplasia with confocal laser endomicroscopy *in vivo*: a prospective study. *Endoscopy*, 40(7), 547-53 (2008)
38. K Banno, Y Niwa, R Miyahara, M Nakamura, T Nagaya, T Nagasaka, O Watanabe, T Ando, H Kawashima, N Ohmiya, A Itoh, Y Hirooka and H Goto: Confocal endomicroscopy for phenotypic diagnosis of gastric cancer. *J Gastroenterol Hepatol*, 25(4), 712-8 (2010)
39. R Ji, X L Zuo, T Yu, X M Gu, Z Li, C J Zhou and Y Q Li: Mucosal barrier defects in gastric intestinal metaplasia: *in vivo* evaluation by confocal endomicroscopy. *Gastrointest Endosc*, 75(5), 980-7 (2012)
40. L G Lim, K G Yeoh, M Salto-Tellez, C J Khor, M Teh, Y H Chan, J B So, A Rajnakova, E Shen, S Srivastava and K Y Ho: Experienced versus inexperienced confocal endoscopists in the diagnosis of gastric adenocarcinoma

## Modern approaches in gastric cancer assessment

and intestinal metaplasia on confocal images. *Gastrointest Endosc*, 73(6), 1141-7 (2011)

41. W B Li, X L Zuo, C Q Li, F Zuo, X M Gu, T Yu, C L Chu, T G Zhang and Y Q Li: Diagnostic value of confocal laser endomicroscopy for gastric superficial cancerous lesions. *Gut*, 60(3), 299-306 (2011)

42. Z Li, T Yu, X L Zuo, X M Gu, C J Zhou, R Ji, C Q Li, P Wang, T G Zhang, K Y Ho and Y Q Li: Confocal laser endomicroscopy for *in vivo* diagnosis of gastric intraepithelial neoplasia: a feasibility study. *Gastrointest Endosc*, 72(6), 1146-53 (2010)

43. S R Jeon, W Y Cho, S Y Jin, Y K Cheon, S R Choi and J Y Cho: Optical biopsies by confocal endomicroscopy prevent additive endoscopic biopsies before endoscopic submucosal dissection in gastric epithelial neoplasias: a prospective, comparative study. *Gastrointest Endosc*, 74(4), 772-80 (2011)

44. R Ji, X L Zuo, C Q Li, C J Zhou and Y Q Li: Confocal endomicroscopy for *in vivo* prediction of completeness after endoscopic mucosal resection. *Surg Endosc*, 25(6), 1933-8 (2011)

45. Z Li, X L Zuo, C Q Li, C J Zhou, J Liu, M Goetz, R Kiesslich, K C Wu, D M Fan and Y Q Li: *In vivo* molecular imaging of gastric cancer by targeting MG7 antigen with confocal laser endomicroscopy. *Endoscopy*, 45(2), 79-85 (2013)

46. E Savarino, M Corbo, P Dulbecco, L Gemignani, E Giambruno, L Mastracci, F Grillo and V Savarino: Narrow-band imaging with magnifying endoscopy is accurate for detecting gastric intestinal metaplasia. *World J Gastroenterol*, 19(17), 2668-75 (2013)

47. N Uedo, R Ishihara, H Iishi, S Yamamoto, T Yamada, K Imanaka, Y Takeuchi, K Higashino, S Ishiguro and M Tatsuta: A new method of diagnosing gastric intestinal metaplasia: narrow-band imaging with magnifying endoscopy. *Endoscopy*, 38(8), 819-24 (2006)

48. R Pittayanon, R Rerknimitr, N Wisedopas, W Ridditid, P Kongkam, S Treeprasertsuk, P Angsuwatcharakon, V Mahachai and P Kullavanijaya: Flexible spectral imaging color enhancement plus probe-based confocal laser endomicroscopy for gastric intestinal metaplasia detection. *J Gastroenterol Hepatol*, 28(6), 1004-9 (2013)

49. M Dinis-Ribeiro, M Areia, A C de Vries, R Marcos-Pinto, M Monteiro-Soares, A O'Connor, C Pereira, P Pimentel-Nunes, R Correia, A Ensari, J M Dumonceau, J C Machado, G Macedo, P Malfertheiner, T Matysiak-Budnik, F Megraud, K Miki, C O'Morain, R M Peek, T Ponchon, A Ristimaki, B Rembacken, F Carneiro and E J Kuipers: Management of precancerous conditions and lesions in the stomach (MAPS): guideline from the European Society of Gastrointestinal Endoscopy (ESGE), European Helicobacter Study Group (EHSg), European Society of Pathology (ESP), and the Sociedade Portuguesa de

Endoscopia Digestiva (SPED). *Endoscopy*, 44(1), 74-94 (2012)

50. E Xirouchakis, F Laoudi, L Tsartsali, C Spiliadi and S D Georgopoulos: Screening for gastric premalignant lesions with narrow band imaging, white light and updated Sydney protocol or both? *Dig Dis Sci*, 58(4), 1084-90 (2013)

51. M F Dixon, R M Genta, J H Yardley and P Correa: Classification and grading of gastritis. The updated Sydney System. International Workshop on the Histopathology of Gastritis, Houston 1994. *Am J Surg Pathol*, 20(10), 1161-81 (1996)

52. M Rugge and R M Genta: Staging and grading of chronic gastritis. *Hum Pathol*, 36(3), 228-33 (2005)

53. M Rugge, M de Boni, G Pennelli, M de Bona, L Giacomelli, M Fassan, D Basso, M Plebani and D Y Graham: Gastritis OLGA-staging and gastric cancer risk: a twelve-year clinico-pathological follow-up study. *Alimentary pharmacology & therapeutics*, 31(10), 1104-11 (2010)

54. I Daugule, A Sudraba, H M Chiu, K Funke, A Ivanauskas, D Janciauskas, L Jonaitis, G Kiudelis, I Tolmanis, A Vanags, J T Lin, L Kupcinkas and M Leja: Gastric plasma biomarkers and Operative Link for Gastritis Assessment gastritis stage. *European journal of gastroenterology & hepatology*, 23(4), 302-7 (2011)

55. L G Capelle, A C de Vries, J Haringsma, F Ter Borg, R A de Vries, M J Bruno, H van Dekken, J Meijer, N C van Grieken and E J Kuipers: The staging of gastritis with the OLGA system by using intestinal metaplasia as an accurate alternative for atrophic gastritis. *Gastrointest Endosc*, 71(7), 1150-8 (2010)

56. M Rugge, M Fassan, M Pizzi, F Farinati, G C Storniolo, M Plebani and D Y Graham: Operative link for gastritis assessment vs operative link on intestinal metaplasia assessment. *World J Gastroenterol*, 17(41), 4596-601 (2011)

57. C M den Hoed, I L Holster, L G Capelle, A C de Vries, B den Hartog, F Ter Borg, K Biermann and E J Kuipers: Follow-up of premalignant lesions in patients at risk for progression to gastric cancer. *Endoscopy*, 45(4), 249-56 (2013)

58. M Fassan, M Pizzi, F Farinati, D Nitti, V Zagonel, R M Genta and M Rugge: Lesions indefinite for intraepithelial neoplasia and OLGA staging for gastric atrophy. *Am J Clin Pathol*, 137(5), 727-32 (2012)

59. M Rugge, M Fassan, M Pizzi, V Zorzetto, G Maddalo, S Realdon, M De Bernard, C Betterle, R Cappellesso, G Pennelli, M de Boni and F Farinati: Autoimmune gastritis: histology phenotype and OLGA staging. *Aliment Pharmacol Ther*, 35(12), 1460-6 (2012)

## Modern approaches in gastric cancer assessment

60. W Hoffmann: Stem Cells, Self-renewal and Cancer of the Gastric Epithelium. *Curr Med Chem* 19(35), 5975-83 (2012)
61. S Nomura, T Baxter, H Yamaguchi, C Leys, A B Vartapetian, J G Fox, J R Lee, T C Wang and J R Goldenring: Spasmolytic polypeptide expressing metaplasia to preneoplasia in H. felis-infected mice. *Gastroenterology*, 127(2), 582-94 (2004)
62. Y C Tsai, W H Hsiao, H B Yang, H C Cheng, W L Chang, C C Lu and B S Sheu: The corpus-predominant gastritis index may serve as an early marker of Helicobacter pylori-infected patients at risk of gastric cancer. *Aliment Pharmacol Ther*, 37(10), 969-78 (2013)
63. A M Halldorsdottir, M Sigurdardottrir, J G Jonasson, M Oddsdottir, J Magnusson, J R Lee and J R Goldenring: Spasmolytic polypeptide-expressing metaplasia (SPEM) associated with gastric cancer in Iceland. *Dig Dis Sci*, 48(3), 431-41 (2003)
64. H Yamaguchi, J R Goldenring, M Kaminishi and J R Lee: Identification of spasmolytic polypeptide expressing metaplasia (SPEM) in remnant gastric cancer and surveillance postgastrectomy biopsies. *Dig Dis Sci*, 47(3), 573-8 (2002)
65. K Nozaki, M Ogawa, J A Williams, B J Lafleur, V Ng, R I Drapkin, J C Mills, S F Konieczny, S Nomura and J R Goldenring: A molecular signature of gastric metaplasia arising in response to acute parietal cell loss. *Gastroenterology*, 134(2), 511-22 (2008)
66. K T Nam, R L O'Neal, R J Coffey, P E Finke, N Barker and J R Goldenring: Spasmolytic polypeptide-expressing metaplasia (SPEM) in the gastric oxyntic mucosa does not arise from Lgr5-expressing cells. *Gut* (2011)
67. T Uehara, D Ma, Y Yao, J P Lynch, K Morales, A Ziober, M Feldman, H Ota and A R Sepulveda: H. pylori infection is associated with DNA damage of Lgr5-positive epithelial stem cells in the stomach of patients with gastric cancer. *Dig Dis Sci*, 58(1), 140-9 (2013)
68. H Oshima and M Oshima: Mouse models of gastric tumors: Wnt activation and PGE2 induction. *Pathol Int*, 60(9), 599-607 (2010)
69. S D Babu, V Jayanthi, N Devaraj, C A Reis and H Devaraj: Expression profile of mucins (MUC2, MUC5AC and MUC6) in Helicobacter pylori infected pre-neoplastic and neoplastic human gastric epithelium. *Mol Cancer*, 5, 10 (2006)
70. T Tsukamoto, K Inada, H Tanaka, T Mizoshita, M Mihara, T Ushijima, Y Yamamura, S Nakamura and M Tatematsu: Down-regulation of a gastric transcription factor, Sox2, and ectopic expression of intestinal homeobox genes, Cdx1 and Cdx2: inverse correlation during progression from gastric/intestinal-mixed to complete intestinal metaplasia. *J Cancer Res Clin Oncol*, 130(3), 135-45 (2004)
71. H S Kim, J S Lee, J N Freund, K W Min, W Kim, S W Juhng and C S Park: CDX-2 homeobox gene expression in human gastric carcinoma and precursor lesions. *J Gastroenterol Hepatol*, 21(2), 438-42 (2006)
72. R Mejias-Luque, S K Linden, M Garrido, H Tye, M Najdovska, B J Jenkins, M Iglesias, M Ernst and C de Bolos: Inflammation modulates the expression of the intestinal mucins MUC2 and MUC4 in gastric tumors. *Oncogene*, 29(12), 1753-62 (2010)
73. H M Kang, N Kim, Y S Park, J H Hwang, J W Kim, S H Jeong, D H Lee, H S Lee, H C Jung and I S Song: Effects of Helicobacter pylori Infection on gastric mucin expression. *J Clin Gastroenterol*, 42(1), 29-35 (2008)
74. R Q Wang and D C Fang: Effects of Helicobacter pylori infection on mucin expression in gastric carcinoma and pericancerous tissues. *J Gastroenterol Hepatol*, 21(2), 425-31 (2006)
75. H Shiroshita, H Watanabe, Y Ajioka, G Watanabe, K Nishikura and S Kitano: Re-evaluation of mucin phenotypes of gastric minute well-differentiated-type adenocarcinomas using a series of HGM, MUC5AC, MUC6, M-GGMC, MUC2 and CD10 stains. *Pathol Int*, 54(5), 311-21 (2004)
76. K Wakatsuki, Y Yamada, M Narikiyo, M Ueno, T Takayama, H Tamaki, K Miki, S Matsumoto, K Enomoto, T Yokotani and Y Nakajima: Clinicopathological and prognostic significance of mucin phenotype in gastric cancer. *J Surg Oncol*, 98(2), 124-9 (2008)
77. K Yamamoto, M Kato, M Takahashi, M Haneda, K Shinada, U Nishida, T Yoshida, N Sonoda, S Ono, M Nakagawa, Y Mori, S Nakagawa, K Mabe, Y Shimizu, J Moriya, K Kubota, Y Matsuno, T Shimoda, H Watanabe and M Asaka: Clinicopathological analysis of early-stage gastric cancers detected after successful eradication of Helicobacter pylori. *Helicobacter*, 16(3), 210-6 (2011)
78. S E Baldus, S P Monig, V Arkenau, F G Hanisch, P M Schneider, J Thiele, A H Holscher and H P Dienes: Correlation of MUC5AC immunoreactivity with histopathological subtypes and prognosis of gastric carcinoma. *Ann Surg Oncol*, 9(9), 887-93 (2002)
79. T Shimamura, H Ito, J Shibahara, A Watanabe, Y Hippo, H Taniguchi, Y Chen, T Kashima, T Ohtomo, F Tanioka, H Iwanari, T Kodama, T Kazui, H Sugimura, M Fukayama and H Aburatani: Overexpression of MUC13 is associated with intestinal-type gastric cancer. *Cancer Sci*, 96(5), 265-73 (2005)
80. D M Maher, B K Gupta, S Nagata, M Jaggi and S C Chauhan: Mucin 13: structure, function, and potential roles in cancer pathogenesis. *Mol Cancer Res*, 9(5), 531-7 (2011)



## Modern approaches in gastric cancer assessment

81. V Barresi, E Vitarelli, M Grosso, G Tuccari and G Barresi: Relationship between immunoexpression of mucin peptide cores MUC1 and MUC2 and Lauren's histologic subtypes of gastric carcinomas. *Eur J Histochem*, 50(4), 301-9 (2006)
82. O Ilhan, U Han, B Onal and S Y Celik: Prognostic significance of MUC1, MUC2 and MUC5AC expressions in gastric carcinoma. *Turk J Gastroenterol*, 21(4), 345-52 (2010)
83. H S Han, S Y Lee, K Y Lee, S N Hong, J H Kim, I K Sung, H S Park, C J Jin and Y I Min: Unclassified mucin phenotype of gastric adenocarcinoma exhibits the highest invasiveness. *J Gastroenterol Hepatol*, 24(4), 658-66 (2009)
84. O J Lee, H J Kim, J R Kim and H Watanabe: The prognostic significance of the mucin phenotype of gastric adenocarcinoma and its relationship with histologic classifications. *Oncol Rep*, 21(2), 387-93 (2009)
85. L Xue, X Zhang, Y Li, H Yang, X Li, J Mi, H Wang, J Wang and X Yan: Differences of immunophenotypic markers and signaling molecules between adenocarcinomas of gastric cardia and distal stomach. *Hum Pathol*, 42(4), 594-601 (2011)
86. F Oz Puyan, N Can, F Ozyilmaz, U Usta, N Sut, E Tastekin and S Altaner: The relationship among PDX1, CDX2, and mucin profiles in gastric carcinomas; correlations with clinicopathologic parameters. *J Cancer Res Clin Oncol*, 137(12), 1749-62 (2011)
87. Y Tajima, K Yamazaki, R Makino, N Nishino, S Aoki, M Kato, K Morohara, T Kaetsu and M Kusano: Gastric and intestinal phenotypic marker expression in early differentiated-type tumors of the stomach: clinicopathologic significance and genetic background. *Clin Cancer Res*, 12(21), 6469-79 (2006)
88. M F Retterspitz, S P Monig, S Schreckenberger, P M Schneider, A H Holscher, H P Dienes and S E Baldus: Expression of beta-catenin, MUC1 and c-met in diffuse-type gastric carcinomas: correlations with tumour progression and prognosis. *Anticancer Res*, 30(11), 4635-41 (2010)
89. B Kocer, A Soran, G Kiyak, S Erdogan, A Eroglu, B Bozkurt, C Solak and O Cengiz: Prognostic significance of mucin expression in gastric carcinoma. *Dig Dis Sci*, 49(6), 954-64 (2004)
90. N Saeki, A Saito, I J Choi, K Matsuo, S Ohnami, H Totsuka, S Chiku, A Kuchiba, Y S Lee, K A Yoon, M C Kook, S R Park, Y W Kim, H Tanaka, K Tajima, H Hirose, F Tanioka, Y Matsuno, H Sugimura, S Kato, T Nakamura, T Nishina, W Yasui, K Aoyagi, H Sasaki, K Yanagihara, H Katai, T Shimoda, T Yoshida, Y Nakamura, S Hirohashi and H Sakamoto: A functional single nucleotide polymorphism in mucin 1, at chromosome 1q22, determines susceptibility to diffuse-type gastric cancer. *Gastroenterology*, 140(3), 892-902 (2011)
91. Y. Jia, C. Persson, L. Hou, Z. Zheng, M. Yeager, J. Lissowska, S. J. Chanock, W. H. Chow and W. Ye: A comprehensive analysis of common genetic variation in MUC1, MUC5AC, MUC6 genes and risk of stomach cancer. *Cancer Causes Control*, 21(2), 313-21 (2010)
92. F Marin, C Bonet, X Munoz, N Garcia, M L Pardo, J M Ruiz-Liso, P Alonso, G Capella, J M Sanz-Anquela, C A Gonzalez and N Sala: Genetic variation in MUC1, MUC2 and MUC6 genes and evolution of gastric cancer precursor lesions in a long-term follow-up in a high-risk area in Spain. *Carcinogenesis*, 33(5), 1072-80 (2012)
93. P Mesquita, A J Peixoto, R Seruca, C Hanski, R Almeida, F Silva, C Reis and L David: Role of site-specific promoter hypomethylation in aberrant MUC2 mucin expression in mucinous gastric carcinomas. *Cancer Lett*, 189(2), 129-36 (2003)
94. L Dardaei, R Shahsavani, A Ghavamzadeh, M Behmanesh, E Aslankoochi, K Alimoghaddam and S H Ghaffari: The detection of disseminated tumor cells in bone marrow and peripheral blood of gastric cancer patients by multimarker (CEA, CK20, TFF1 and MUC2) quantitative real-time PCR. *Clin Biochem*, 44(4), 325-30 (2011)
95. K Klaamas, O Kurtenkov, S von Mensdorff-Pouilly, L Shljapnikova, L Miljukhina, V Brjalin and A Lipping: Impact of Helicobacter pylori infection on the humoral immune response to MUC1 peptide in patients with chronic gastric diseases and gastric cancer. *Immunol Invest*, 36(4), 371-86 (2007)
96. M Kim, H J Kim, B Y Choi, J H Kim, K S Song, S M Noh, J C Kim, D S Han, S Y Kim and Y S Kim: Identification of potential serum biomarkers for gastric cancer by a novel computational method, multiple normal tissues corrected differential analysis. *Clin Chim Acta*, 413(3-4), 428-33 (2012)
97. Y J Bang, E Van Cutsem, A Feyereislova, H C Chung, L Shen, A Sawaki, F Lordick, A Ohtsu, Y Omuro, T Satoh, G Aprile, E Kulikov, J Hill, M Lehle, J Ruschoff and Y K Kang: Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): a phase 3, open-label, randomised controlled trial. *Lancet*, 376(9742), 687-97 (2010)
98. Y Kataoka, H Okabe, A Yoshizawa, S Minamiguchi, K Yoshimura, H Haga and Y Sakai: HER2 expression and its clinicopathological features in resectable gastric cancer. *Gastric Cancer* (2012)
99. C Gomez-Martin, E Garralda, M J Echarri, A Ballesteros, A Arcediano, J L Rodriguez-Peralto, M Hidalgo and F Lopez-Rios: HER2/neu testing for anti-HER2-based therapies in patients with unresectable and/or

## Modern approaches in gastric cancer assessment

- metastatic gastric cancer. *J Clin Pathol*, 65(8), 751-7 (2012)
100. K R Jones, S Jang, J Y Chang, J Kim, I S Chung, C H Olsen, D S Merrell and J H Cha: Polymorphisms in the intermediate region of VacA impact Helicobacter pylori-induced disease development. *J Clin Microbiol*, 49(1), 101-10 (2011)
101. Y Y Janjigian, D Werner, C Pauligk, K Steinmetz, D P Kelsen, E Jager, H M Altmannsberger, E Robinson, L J Tafe, L H Tang, M A Shah and S E Al-Batran: Prognosis of metastatic gastric and gastroesophageal junction cancer by HER2 status: a European and USA International collaborative analysis. *Ann Oncol*, 23(10), 2656-62 (2012)
102. T C Chua and N D Merrett: Clinicopathologic factors associated with HER2-positive gastric cancer and its impact on survival outcomes--a systematic review. *Int J Cancer*, 130(12), 2845-56 (2012)
103. B Yan, E X Yau, S S Bte Omar, C W Ong, B Pang, K G Yeoh and M Salto-Tellez: A study of HER2 gene amplification and protein expression in gastric cancer. *J Clin Pathol*, 63(9), 839-42 (2010)
104. A A Jacome, D R Wohnrath, C Scapulatempo Neto, E C Carnesecca, S V Serrano, L S Viana, J S Nunes, E Z Martinez and J S Santos: Prognostic value of epidermal growth factor receptors in gastric cancer: a survival analysis by Weibull model incorporating long-term survivors. *Gastric Cancer* (2013)
105. M Terashima, K Kitada, A Ochiai, W Ichikawa, I Kurahashi, S Sakuramoto, H Katai, T Sano, H Imamura and M Sasako: Impact of expression of human epidermal growth factor receptors EGFR and ERBB2 on survival in stage II/III gastric cancer. *Clin Cancer Res*, 18(21), 5992-6000 (2012)
106. K Shitara, Y Yatabe, K Matsuo, M Sugano, C Kondo, D Takahari, T Ura, M Tajika, S Ito and K Muro: Prognosis of patients with advanced gastric cancer by HER2 status and trastuzumab treatment. *Gastric Cancer* (2012)
107. J T Jorgensen and M Hersom: HER2 as a Prognostic Marker in Gastric Cancer - A Systematic Analysis of Data from the Literature. *J Cancer*, 3, 137-44 (2012)
108. M Fassan, L Mastracci, F Grillo, V Zagonel, S Bruno, G Battaglia, F Pitto, D Nitti, T Celiento, G Zaninotto, R Fiocca and M Ruge: Early HER2 dysregulation in gastric and oesophageal carcinogenesis. *Histopathology*, 61(5), 769-776 (2012)
109. C Bozzetti, F V Negri, C A Lagrasta, P Crafa, C Bassano, I Tamagnini, G Gardini, R Nizzoli, F Leonardi, D Gasparro, R Camisa, S Cavalli, E M Silini and A Ardizzoni: Comparison of HER2 status in primary and paired metastatic sites of gastric carcinoma. *Br J Cancer*, 104(9), 1372-6 (2011)
110. F Pagni, S Zannella, S Ronchi, C Garanzini and B E Leone: HER2 Status of Gastric Carcinoma and Corresponding Lymph Node Metastasis. *Pathol Oncol Res* (2012)
111. A F Okines, L C Thompson, D Cunningham, A Wotherspoon, J S Reis-Filho, R E Langley, T S Waddell, D Noor, Z Eltahir, R Wong and S Stenning: Effect of HER2 on prognosis and benefit from peri-operative chemotherapy in early oesophago-gastric adenocarcinoma in the MAGIC trial. *Ann Oncol*, 24(5), 1253-1261 (2013)
112. M Pirrelli, M L Caruso, M Di Maggio, R Armentano and A M Valentini: Are Biopsy Specimens Predictive of HER2 Status in Gastric Cancer Patients? *Dig Dis Sci* (2012)
113. V S Warneke, H M Behrens, C Boger, T Becker, F Lordick, M P Ebert and C Rocken: Her2/neu testing in gastric cancer: evaluating the risk of sampling errors. *Ann Oncol*, 24(3), 725-33 (2013)
114. N Fusco, E G Rocco, C Del Conte, C Pellegrini, G Bulfamante, F Di Nuovo, S Romagnoli and S Bosari: HER2 in gastric cancer: a digital image analysis in pre-neoplastic, primary and metastatic lesions. *Mod Pathol* (2013)
115. T Narita, A Seshimo, M Suzuki, J Murata and S Kameoka: Status of Tissue Expression and Serum Levels of HER2 in Gastric Cancer Patients in Japan. *Hepato-gastroenterology*, 60(125) (2013)
116. F di Mario and L G Cavallaro: Non-invasive tests in gastric diseases. *Dig Liver Dis*, 40(7), 523-30 (2008)
117. K Borch, C K Axelsson, H Halgreen, M D Damkjaer Nielsen, T Ledin and P B Szesci: The ratio of pepsinogen A to pepsinogen C: a sensitive test for atrophic gastritis. *Scand J Gastroenterol*, 24(7), 870-6 (1989)
118. K Miki: Gastric cancer screening using the serum pepsinogen test method. *Gastric Cancer*, 9(4), 245-53 (2006)
119. K Miki: Gastric cancer screening by combined assay for serum anti-Helicobacter pylori IgG antibody and serum pepsinogen levels - "ABC method". *Proc Jpn Acad Ser B Phys Biol Sci*, 87(7), 405-14 (2011)
120. M Leja, L Kupcinskas, K Funke, A Sudraba, L Jonaitis, A Ivanauskas, D Janciauskas, G Kiudelis, H M Chiu and J T Lin: The validity of a biomarker method for indirect detection of gastric mucosal atrophy versus standard histopathology. *Dig Dis Sci*, 54(11), 2377-84 (2009)
121. Y Hattori, H Tashiro, T Kawamoto and Y Kodama: Sensitivity and specificity of mass screening for gastric cancer using the measurement of serum pepsinogens. *Jpn J Cancer Res*, 86(12), 1210-5 (1995)

## Modern approaches in gastric cancer assessment

122. S Kikuchi, M Kato, T Katsuyama, S Tominaga and M Asaka: Design and planned analyses of an ongoing randomized trial assessing the preventive effect of *Helicobacter pylori* eradication on occurrence of new gastric carcinomas after endoscopic resection. *Helicobacter*, 11(3), 147-51 (2006)
123. F Kitahara, K Kobayashi, T Sato, Y Kojima, T Araki and M A Fujino: Accuracy of screening for gastric cancer using serum pepsinogen concentrations. *Gut*, 44(5), 693-7 (1999)
124. S Mizuno, M Kobayashi, S Tomita, I Miki, A Masuda, M Onoyama, Y Habu, H Inokuchi and Y Watanabe: Validation of the pepsinogen test method for gastric cancer screening using a follow-up study. *Gastric Cancer*, 12(3), 158-63 (2009)
125. K Yanaoka, M Oka, C Mukoubayashi, N Yoshimura, S Enomoto, M Iguchi, H Magari, H Utsunomiya, H Tamai, K Arai, H Ohata, M Fujishiro, T Takeshita, O Mohara and M Ichinose: Cancer high-risk subjects identified by serum pepsinogen tests: outcomes after 10-year follow-up in asymptomatic middle-aged males. *Cancer Epidemiol Biomarkers Prev*, 17(4), 838-45 (2008)
126. J M Kang, N Kim, J Y Yoo, Y S Park, D H Lee, H Y Kim, H S Lee, G Choe, J S Kim, H C Jung and I S Song: The role of serum pepsinogen and gastrin test for the detection of gastric cancer in Korea. *Helicobacter*, 13(2), 146-56 (2008)
127. J Bornschein, M Selgrad, T Wex, D Kuester and P Malfertheiner: Serological assessment of gastric mucosal atrophy in gastric cancer. *BMC Gastroenterol*, 12, 10 (2012)
128. M S Kwak, N Kim, H S Lee, H E Lee, H C Jung and I S Song: Predictive power of serum pepsinogen tests for the development of gastric cancer in comparison to the histologic risk index. *Dig Dis Sci*, 55(8), 2275-82 (2010)
129. J Bornschein, M Selgrad, M Warnecke, D Kuester, T Wex and P Malfertheiner: *H. pylori* infection is a key risk factor for proximal gastric cancer. *Dig Dis Sci*, 55(11), 3124-31 (2010)
130. A Boussioutas, H Li, J Liu, P Waring, S Lade, A J Holloway, D Taupin, K Gorringer, I Haviv, P V Desmond and D D Bowtell: Distinctive patterns of gene expression in premalignant gastric mucosa and gastric cancer. *Cancer Res*, 63(10), 2569-77 (2003)
131. P Sipponen, P Ranta, T Helske, I Kaariainen, T Maki, A Linnala, O Suovaniemi, A Alanko and M Harkonen: Serum levels of amidated gastrin-17 and pepsinogen I in atrophic gastritis: an observational case-control study. *Scand J Gastroenterol*, 37(7), 785-91 (2002)
132. H Vaananen, M Vauhkonen, T Helske, I Kaariainen, M Rasmussen, H Tunturi-Hihnala, J Koskenpato, M Sotka, M Turunen, R Sandstrom, M Ristikankare, A Jussila and P Sipponen: Non-endoscopic diagnosis of atrophic gastritis with a blood test. Correlation between gastric histology and serum levels of gastrin-17 and pepsinogen I: a multicentre study. *Eur J Gastroenterol Hepatol*, 15(8), 885-91 (2003)
133. L Agreus, E J Kuipers, L Kupcinskas, P Malfertheiner, F Di Mario, M Leja, V Mahachai, N Yaron, M van Oijen, G Perez Perez, M Rugge, J Ronkainen, M Salaspuro, P Sipponen, K Sugano and J Sung: Rationale in diagnosis and screening of atrophic gastritis with stomach-specific plasma biomarkers. *Scand J Gastroenterol*, 47(2), 136-47 (2012)
134. P Sipponen, M Harkonen, A Alanko and O Suovaniemi: Diagnosis of atrophic gastritis from a serum sample. *Minerva Gastroenterol Dietol*, 49(1), 11-21 (2003)
135. G Nardone, A Rocco, S Staibano, E Mezza, G Autiero, D Compare, G De Rosa and G Budillon: Diagnostic accuracy of the serum profile of gastric mucosa in relation to histological and morphometric diagnosis of atrophy. *Aliment Pharmacol Ther*, 22(11-12), 1139-46 (2005)
136. S Calatayud, A Alvarez and V M Victor: Gastrin: an acid-releasing, proliferative and immunomodulatory peptide? *Mini Rev Med Chem*, 10(1), 8-19 (2010)
137. P Sipponen, M Vauhkonen, T Helske, I Kaariainen and M Harkonen: Low circulating levels of gastrin-17 in patients with Barrett's esophagus. *World J Gastroenterol*, 11(38), 5988-92 (2005)
138. Y Abe, K Iijima, T Koike, K Asanuma, A Imatani, S Ohara and T Shimosegawa: Barrett's esophagus is characterized by the absence of *Helicobacter pylori* infection and high levels of serum pepsinogen I concentration in Japan. *J Gastroenterol Hepatol*, 24(1), 129-34 (2009)
139. M Leja, L Kupcinskas, K Funke, A Sudraba, L Jonaitis, A Ivanauskas, D Janciauskas, G Kuidelis, H M Chiu and J T Lin: Value of gastrin-17 in detecting antral atrophy. *Adv Med Sci*, 56(2), 145-50 (2011)
140. H Watabe, T Mitsushima, Y Yamaji, M Okamoto, R Wada, T Kokubo, H Doi, H Yoshida, T Kawabe and M Omata: Predicting the development of gastric cancer from combining *Helicobacter pylori* antibodies and serum pepsinogen status: a prospective endoscopic cohort study. *Gut*, 54(6), 764-8 (2005)
141. T Kudo, S Kakizaki, N Sohara, Y Onozato, S Okamura, Y Inui and M Mori: Analysis of ABC (D) stratification for screening patients with gastric cancer. *World J Gastroenterol*, 17(43), 4793-8 (2011)
142. K Miki, M Fujishiro, S Kodashima and N Yahagi: Long-term results of gastric cancer screening using the serum pepsinogen test method among an asymptomatic middle-aged Japanese population. *Dig Endosc*, 21(2), 78-81 (2009)

## Modern approaches in gastric cancer assessment

143. Y Oishi, Y Kiyohara, M Kubo, K Tanaka, Y Tanizaki, T Ninomiya, Y Doi, K Shikata, K Yonemoto, T Shiota, T Matsumoto and M Iida: The serum pepsinogen test as a predictor of gastric cancer: the Hisayama study. *Am J Epidemiol*, 163(7), 629-37 (2006)
144. K Yanaoka, M Oka, C Mukoubayashi, N Yoshimura, S Enomoto, M Iguchi, H Magari, H Utsunomiya, H Tamai, K Arii, H Ohata, M Fujishiro, T Takeshita, O Mohara and M Ichinose: Cancer high-risk subjects identified by serum pepsinogen tests: outcomes after 10-year follow-up in asymptomatic middle-aged males. *Cancer epidemiology, biomarkers & prevention : a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology*, 17(4), 838-45 (2008)
145. X Zhang, L Xue, L Xing, J Wang, J Cui, J Mi, X Xing, Z Du, J Misumi, Q Tian and L Wang: Low serum pepsinogen I and pepsinogen I/II ratio and Helicobacter pylori infection are associated with increased risk of gastric cancer: 14-year follow up result in a rural Chinese community. *Int J Cancer*, 130(7), 1614-9 (2012)
146. O V Reshetnikov, T G Openko, G I Simonova, S A Kurilovich, S K Maliushina, I Ragino Iu and M I Voevoda: [Risk of gastric cancer dependent on serological markers of atrophic gastritis: cohort study]. *Vopr Onkol*, 58(5), 644-8 (2012)
147. M Yoshihara, T Hiyama, S Yoshida, M Ito, S Tanaka, Y Watanabe and K Haruma: Reduction in gastric cancer mortality by screening based on serum pepsinogen concentration: a case-control study. *Scand J Gastroenterol*, 42(6), 760-4 (2007)
148. R Lomba-Viana, M Dinis-Ribeiro, F Fonseca, A S Vieira, M J Bento and H Lomba-Viana: Serum pepsinogen test for early detection of gastric cancer in a European country. *Eur J Gastroenterol Hepatol*, 24(1), 37-41 (2012)
149. H Brenner, D Rothenbacher and M N Weck: Epidemiologic findings on serologically defined chronic atrophic gastritis strongly depend on the choice of the cutoff-value. *Int J Cancer*, 121(12), 2782-6 (2007)
150. K Miki and M Fujishiro: Cautious comparison between East and West is necessary in terms of the serum pepsinogen test. *Dig Endosc*, 21(2), 134-5 (2009)
151. P Malfertheiner, F Megraud, C A O'Morain, J Atherton, A T Axon, F Bazzoli, G F Gensini, J P Gisbert, D Y Graham, T Rokkas, E M El-Omar and E J Kuipers: Management of Helicobacter pylori infection--the Maastricht IV/ Florence Consensus Report. *Gut*, 61(5), 646-64 (2012)
152. O Tureci, U Sahin and M Pfreundschuh: Serological analysis of human tumor antigens: molecular definition and implications. *Mol Med Today*, 3(8), 342-9 (1997)
153. K D Preuss, C Zwick, C Bormann, F Neumann and M Pfreundschuh: Analysis of the B-cell repertoire against antigens expressed by human neoplasms. *Immunol Rev*, 188, 43-50 (2002)
154. L Dai, N Lei, M Liu and J Y Zhang: Autoantibodies to tumor-associated antigens as biomarkers in human hepatocellular carcinoma (HCC). *Exp Hematol Oncol*, 2(1), 15 (2013)
155. P Zayakin, G Ancans, K Silina, I Meistere, Z Kalnina, D Andrejeva, E Endzelins, L Ivanova, A Pismennaja, A Ruskule, S Donina, T Wex, P Malfertheiner, M Leja and A Line: Tumor-associated autoantibody signature for the early detection of gastric cancer. *Int J Cancer*, 132(1), 137-47 (2013)
156. L A Liotta, J Kleinerman and G M Sidel: Quantitative relationships of intravascular tumor cells, tumor vessels, and pulmonary metastases following tumor implantation. *Cancer Res*, 34(5), 997-1004 (1974)
157. S J Kim, A Masago, Y Tamaki, K Akazawa, F Tsukamoto, J Sato, T Ozawa, Y Tsujino and S Noguchi: A novel approach using telomerase-specific replication-selective adenovirus for detection of circulating tumor cells in breast cancer patients. *Breast Cancer Res Treat*, 128(3), 765-73 (2011)
158. M G Krebs, R Sloane, L Priest, L Lancashire, J M Hou, A Greystoke, T H Ward, R Ferraldeschi, A Hughes, G Clack, M Ranson, C Dive and F H Blackhall: Evaluation and prognostic significance of circulating tumor cells in patients with non-small-cell lung cancer. *J Clin Oncol*, 29(12), 1556-63 (2011)
159. H Ito, H Inoue, N Sando, S Kimura, K Gohda, J Sato, K Murakami, S Ito, N Odaka, H Satodate and S E Kudo: Prognostic impact of detecting viable circulating tumour cells in gastric cancer patients using a telomerase-specific viral agent: a prospective study. *BMC Cancer*, 12(1), 346 (2012)
160. K Hiraiwa, H Takeuchi, H Hasegawa, Y Saikawa, K Suda, T Ando, K Kumagai, T Irino, T Yoshikawa, S Matsuda, M Kitajima and Y Kitagawa: Clinical significance of circulating tumor cells in blood from patients with gastrointestinal cancers. *Ann Surg Oncol*, 15(11), 3092-100 (2008)
161. L Cui, Y Lou, X Zhang, H Zhou, H Deng, H Song, X Yu, B Xiao, W Wang and J Guo: Detection of circulating tumor cells in peripheral blood from patients with gastric cancer using piRNAs as markers. *Clin Biochem*, 44(13), 1050-7 (2011)
162. H Zhou, J M Guo, Y R Lou, X J Zhang, F D Zhong, Z Jiang, J Cheng and B X Xiao: Detection of circulating tumor cells in peripheral blood from patients with gastric cancer using microRNA as a marker. *J Mol Med (Berl)*, 88(7), 709-17 (2010)
163. S Jahr, H Hentze, S Englisch, D Hardt, F O Fackelmayer, R D Hesch and R Knippers: DNA fragments

## Modern approaches in gastric cancer assessment

in the blood plasma of cancer patients: quantitations and evidence for their origin from apoptotic and necrotic cells. *Cancer Res*, 61(4), 1659-65 (2001)

164. F Diehl, K Schmidt, M A Choti, K Romans, S Goodman, M Li, K Thornton, N Agrawal, L Sokoll, S A Szabo, K W Kinzler, B Vogelstein and L A Diaz: Circulating mutant DNA to assess tumor dynamics. *Nat Med*, 14(9), 985-90 (2008)

165. T Forshew, M Murtaza, C Parkinson, D Gale, D W Tsui, F Kaper, S J Dawson, A M Piskorz, M Jimenez-Linan, D Bentley, J Hadfield, A P May, C Caldas, J D Brenton and N Rosenfeld: Noninvasive identification and monitoring of cancer mutations by targeted deep sequencing of plasma DNA. *Sci Transl Med*, 4(136), 136ra68 (2012)

166. S J Dawson, D W Tsui, M Murtaza, H Biggs, O M Rueda, S F Chin, M J Dunning, D Gale, T Forshew, B Mahler-Araujo, S Rajan, S Humphray, J Becq, D Halsall, M Wallis, D Bentley, C Caldas and N Rosenfeld: Analysis of circulating tumor DNA to monitor metastatic breast cancer. *N Engl J Med*, 368(13), 1199-209 (2013)

167. K Hibi, T Goto, A Shirahata, M Saito, G Kigawa, H Nemoto and Y Sanada: Detection of TFPI2 methylation in the serum of gastric cancer patients. *Anticancer Res*, 31(11), 3835-8 (2011)

168. C Sakakura, T Hamada, K Miyagawa, M Nishio, A Miyashita, H Nagata, H Ida, S Yazumi, E Otsuji, T Chiba, K Ito and Y Ito: Quantitative analysis of tumor-derived methylated RUNX3 sequences in the serum of gastric cancer patients. *Anticancer Res*, 29(7), 2619-25 (2009)

169. M Hakim, Y Y Broza, O Barash, N Peled, M Phillips, A Amann and H Haick: Volatile organic compounds of lung cancer and possible biochemical pathways. *Chem. Rev.*, 112, 5949-5966 (2012)

170. U Tisch, S Billan, M Ilouze, M Phillips, N Peled and H Haick: Volatile organic compounds in exhaled breath as biomarkers for the early detection and screening of lung cancer *CML-Lung Cancer*, 5, 107-117 (2012)

171. J D Pleil, M A Stiegel and T H Risby: Clinical breath analysis: discriminating between human endogenous compounds and exogenous (environmental) chemical confounders. *J Breath Res*, 7(1), 017107 (2013)

172. S Peng, M Hakim, Y Broza, S Billan, R Abdah-Brotnyak, A Kuten, U Tisch and H Haick: Detection of lung, breast, colorectal, and prostate cancers from exhaled breath using a single array of nanosensors. *Br J Cancer*, 103, 542-551 (2010)

173. D F Altomare, M Di Lena, F Porcelli, L Trizio, E Travaglio, M Tutino, S Dragonieri, V Memeo and G de Gennaro: Exhaled volatile organic compounds identify patients with colorectal cancer. *Br J Surg*, 100(1), 144-50 (2013)

174. H Amal, L Ding, B B Liu, U Tisch, Z Q Xu, D Y Shi, Y Zhao, J Chen, R X Sun, H Liu, S L Ye, Z Y Tang and H Haick: The scent fingerprint of hepatocarcinoma: in-vitro metastasis prediction with volatile organic compounds (VOCs). *Int J Nanomedicine*, 7, 4135-46 (2012)

175. Z Q Xu, Y Y Broza, R Ionsecu, U Tisch, L Ding, H Liu, Q Song, Y Y Pan, F X Xiong, K S Gu, G P Sun, Z D Chen, M Leja and H Haick: A nanomaterial-based breath test for distinguishing gastric cancer from benign gastric conditions. *Br J Cancer*, 108(4), 941-50 (2013)

176. Z Q Xu, Y Y Broza, R Ionsecu, U Tisch, L Ding, H Liu, Q Song, Y Y Pan, F X Xiong, K S Gu, G P Sun, Z D Chen, M Leja and H Haick: A nanomaterial-based breath test for distinguishing gastric cancer from benign gastric conditions. *Br. J. Cancer*, 108, 941-50 (2013)

177. V Ambros: The functions of animal microRNAs. *Nature*, 431(7006), 350-5 (2004)

178. G A Calin and C M Croce: MicroRNA signatures in human cancers. *Nat Rev Cancer*, 6(11), 857-66 (2006)

179. W Filipowicz, S N Bhattacharyya and N Sonenberg: Mechanisms of post-transcriptional regulation by microRNAs: are the answers in sight? *Nat Rev Genet*, 9(2), 102-14 (2008)

180. J Liu, M A Carmell, F V Rivas, C G Marsden, J M Thomson, J J Song, S M Hammond, L Joshua-Tor and G J Hannon: Argonaute2 is the catalytic engine of mammalian RNAi. *Science*, 305(5689), 1437-41 (2004)

181. A Goel and C R Boland: Epigenetics of colorectal cancer. *Gastroenterology*, 143(6), 1442-1460 e1 (2012)

182. A Link, J Kupcinkas, T Wex and P Malfertheiner: Macro-role of microRNA in gastric cancer. *Dig Dis* 30(3), 255-267 (2012)

183. J H Song and S J Meltzer: MicroRNAs in pathogenesis, diagnosis, and treatment of gastroesophageal cancers. *Gastroenterology*, 143(1), 35-47 (2012)

184. J Lu, G Getz, E A Miska, E Alvarez-Saavedra, J Lamb, D Peck, A Sweet-Cordero, B L Ebert, R H Mak, A A Ferrando, J R Downing, T Jacks, H R Horvitz and T R Golub: MicroRNA expression profiles classify human cancers. *Nature*, 435(7043), 834-838 (2005)

185. T Ueda, S Volinia, H Okumura, M Shimizu, C Taccioli, S Rossi, H Alder, C G Liu, N Oue, W Yasui, K Yoshida, H Sasaki, S Nomura, Y Seto, M Kaminishi, G A Calin and C M Croce: Relation between microRNA expression and progression and prognosis of gastric cancer: a microRNA expression analysis. *Lancet Oncol*, 11(2), 136-146 (2010)

186. K Matsushima, H Isomoto, N Inoue, T Nakayama, T Hayashi, M Nakayama, K Nakao, T Hirayama and S Kohno: MicroRNA signatures in *Helicobacter pylori*-

## Modern approaches in gastric cancer assessment

- infected gastric mucosa. *Int J Cancer*, 128(2), 361-70 (2011)
187. F Petrocca, R Visone, M R Onelli, M H Shah, M S Nicoloso, I de Martino, D Iliopoulos, E Pilozi, C G Liu, M Negrini, L Cavazzini, S Volinia, H Alder, L P Ruco, G Baldassarre, C M Croce and A Vecchione: E2F1-regulated microRNAs impair TGFbeta-dependent cell-cycle arrest and apoptosis in gastric cancer. *Cancer Cell*, 13(3), 272-86 (2008)
188. X Chen, Y Ba, L Ma, X Cai, Y Yin, K Wang, J Guo, Y Zhang, J Chen, X Guo, Q Li, X Li, W Wang, J Wang, X Jiang, Y Xiang, C Xu, P Zheng, J Zhang, R Li, H Zhang, X Shang, T Gong, G Ning, K Zen and C Y Zhang: Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res*, 18(10), 997-1006 (2008)
189. P S Mitchell, R K Parkin, E M Kroh, B R Fritz, S K Wyman, E L Pogosova-Agadjanyan, A Peterson, J Noteboom, K C O'Briant, A Allen, D W Lin, N Urban, C W Drescher, B S Knudsen, D L Stirewalt, R Gentleman, R L Vessella, P S Nelson, D B Martin and M Tewari: Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A*, 105(30), 10513-8 (2008)
190. L Cui, X Zhang, G Ye, T Zheng, H Song, H Deng, B Xiao, T Xia, X Yu, Y Le and J Guo: Gastric juice MicroRNAs as potential biomarkers for the screening of gastric cancer. *Cancer*, 119(9), 1618-26 (2013)
191. X Zhang, L Cui, G Ye, T Zheng, H Song, T Xia, X Yu, B Xiao, Y Le and J Guo: Gastric juice microRNA-421 is a new biomarker for screening gastric cancer. *Tumour Biol*, 33(6), 2349-55 (2012)
192. A Link, F Balaguer, Y Shen, T Nagasaka, J J Lozano, C R Boland and A Goel: Fecal MicroRNAs as novel biomarkers for colon cancer screening. *Cancer Epidemiol Biomarkers Prev*, 19(7), 1766-74 (2010)
193. A Link, V Becker, A Goel, T Wex and P Malfertheiner: Feasibility of fecal microRNAs as novel biomarkers for pancreatic cancer. *PLoS One*, 7(8), e42933 (2012)
194. L Xing, N W Todd, L Yu, H Fang and F Jiang: Early detection of squamous cell lung cancer in sputum by a panel of microRNA markers. *Mod Pathol*, 23(8), 1157-64 (2010)
195. J A Weber, D H Baxter, S Zhang, D Y Huang, K H Huang, M J Lee, D J Galas and K Wang: The microRNA spectrum in 12 body fluids. *Clin Chem*, 56(11), 1733-41 (2010)
196. M P Hunter, N Ismail, X Zhang, B D Aguda, E J Lee, L Yu, T Xiao, J Schafer, M L Lee, T D Schmittgen, S P Nana-Sinkam, D Jarjoura and C B Marsh: Detection of microRNA expression in human peripheral blood microvesicles. *PLoS One*, 3(11), e3694 (2008)
197. L Pigati, S C Yaddanapudi, R Iyengar, D J Kim, S A Hearn, D Danforth, M L Hastings and D M Duelli: Selective release of microRNA species from normal and malignant mammary epithelial cells. *PLoS One*, 5(10), e13515 (2010)
198. J D Arroyo, J R Chevillet, E M Kroh, I K Ruf, C C Pritchard, D F Gibson, P S Mitchell, C F Bennett, E L Pogosova-Agadjanyan, D L Stirewalt, J F Tait and M Tewari: Argonaute2 complexes carry a population of circulating microRNAs independent of vesicles in human plasma. *Proc Natl Acad Sci U S A*, 108(12), 5003-8 (2011)
199. B S Li, Y L Zhao, G Guo, W Li, E D Zhu, X Luo, X H Mao, Q M Zou, P W Yu, Q F Zuo, N Li, B Tang, K Y Liu and B Xiao: Plasma microRNAs, miR-223, miR-21 and miR-218, as novel potential biomarkers for gastric cancer detection. *PLoS One*, 7(7), e41629 (2012)
200. R Liu, C Zhang, Z Hu, G Li, C Wang, C Yang, D Huang, X Chen, H Zhang, R Zhuang, T Deng, H Liu, J Yin, S Wang, K Zen, Y Ba and C Y Zhang: A five-microRNA signature identified from genome-wide serum microRNA expression profiling serves as a fingerprint for gastric cancer diagnosis. *Eur J Cancer*, 47(5), 784-91 (2011)
201. M Tsujiura, D Ichikawa, S Komatsu, A Shiozaki, H Takeshita, T Kosuga, H Konishi, R Morimura, K Deguchi, H Fujiwara, K Okamoto and E Otsuji: Circulating microRNAs in plasma of patients with gastric cancers. *Br J Cancer*, 102(7), 1174-9 (2010)
202. X Yu, L Luo, Y Wu, Y Liu, X Zhao, X Zhang, L Cui, G Ye, Y Le and J Guo: Gastric juice miR-129 as a potential biomarker for screening gastric cancer. *Med Oncol*, 30(1), 365 (2013)
203. R Kogo, K Mimori, F Tanaka, S Komune and M Mori: Clinical significance of miR-146a in gastric cancer cases. *Clin Cancer Res*, 17(13), 4277-84 (2011)
204. M Okubo, T Tahara, T Shibata, H Yamashita, M Nakamura, D Yoshioka, J Yonemura, T Ishizuka, T Arisawa and I Hirata: Association between common genetic variants in pre-microRNAs and gastric cancer risk in Japanese population. *Helicobacter*, 15(6), 524-531 (2010)
205. M Y Song, H J Su, L Zhang, J L Ma, J Y Li, K F Pan and W C You: Genetic Polymorphisms of miR-146a and miR-27a, H. pylori Infection, and Risk of Gastric Lesions in a Chinese Population. *PLoS One*, 8(4), e61250 (2013)
206. T Ando, T Yoshida, S Enomoto, K Asada, M Tatematsu, M Ichinose, T Sugiyama and T Ushijima: DNA methylation of microRNA genes in gastric mucosae of gastric cancer patients: its possible involvement in the

## Modern approaches in gastric cancer assessment

- formation of epigenetic field defect. *Int J Cancer*, 124(10), 2367-74 (2009)
207. Q Chen, X Chen, M Zhang, Q Fan, S Luo and X Cao: miR-137 is frequently down-regulated in gastric cancer and is a negative regulator of Cdc42. *Dig Dis Sci*, 56(7), 2009-16 (2011)
208. K W Tsai, C W Wu, L Y Hu, S C Li, Y L Liao, C H Lai, H W Kao, W L Fang, K H Huang, W C Chan and W C Lin: Epigenetic regulation of miR-34b and miR-129 expression in gastric cancer. *Int J Cancer*, 129(11), 2600-10 (2011)
209. Z Chen, R Saad, P Jia, D Peng, S Zhu, M K Washington, Z Zhao, Z Xu and W El-Rifai: Gastric adenocarcinoma has a unique microRNA signature not present in esophageal adenocarcinoma. *Cancer*, 119(11), 1985-93 (2013)
210. C O Gigeck, E S Chen, D Q Calcagno, F Wisnieski, R R Burbano and M A Smith: Epigenetic mechanisms in gastric cancer. *Epigenomics*, 4(3), 279-94 (2012)
211. P A Jones and S B Baylin: The fundamental role of epigenetic events in cancer. *Nat Rev Genet*, 3(6), 415-28 (2002)
212. F Y Huang, A O Chan, A Rashid, D K Wong, C H Cho and M F Yuen: Helicobacter pylori induces promoter methylation of E-cadherin via interleukin-1beta activation of nitric oxide production in gastric cancer cells. *Cancer*, 118(20), 4969-4980 (2012)
213. A S Cheng, M S Li, W Kang, V Y Cheng, J L Chou, S S Lau, M Y Go, C C Lee, T K Ling, E K Ng, J Yu, T H Huang, K F To, M W Chan, J J Sung and F K Chan: Helicobacter pylori Causes Epigenetic Dysregulation of FOXD3 to Promote Gastric Carcinogenesis. *Gastroenterology* 144(1), 122-33 (2013)
214. X X Lu, J L Yu, L S Ying, J Han, S Wang, Q M Yu, X B Wang, X H Fang and Z Q Ling: Stepwise cumulation of RUNX3 methylation mediated by Helicobacter pylori infection contributes to gastric carcinoma progression. *Cancer*, 118(22), 5507-17 (2012)
215. A Ivanauskas, J Hoffmann, L V Jonaitis, R Markelis, E Juozaityte, L Kupcinskas, C Lofton-Day, C Rocken and P Malfertheiner: Distinct TPEF/HPP1 gene methylation patterns in gastric cancer indicate a field effect in gastric carcinogenesis. *Dig Liver Dis* 40(12), 920-926 (2008)
216. D Q Calcagno, C O Gigeck, E S Chen, R R Burbano and A Smith: DNA and histone methylation in gastric carcinogenesis. *World J Gastroenterol*, 19(8), 1182-92 (2013)
217. X T Hu and C He: Recent progress in the study of methylated tumor suppressor genes in gastric cancer. *Chin J Cancer*, 32(1), 31-41 (2013)
218. N S Sapari, M Loh, A Vaithilingam and R Soong: Clinical potential of DNA methylation in gastric cancer: a meta-analysis. *PLoS One*, 7(4), e36275 (2012)
219. M Fukayama: Epstein-Barr virus and gastric carcinoma. *Pathol Int*, 60(5), 337-50 (2010)
220. T Niwa and T Ushijima: Induction of epigenetic alterations by chronic inflammation and its significance on carcinogenesis. *Adv Genet*, 71, 41-56 (2010)
221. X P Zou, B Zhang, X Q Zhang, M Chen, J Cao and W J Liu: Promoter hypermethylation of multiple genes in early gastric adenocarcinoma and precancerous lesions. *Hum Pathol*, 40(11), 1534-42 (2009)
222. F M Selaru, S David, S J Meltzer and J P Hamilton: Epigenetic events in gastrointestinal cancer. *Am J Gastroenterol*, 104(8), 1910-2 (2009)
223. K Balassiano, S Lima, M Jenab, K Overvad, A Tjonneland, M C Boutron-Ruault, F Clavel-Chapelon, F Canzian, R Kaaks, H Boeing, K Meidtner, A Trichopoulou, P Laglou, P Vineis, S Panico, D Palli, S Grioni, R Tumino, E Lund, H B Bueno-de-Mesquita, M E Numans, P H Peeters, Q J Ramon, M J Sanchez, C Navarro, E Ardanaz, M Dorransoro, G Hallmans, R Stenling, R Ehrnstrom, S Regner, N E Allen, R C Travis, K T Khaw, G J Offerhaus, N Sala, E Riboli, P Hainaut, J Y Scazec, B S Sylla, C A Gonzalez and Z Herceg: Aberrant DNA methylation of cancer-associated genes in gastric cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC-EURGAST). *Cancer Lett*, 311(1), 85-95 (2011)
224. Y Shigematsu, T Niwa, S Yamashita, H Taniguchi, R Kushima, H Katai, S Ito, T Tsukamoto, M Ichinose and T Ushijima: Identification of a DNA methylation marker that detects the presence of lymph node metastases of gastric cancers. *Oncol Lett*, 4(2), 268-274 (2012)
225. D Compare, A Rocco, E Liguori, F P D'Armiento, G Persico, S Masone, E Coppola-Bottazzi, R Suriani, M Romano and G Nardone: Global DNA hypomethylation is an early event in Helicobacter pylori-related gastric carcinogenesis. *J Clin Pathol*, 64(8), 677-82 (2011)
226. Z Zheng, A F Andersson, W Ye, O Nyren, S Normark and L Engstrand: A method for metagenomics of Helicobacter pylori from archived formalin-fixed gastric biopsies permitting longitudinal studies of carcinogenic risk. *PLoS One*, 6(10), e26442 (2011)
227. O Fletcher and R S Houlston: Architecture of inherited susceptibility to common cancer. *Nat Rev Cancer*, 10(5), 353-61 (2010) doi:10.1038/nrc2840
228. A Hishida, K Matsuo, Y Goto and N Hamajima: Genetic predisposition to Helicobacter pylori-induced gastric precancerous conditions. *World Journal of Gastrointestinal Oncology*, 2(10), 369 (2010)

## Modern approaches in gastric cancer assessment

229. P Guilford, B Humar and V Blair: Hereditary diffuse gastric cancer: translation of CDH1 germline mutations into clinical practice. *Gastric Cancer*, 13(1), 1-10 (2010)
230. L Fuccio, R M Zagari, L H Eusebi, L Laterza, V Cennamo, L Ceroni, D Grilli and F Bazzoli: Meta-analysis: can *Helicobacter pylori* eradication treatment reduce the risk for gastric cancer? *Ann Intern Med*, 151(2), 121-128 (2009)
231. T Watanabe, M Tada, H Nagai, S Sasaki and M Nakao: *Helicobacter pylori* infection induces gastric cancer in Mongolian gerbils. *Gastroenterology*, 115(3), 642-648 (1998)
232. E M El-Omar, M Carrington, W H Chow, K E McColl, J H Bream, H A Young, J Herrera, J Lissowska, C C Yuan, N Rothman, G Lanyon, M Martin, J F Fraumeni and C S Rabkin: Interleukin-1 polymorphisms associated with increased risk of gastric cancer. *Nature*, 404(6776), 398-402 (2000)
233. A Hishida, K Matsuo, Y Goto, M Naito, K Wakai, K Tajima and N Hamajima: No associations of Toll-like receptor 2 (TLR2) -196 to -174del polymorphism with the risk of *Helicobacter pylori* seropositivity, gastric atrophy, and gastric cancer in Japanese. *Gastric Cancer*, 13(4), 251-7 (2010)
234. J Kupcinskas, T Wex, J Bornschein, M Selgrad, M Leja, E Juozaityte, G Kiudelis, L Jonaitis and P Malfertheiner: Lack of association between gene polymorphisms of Angiotensin converting enzyme, Nod-like receptor 1, Toll-like receptor 4, FAS/FASL and the presence of *Helicobacter pylori*-induced premalignant gastric lesions and gastric cancer in Caucasians. *BMC Med Genet*, 12, 112 (2011)
235. L Kupcinskas, T Wex, J Kupcinskas, M Leja, A Ivanauskas, L V Jonaitis, D Janciauskas, G Kiudelis, K Funka, A Sudraba, H M Chiu, J T Lin and P Malfertheiner: Interleukin-1B and interleukin-1 receptor antagonist gene polymorphisms are not associated with premalignant gastric conditions: a combined haplotype analysis. *Eur J Gastroenterol Hepatol*, 22(10), 1189-1195 (2010)
236. T Wex, A Leodolter, J Bornschein, D Kuester, T Kahne, S Kropf, C Albrecht, M Naumann, A Roessner and P Malfertheiner: Interleukin 1 beta (IL1B) gene polymorphisms are not associated with gastric carcinogenesis in Germany. *Anticancer Res*, 30(2), 505-11 (2010)
237. E M El Omar, M Carrington, W H Chow, K E L McColl, J H Bream, H A Young, J Herrera, J Lissowska, C C Yuan and N Rothman: Interleukin-1 polymorphisms associated with increased risk of gastric cancer. *Nature*, 404, 398-402 (2000)
238. T Tahara, T Shibata, H Yamashita, D Yoshioka, M Okubo, J Yonemura, Y Kamiya, T Ishizuka, Y Nakagawa, M Nagasaka, M Iwata, M Nakamura, I Hirata and T Arisawa: Synergistic effect of IL-1beta and TNF-alpha polymorphisms on the *H. pylori*-related gastric premalignant condition. *Hepatogastroenterology*, 59(120), 2416-20 (2012)
239. C A Yang, C Scheibenbogen, S Bauer, C Kleinle, T Wex, J Bornschein, P Malfertheiner, S Hellmig, R R Schumann and L Hamann: A frequent Toll-like receptor 1 gene polymorphism affects NK- and T-cell IFN-gamma production and is associated with *Helicobacter pylori*-induced gastric disease. *Helicobacter*, 18(1), 13-21 (2013)
240. M C Camargo, R Mera, P Correa, R M Peek, E T Fonham, K J Goodman, M B Piazuelo, L Sicinski, J Zabaleta and B G Schneider: Interleukin-1beta and interleukin-1 receptor antagonist gene polymorphisms and gastric cancer: a meta-analysis. *Cancer Epidemiol Biomarkers Prev*, 15(9), 1674-87 (2006)
241. F Kamangar, C Cheng, C C Abnet and C S Rabkin: Interleukin-1B polymorphisms and gastric cancer risk: a meta-analysis. *Cancer Epidemiology Biomarkers & Prevention*, 15(10), 1920-1928 (2006)
242. B Peleteiro, N Lunet, C Carrilho, C Duraes, J C Machado, C La Vecchia and H Barros: Association between cytokine gene polymorphisms and gastric precancerous lesions: systematic review and meta-analysis. *Cancer Epidemiology Biomarkers & Prevention*, 19(3), 762-776 (2010)
243. B Vincenzi, G Patti, S Galluzzo, F Pantano, O Venditti, D Santini, A Ruzzo, G Schiavon, M Caraglia, M Marra, F Graziano and G Tonini: Interleukin 1beta-511T gene (IL1beta) polymorphism is correlated with gastric cancer in the Caucasian population: results from a meta-analysis. *Oncol Rep*, 20(5), 1213-20 (2008)
244. N Saeki, H Ono, H Sakamoto and T Yoshida: Genetic factors related to gastric cancer susceptibility identified using a genome-wide association study. *Cancer Sci*, 104(1), 1-8 (2013)
245. Y Shi, Z Hu, C Wu, J Dai, H Li, J Dong, M Wang, X Miao, Y Zhou, F Lu, H Zhang, L Hu, Y Jiang, Z Li, M Chu, H Ma, J Chen, G Jin, W Tan, T Wu, Z Zhang, D Lin and H Shen: A genome-wide association study identifies new susceptibility loci for non-cardia gastric cancer at 3q13.31 and 5p13.1. *Nat Genet*, 43(12), 1215-8 (2011)
246. P Lochhead, B Frank, G L Hold, C S Rabkin, M T Ng, T L Vaughan, H A Risch, M D Gammon, J Lissowska, M N Weck, E Raum, H Muller, T Illig, N Klopp, A Dawson, K E McColl, H Brenner, W H Chow and E M El-Omar: Genetic variation in the prostate stem cell antigen gene and upper gastrointestinal cancer in white individuals. *Gastroenterology*, 140(2), 435-41 (2011)
247. J Bornschein, A Kandulski, M Selgrad and P Malfertheiner: From gastric inflammation to gastric cancer. *Dig Dis*, 28(4-5), 609-14 (2010)



## Modern approaches in gastric cancer assessment

248. E Segal, N Friedman, N Kaminski, A Regev and D Koller: From signatures to models: understanding cancer using microarrays. *Nat Genet*, 37 Suppl, S38-45 (2005)
249. E Segal, M Shapira, A Regev, D Pe'er, D Botstein, D Koller and N Friedman: Module networks: identifying regulatory modules and their condition-specific regulators from gene expression data. *Nat Genet*, 34(2), 166-76 (2003)
250. K Garber: Genomic medicine. Gene expression tests foretell breast cancer's future. *Science*, 303(5665), 1754-5 (2004)
251. I Drozdov, C A Ouzounis, A M Shah and S Tsoka: Functional Genomics Assistant (FUGA): a toolbox for the analysis of complex biological networks. *BMC Res Notes*, 4(1), 462 (2011)
252. V K Mootha, C M Lindgren, K F Eriksson, A Subramanian, S Sihag, J Lehar, P Puigserver, E Carlsson, M Ridderstrale, E Laurila, N Houstis, M J Daly, N Patterson, J P Mesirov, T R Golub, P Tamayo, B Spiegelman, E S Lander, J N Hirschhorn, D Altshuler and L C Groop: PGC-1 $\alpha$ -responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes. *Nat Genet*, 34(3), 267-73 (2003)
253. M Ronen, R Rosenberg, B I Shraiman and U Alon: Assigning numbers to the arrows: parameterizing a gene regulation network by using accurate expression kinetics. *Proc Natl Acad Sci U S A*, 99(16), 10555-60 (2002)
254. I Drozdov, S Tsoka, C A Ouzounis and A M Shah: Genome-wide expression patterns in physiological cardiac hypertrophy. *BMC Genomics*, 11, 557 (2010)
255. R Guimera and L A Nunes Amaral: Functional cartography of complex metabolic networks. *Nature*, 433(7028), 895-900 (2005)
256. S M Kim, S H Leem, I S Chu, Y Y Park, S C Kim, S B Kim, E S Park, J Y Lim, J Heo, Y J Kim, D G Kim, A Kaseb, Y N Park, X W Wang, S S Thorgeirsson and J S Lee: 65-gene-based risk score classifier predicts overall survival in hepatocellular carcinoma. *Hepatology*, 55(5), 1443-52 (2012)
257. I Drozdov, M Kidd, B Nadler, R L Camp, S M Mane, O Hauso, B I Gustafsson and I M Modlin: Predicting neuroendocrine tumor (carcinoid) neoplasia using gene expression profiling and supervised machine learning. *Cancer*, 115(8), 1638-50 (2009)
258. I Drozdov, B Svejda, B I Gustafsson, S Mane, R Pfragner, M Kidd and I M Modlin: Gene network inference and biochemical assessment delineates GPCR pathways and CREB targets in small intestinal neuroendocrine neoplasia. *PLoS One*, 6(8), e22457 (2011)
259. I M Modlin, B I Gustafsson, I Drozdov, B Nadler, R Pfragner and M Kidd: Principal component analysis, hierarchical clustering, and decision tree assessment of plasma mRNA and hormone levels as an early detection strategy for small intestinal neuroendocrine (carcinoid) tumors. *Ann Surg Oncol*, 16(2), 487-98 (2009)
260. A J Enright, S Van Dongen and C A Ouzounis: An efficient algorithm for large-scale detection of protein families. *Nucleic Acids Res*, 30(7), 1575-84 (2002)
261. E Birney, T D Andrews, P Bevan, M Caccamo, Y Chen, L Clarke, G Coates, J Cuff, V Curwen, T Cutts, T Down, E Eyra, X M Fernandez-Suarez, P Gane, B Gibbins, J Gilbert, M Hammond, H R Hotz, V Iyer, K Jekosch, A Kahari, A Kasprzyk, D Keefe, S Keenan, H Lehtvaslainen, G McVicker, C Melsopp, P Meidl, E Mongin, R Pettett, S Potter, G Proctor, M Rae, S Searle, G Slater, D Smedley, J Smith, W Spooner, A Stabenau, J Stalker, R Storey, A Ureta-Vidal, K C Woodwark, G Cameron, R Durbin, A Cox, T Hubbard and M Clamp: An overview of Ensembl. *Genome Res*, 14(5), 925-8 (2004)
262. K Furuta, T Arao, K Sakai, H Kimura, T Nagai, D Tamura, K Aomatsu, K Kudo, H Kaneda, Y Fujita, K Matsumoto, Y Yamada, K Yanagihara, M Sekijima and K Nishio: Integrated analysis of whole genome exon array and array-comparative genomic hybridization in gastric and colorectal cancer cells. *Cancer Sci*, 103(2), 221-7 (2012)
263. Y Z Zhang, L H Zhang, Y Gao, C H Li, S Q Jia, N Liu, F Cheng, D Y Niu, W C Cho, J F Ji and C Q Zeng: Discovery and validation of prognostic markers in gastric cancer by genome-wide expression profiling. *World J Gastroenterol*, 17(13), 1710-7 (2011)
264. E Rossi, C Klersy, R Manca, O Zuffardi and E Solcia: Correlation between genomic alterations assessed by array comparative genomic hybridization, prognostically informative histologic subtype, stage, and patient survival in gastric cancer. *Hum Pathol*, 42(12), 1937-45 (2011)
265. L Cheng, P Wang, S Yang, Y Yang, Q Zhang, W Zhang, H Xiao and H Gao: Identification of genes with a correlation between copy number and expression in gastric cancer. *BMC Med Genomics*, 5, 14 (2012)
266. A Kuroda, Y Tsukamoto, L T Nguyen, T Noguchi, I Takeuchi, M Uchida, T Uchida, N Hijiya, C Nakada, T Okimoto, M Kodama, K Murakami, K Matsuura, M Seto, H Ito, T Fujioka and M Moriyama: Genomic profiling of submucosal-invasive gastric cancer by array-based comparative genomic hybridization. *PLoS One*, 6(7), e22313 (2011)
267. N Deng, L K Goh, H Wang, K Das, J Tao, I B Tan, S Zhang, M Lee, J Wu, K H Lim, Z Lei, G Goh, Q Y Lim, A L Tan, D Y Sin Poh, S Riahi, S Bell, M M Shi, R Linnartz, F Zhu, K G Yeoh, H C Toh, W P Yong, H C Cheong, S Y Rha, A Boussioutas, H Grabsch, S Rozen and P Tan: A comprehensive survey of genomic alterations in gastric cancer reveals systematic patterns of molecular exclusivity and co-occurrence among distinct therapeutic targets. *Gut*, 61(5), 673-84 (2012)

## Modern approaches in gastric cancer assessment

268. A M Dulak, S E Schumacher, J van Lieshout, Y Imamura, C Fox, B Shim, A H Ramos, G Saksena, S C Baca, J Baselga, J Tabernero, J Barretina, P C Enzinger, G Corso, F Roviello, L Lin, S Bandla, J D Luketich, A Pennathur, M Meyerson, S Ogino, R A Shivdasani, D G Beer, T E Godfrey, R Beroukhim and A J Bass: Gastrointestinal adenocarcinomas of the esophagus, stomach, and colon exhibit distinct patterns of genome instability and oncogenesis. *Cancer Res*, 72(17), 4383-93 (2012)
269. Y Zhang, S Wang, D Li, J Zhnag, D Gu, Y Zhu and F He: A systems biology-based classifier for hepatocellular carcinoma diagnosis. *PLoS One*, 6(7), e22426 (2011)
270. J Saez-Rodriguez, L G Alexopoulos, M Zhang, M K Morris, D A Lauffenburger and P K Sorger: Comparing signaling networks between normal and transformed hepatocytes using discrete logical models. *Cancer Res*, 71(16), 5400-11 (2011)
271. X Y Goh, J R Rees, A L Paterson, S F Chin, J C Marionni, V Save, M O'Donovan, P P Eijk, D Alderson, B Ylstra, C Caldas and R C Fitzgerald: Integrative analysis of array-comparative genomic hybridisation and matched gene expression profiling data reveals novel genes with prognostic significance in oesophageal adenocarcinoma. *Gut*, 60(10), 1317-26 (2011)
272. A Aggarwal, D L Guo, Y Hoshida, S T Yuen, K M Chu, S So, A Boussioutas, X Chen, D Bowtell, H Aburatani, S Y Leung and P Tan: Topological and functional discovery in a gene coexpression meta-network of gastric cancer. *Cancer Res*, 66(1), 232-41 (2006)
273. J Cui, F Li, G Wang, X Fang, J D Puett and Y Xu: Gene-expression signatures can distinguish gastric cancer grades and stages. *PLoS One*, 6(3), e17819 (2011)
274. J Y Cho, J Y Lim, J H Cheong, Y Y Park, S L Yoon, S M Kim, S B Kim, H Kim, S W Hong, Y N Park, S H Noh, E S Park, I S Chu, W K Hong, J A Ajani and J S Lee: Gene expression signature-based prognostic risk score in gastric cancer. *Clin Cancer Res*, 17(7), 1850-7 (2011)
275. S M Kim, Y Y Park, E S Park, J Y Cho, J G Izzo, D Zhang, S B Kim, J H Lee, M S Bhutani, S G Swisher, X Wu, K R Coombes, D Maru, K K Wang, N S Buttar, J A Ajani and J S Lee: Prognostic biomarkers for esophageal adenocarcinoma identified by analysis of tumor transcriptome. *PLoS One*, 5(11), e15074 (2010)
276. C J Peters, J R Rees, R H Hardwick, J S Hardwick, S L Vowler, C A Ong, C Zhang, V Save, M O'Donovan, D Rassl, D Alderson, C Caldas and R C Fitzgerald: A 4-gene signature predicts survival of patients with resected adenocarcinoma of the esophagus, junction, and gastric cardia. *Gastroenterology*, 139(6), 1995-2004 e15 (2010)
277. A Saadi, N B Shannon, P Lao-Sirieix, M O'Donovan, E Walker, N J Clemons, J S Hardwick, C Zhang, M Das, V Save, M Novelli, F Balkwill and R C Fitzgerald: Stromal genes discriminate preinvasive from invasive disease, predict outcome, and highlight inflammatory pathways in digestive cancers. *Proc Natl Acad Sci U S A*, 107(5), 2177-82 (2010)
278. C H Ooi, T Ivanova, J Wu, M Lee, I B Tan, J Tao, L Ward, J H Koo, V Gopalakrishnan, Y Zhu, L L Cheng, J Lee, S Y Rha, H C Chung, K Ganesan, J So, K C Soo, D Lim, W H Chan, W K Wong, D Bowtell, K G Yeoh, H Grabsch, A Boussioutas and P Tan: Oncogenic pathway combinations predict clinical prognosis in gastric cancer. *PLoS Genet*, 5(10), e1000676 (2009)
279. Y Y Wang, Z Y Ye, Z S Zhao, H Q Tao and S G Li: Systems biology approach to identification of biomarkers for metastatic progression in gastric cancer. *J Cancer Res Clin Oncol*, 136(1), 135-41 (2010)
280. M A Shah, R Khanin, L Tang, Y Y Janjigian, D S Klimstra, H Gerdes and D P Kelsen: Molecular classification of gastric cancer: a new paradigm. *Clin Cancer Res*, 17(9), 2693-701 (2011)
281. G Wang, N Hu, H H Yang, L Wang, H Su, C Wang, R Clifford, E M Dawsey, J M Li, T Ding, X Y Han, C Giffen, A M Goldstein, P R Taylor and M P Lee: Comparison of global gene expression of gastric cardia and noncardia cancers from a high-risk population in china. *PLoS One*, 8(5), e63826 (2013)
282. Z Lei, I B Tan, K Das, N Deng, H Zouridis, S Pattison, C Chua, Z Feng, Y K Guan, C H Ooi, T Ivanova, S Zhang, M Lee, J Wu, A Ngo, S Manesh, E Tan, B T Teh, J B Yan So, L K Goh, A Boussioutas, T K Hon Lim, H Flotow, P Tan and S G Rozen: Identification of Molecular Subtypes of Gastric Cancer with Different Responses to PI3-Kinase Inhibitors and 5-Fluorouracil. *Gastroenterology* (2013)
283. H Wong and T Yau: Molecular targeted therapies in advanced gastric cancer: does tumor histology matter? *Therap Adv Gastroenterol*, 6(1), 15-31 (2013)
284. D Wang, F Ye, Y Sun, W Li, H Liu, J Jiang, Y Zhang, C Liu, W Tong, L Gao, W Zhang, T Seetoe, P Lee, J Suo and D Y Zhang: Protein signatures for classification and prognosis of gastric cancer a signaling pathway-based approach. *Am J Pathol*, 179(4), 1657-66 (2011)
285. J Qian, F Li, J Yu, Z Yang and J Chen: Mining predictive biomarker for neoadjuvant chemotherapy in gastric cancer by proteomics. *Hepatogastroenterology*, 58(110-111), 1828-33 (2011)
286. D Torti, F Sassi, F Galimi, S Gastaldi, T Perera, P M Comoglio, L Trusolino and A Bertotti: A preclinical algorithm of soluble surrogate biomarkers that correlate with therapeutic inhibition of the MET oncogene in gastric tumors. *Int J Cancer*, 130(6), 1357-66 (2012)
287. S O Deiningner, M P Ebert, A Futterer, M Gerhard and C Rocken: MALDI imaging combined with hierarchical

## Modern approaches in gastric cancer assessment

clustering as a new tool for the interpretation of complex human cancers. *J Proteome Res*, 7(12), 5230-6 (2008)

288. S Meding, U Nitsche, B Balluff, M Elsner, S Rauser, C Schone, M Nipp, M Maak, M Feith, M P Ebert, H Friess, R Langer, H Hofler, H Zitzelsberger, R Rosenberg and A Walch: Tumor classification of six common cancer types based on proteomic profiling by MALDI imaging. *J Proteome Res*, 11(3), 1996-2003 (2012)

289. W Liu, B Liu, Q Cai, J Li, X Chen and Z Zhu: Proteomic identification of serum biomarkers for gastric cancer using multi-dimensional liquid chromatography and 2D differential gel electrophoresis. *Clin Chim Acta*, 413(13-14), 1098-106 (2012)

290. K M Fock, P Katelaris, K Sugano, T L Ang, R Hunt, N J Talley, S K Lam, S D Xiao, H J Tan, C Y Wu, H C Jung, B H Hoang, U Kachintorn, K L Goh, T Chiba and A A Rani: Second Asia-Pacific Consensus Guidelines for Helicobacter pylori infection. *Journal of gastroenterology and hepatology*, 24(10), 1587-600 (2009)

**Key Words:** Gastric cancer, endoscopic imaging, Her2, pepsinogens, miRNA, array comparative genomic hybridisation, mucins, Review

**Send correspondence to:** Peter Malfertheiner, Dept. of Gastroenterology, Hepatology and Infectious Diseases, Otto-von-Guericke University, Leipziger Str. 44, 39120 Magdeburg, Germany, Tel: 0049-391-6713100, Fax: 0049-391-6713105, E-mail: peter.malfertheiner@med.ovgu.de