

Clinical applications of coenzyme Q10

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1. ABSTRACT

Coenzyme Q10 (CoQ10) or ubiquinone was known for its key role in mitochondrial bioenergetics as electron and proton carrier; later studies demonstrated its presence in other cellular membranes and in blood plasma, and extensively investigated its antioxidant role. These two functions constitute the basis for supporting the clinical indication of CoQ10. Furthermore, recent data indicate that CoQ10 affects expression of genes involved in human cell signalling, metabolism and transport and some of the effects of CoQ10 supplementation may be due to this property. CoQ10 deficiencies are due to autosomal recessive mutations, mitochondrial diseases, ageing-related

oxidative stress and carcinogenesis processes, and also a secondary effect of statin treatment. Many neurodegenerative disorders, diabetes, cancer, fibromyalgia, muscular and cardiovascular diseases have been associated with low CoQ10 levels. CoQ10 treatment does not cause serious adverse effects in humans and new formulations have been developed that increase CoQ10 absorption and tissue distribution. Oral CoQ10 treatment is a frequent mitochondrial energizer and antioxidant strategy in many diseases that may provide a significant symptomatic benefit.

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Table 1. Most frequent physiologic and clinical applications of coenzyme Q₁₀

Physiologic and clinical applications	References
Human coenzyme Q ₁₀ deficiencies	(34)
Mitochondrial diseases	(40)
Fibromyalgia	(51, 52)
Cardiac failure	(139)
Ischemic heart disease	(140)
Interaction with statins	(141)
Hypertension	(73)
Diabetes	(102)
Endothelial function	(142)
Pre-eclampsia	(117)
Neurodegenerative diseases	
Parkinson's disease	(143)
Huntington's disease	(86)
Alzheimer's disease	(88)
Friedreich's ataxia	(144)
Cancer	(99)
Aging	(120)
Other pathological conditions	
Migraine	(145)
Down's syndrome	(119)
Periodontal Disease	(112)
Asthenozoospermia	(108)

2. INTRODUCTION

Coenzyme Q₁₀ (CoQ₁₀) is an essential compound found naturally in virtually every cell in the human body. Because of its ubiquitous presence in nature and its quinone structure CoQ₁₀ is also known as ubiquinone. It is found in cell membranes and is particularly well known for its role in the electron transport chain in mitochondrial membranes during aerobic cellular respiration. Adequate amounts of CoQ₁₀ are necessary for cellular respiration and ATP production. CoQ₁₀ also functions as an intercellular antioxidant and its presence was then demonstrated in all cell membranes and in blood both, in high- and in low-density lipoproteins, where it is endowed with antioxidant properties (1). CoQ₁₀ was also recognized to have an effect on gene expression that might account for its effects on overall tissue metabolism (2, 3).

Although the chemical structure of CoQ₁₀ is similar to that of vitamin K, CoQ₁₀ is not considered a vitamin because it is the only lipid-soluble antioxidant that animal cells synthesize *de novo* in the body (4). Cells generally rely on biosynthesis for their supply of CoQ₁₀. Endogenous levels are subject to regulation by physiological factors that are related to the oxidative activity of the organism (5, 6). Para-hydroxybenzoic acid from the amino acid tyrosine is the first aromatic precursor in the biosynthetic pathway of CoQ₁₀ in humans and constitutes the quinoid ring structure of the CoQ₁₀ molecule. The tail, consisting of 10 isoprenoid units, is derived from the mevalonate pathway (7). Endogenous CoQ₁₀ levels are determined by both the rate of production and the rate of consumption in the body. These levels can be altered in a number of disease states, among which cardiovascular disease and degenerative muscle disorders have been well documented in humans (8).

Dietary supplementation affecting CoQ₁₀ levels has been shown in a number of organisms to cause multiple phenotypic effects, which can be explained on the basis of its significant impact on the expression of many genes

mainly involved in cell signalling, intermediary metabolism, transport and transcription control and inflammation, among others, indicating an important role for CoQ₁₀ as a potent gene regulator (2, 9). However, the molecular mechanisms whereby CoQ₁₀ is inducing these pleiotropic effects has yet to be completely understood (3).

Numerous disease processes, associated with CoQ₁₀ deficiency, can benefit from CoQ₁₀ supplementation including primary and secondary CoQ₁₀ deficiencies, mitochondrial diseases, fibromyalgia, cardiovascular disease, neurodegenerative diseases, cancer, diabetes mellitus, male infertility and periodontal disease (Table 1).

Tissue deficiencies or subnormal serum levels of CoQ₁₀ have been reported in a wide range of medical conditions, including primary CoQ₁₀ deficiencies (10) and secondary CoQ₁₀ deficiencies such as, mitochondrial diseases (11). CoQ₁₀ levels decline with advancing age, and this decline might contribute in part to some of the manifestations of aging (12). CoQ₁₀ deficiency could result from: (1) impaired CoQ₁₀ synthesis due to nutritional deficiencies (such as vitamin B6 deficiency, a cofactor essential for CoQ₁₀ biosynthesis), (2) a genetic or acquired defect in CoQ₁₀ synthesis or utilization, or (3) increased tissue needs resulting from a particular disease. Clinical presentations of severe CoQ₁₀ deficiency include encephalomyopathy, severe infantile multisystemic disease, cerebellar ataxia, Leigh syndrome with growth retardation and isolated myopathy. Since oral administration of CoQ₁₀ can increase tissue levels of the nutrient, it is possible to correct CoQ₁₀ deficiency and is particularly essential in the life-threatening infantile encephalopathy (13).

3. ABSORPTION, TISSUE UPTAKE AND PHARMACOKINETICS

Plasma CoQ₁₀ concentrations are usually used for the estimation of CoQ₁₀ status in humans primarily because of the ease of sample collection. Reported plasma CoQ₁₀ ranged from 0.40 to 1.91 µmol/l (0.34-1.65 µg/ml) (4).

Table 2. Functions of coenzyme Q₁₀

Function	References
Electron and proton carrier in the mitochondrial respiratory chain	(146)
Participation in extra-mitochondrial electron transport (plasma membranes, lysosomes)	(147, 148)
Endogenously synthesized, lipid-soluble antioxidant	(5, 149)
Regulation of mitochondrial permeability transition pores	(150)
Required for activation of mitochondrial uncoupling proteins	(151)
CoQ ₁₀ exerts multiple anti-inflammatory effects by influencing the expression of NFκ-B ¹ -dependent genes	(152)
Regulation of the physicochemical properties of membranes	(8)
By protecting LDL ² from oxidation, this lipid also has anti-atherosclerotic properties	(153)
Modulation of the amount of h2-integrins on the surface of blood monocytes which counteracts monocyte-endothelial cell interactions	(154)
Improvement of endothelial dysfunction (probably by increasing nitric oxide)	(155)
It is required for the biosynthesis of pyrimidine nucleotides because it is an essential co-factor for dihydro-orotate dehydrogenase	(156)
Mitophagy modulator	(45)
Inflammasome modulator	(157)

Abbreviations: Nuclear Transcription Factor-kappa B¹; Low Density Lipoprotein².

However, these measurements reflect dietary intake rather than tissue status. Moreover, the relationship between plasma and tissue CoQ₁₀ levels is not yet clear, and plasma levels should only be regarded as a surrogate for tissue (14), and in particular mitochondrial levels, where any therapeutic effect of CoQ₁₀ may be expected to be most important. The primary problem with measuring tissue levels is access to tissue samples. Skin fibroblasts, muscle biopsies, and blood mononuclear cells (BMCs) may reflect better actual tissue CoQ₁₀ levels. Blood cells have been used for estimates of CoQ₁₀ in tissues (15). CoQ₁₀ content of BMCs was shown to correlate with skeletal muscle CoQ₁₀ in un-supplemented subjects whereas the plasma concentrations did not (16). There would appear to be no clinical value in measuring erythrocyte CoQ₁₀, but there may be a possible case for considering its measurement in platelets or other mitochondria-containing blood cells such as BMCs, though pertinent reference ranges would need to be established (17).

CoQ₁₀ is naturally found in dietary sources, with large amounts present in heart, chicken leg, herring and trout. The daily intake from food was estimated to be 3-5 mg CoQ₁₀ a day. However, in tissues with unimpaired synthetic capacity, it appears that CoQ₁₀ reaches a saturation level, and nutritional supplement of CoQ₁₀ in the diet does not increase tissue levels above normal (18, 19). Intestinal absorption is threefold faster if CoQ₁₀ is administered with food intake (20). Following absorption, CoQ₁₀ appears in plasma lipoproteins and in liver, but usually not in heart or kidney (21). However, with higher supplementations (150 mg/kg/d), heart and the skeletal muscle showed a significant increase in total CoQ₁₀ suggesting that higher plasma CoQ₁₀ concentrations are necessary to facilitate uptake by peripheral tissues (22). Biochemical characteristics of CoQ₁₀ are important for our understanding of uptake and distribution following oral ingestion. CoQ₁₀ is absorbed slowly from the small intestine, possibly because it has a high molecular weight and is not very water soluble, passes into the lymphatics, and finally to the blood and tissues. Research on exogenous CoQ₁₀ absorption and bioavailability varies greatly depending on the type of CoQ₁₀ preparation studied. CoQ₁₀ absorption is probably a complex process and dependent upon active and passive transport mechanisms (23). A

study on intestinal absorption of 30 mg CoQ₁₀ administered in a meal or as powder in capsules to healthy subjects found no significant difference in absorption for these two routes of administration (24). Although not all research is in agreement, the general consensus is that slightly better absorption is achieved with oil-based forms of CoQ₁₀ (25, 26). Further studies are needed to elucidate whether age, gender, lipoprotein status, diet, dosage formulation, or other factors may affect the bioavailability of CoQ₁₀ with chronic dosing (27).

CoQ₁₀ dosage guidelines, which appeared to be safe and well tolerated, were suggested for adults (up to 1,200 mg/day) (28) and for children (up to 10 mg/kg/day) (29), although higher doses are recommended in particular pathological conditions. Monitoring trough CoQ₁₀ plasma concentrations may be considered after 3-4 weeks of constant dosing, when steady-state conditions exist (30). Steady-state plasma concentrations at these dosage levels generally ranged from 5 to 10 µg/mL (27).

4. MECHANISM OF ACTION

Due to its involvement in ATP synthesis, CoQ₁₀ affects the function of all cells in the body, especially those with high-energy demand, making it essential for the health of all tissues and organs. CoQ₁₀ is our only lipid-soluble antioxidant synthesized endogenously and efficiently prevents oxidation of proteins, lipids and DNA. The fundamental role of CoQ₁₀ in mitochondrial bioenergetics and its well-acknowledged antioxidant properties constitute the basis for its clinical applications, although some of its effects may be related to a gene induction mechanism (31). Today, several other important functions are also associated with CoQ₁₀ (Table 2).

5. CLINICAL INDICATIONS

5.1. Treatment of coenzyme Q₁₀ deficiencies

CoQ₁₀ deficiency is a treatable condition and therefore its diagnosis is essential, especially for pediatricians and infantile neurologists. The diagnosis can be made by direct measurement of CoQ₁₀ in muscle, and reinforced by the presence of reduced biochemical activities of respiratory chain complexes, in particular,

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complexes I+III and II+III. Molecular genetic testing has revealed causative mutations in a small proportion of patients indicating that screening for DNA mutations is not yet effective for diagnosing CoQ₁₀ deficiency (10). An early treatment with high-dose CoQ₁₀ might radically change the natural history of this group of diseases (32). Patients with all forms of CoQ₁₀ deficiency have shown clinical improvement with oral CoQ₁₀ supplementation, but cerebral symptoms are only partially ameliorated, probably because of irreversible structural brain damage before treatment and because of poor penetration of CoQ₁₀ across the blood-brain barrier (33).

CoQ₁₀ deficiency is involved in cardiomyopathies and degenerative muscle and neuronal diseases. The major phenotypes provoked by CoQ₁₀ deficiencies are encephalomyopathy, severe infantile multisystemic disease, cerebellar ataxia, Leigh syndrome with growth retardation, ataxia, nephrotic syndrome and isolated myopathy (34).

The cerebellum may have the narrowest safety margin and, therefore, would be the first tissue to suffer from a pathological shortage of CoQ₁₀ (35). The most severe human CoQ₁₀ deficiencies are due to autosomal recessive mutations and can be classified as primary deficiencies when mutations affect CoQ₁₀ biosynthetic genes (COQ genes) or secondary if the cause is related to other genetic defects (34). In 1989, Ogasahara and colleagues reported the first case of primary CoQ₁₀ deficiency in skeletal muscle (36). Currently, more than 100 patients with CoQ₁₀ deficiency have been reported. Most patients with the infantile-onset multisystemic variant have genetically confirmed primary CoQ₁₀ deficiency (10). Mutations have been described in COQ2, PDSS2, COQ9, PDSS1, and COQ6. Patients with COQ2 mutations have presented with either infantile multisystemic syndrome or isolated nephropathy. A subgroup of patients with juvenile-onset cerebellar ataxia has primary CoQ₁₀ deficiency due to mutations in the ADCK3 gene. Secondary deficiencies include diseases caused by mutations in genes unrelated to ubiquinone biosynthesis, for example aprataxin (APTX) gene, causing ataxia and oculomotor apraxia (37), electron-transferring-flavoprotein dehydrogenase gene (ETFDH), causing isolated myopathy (38), and BRAF gene, causing cardiofaciocutaneous syndrome (39). However, the majority of patients with cerebellar ataxia and CoQ₁₀ deficiency still lack molecular diagnosis. Patients with CoQ₁₀ deficiency showed variable responses to CoQ₁₀ treatment. The recommend oral supplementation doses are up to 2,400 mg daily in adult patients and up to 30 mg/kg daily in pediatric patients, divided into three doses per day (10). Clinical improvement after CoQ₁₀ supplementation was reported in many patients, but treatment protocols have not been standardized, and results have not been uniform in all the patients.

5.2. Mitochondrial disorders

CoQ₁₀ is frequently reduced in muscle of patients with mitochondrial myopathy (11) and CoQ₁₀ is very widely used for primary mitochondrial disorders treatment (40). Numerous case reports and small, open-label studies describe mitochondrial diseases of varying severity that

have responded to CoQ₁₀ supplementation, typically in dosages from 30-300 mg/day (41, 42). A three-month trial included eight patients with mitochondrial encephalomyopathies supplemented with 160 mg CoQ₁₀/day. Although the researchers reported a trend toward improved muscle endurance, less fatigue during daily duties, and decreased serum lactate and pyruvate levels, only the muscle endurance results reached statistical significance. The study authors hypothesized the dosage was too low to provide significant benefit (43). In a six months double-blind clinical trial, 44 patients with mitochondrial myopathies from multiple centers were treated with 2 mg/kg CoQ₁₀ daily. Sixteen of 24 patients experienced at least a 25 percent decrease in post-exercise lactate levels and were selected as “responders” to continue the study. After a further three months at the same dose, no significant differences were observed between the responder and placebo groups. The lack of long-term therapeutic effect in the responders may be attributed to the relatively low dose and short duration of the study (44). Overall, it appears that larger CoQ₁₀ dosages are indicated for mitochondrial disorders. Recently, our group has demonstrated the benefits of CoQ₁₀ supplementation in several cellular models of mitochondrial diseases (45-48). However, the clinical evidence supporting a treatment benefit for CoQ₁₀ in primary mitochondrial disease whilst positive is limited. Reasons for this include the relative rarity and the heterogeneity of mitochondrial diseases included in clinical trials (49).

5.3. Fibromyalgia

Fibromyalgia (FM) is a chronic pain syndrome with unknown etiology and a wide spectrum of symptoms such as allodynia, debilitating fatigue, joint stiffness and migraine. Recent studies have shown some evidences demonstrating that oxidative stress is associated to clinical symptoms in FM of fibromyalgia. Recent findings of our group has shown reduced levels of CoQ₁₀, decreased mitochondrial membrane potential, increased levels of mitochondrial superoxide and increased levels of lipid peroxidation in blood mononuclear cells (BMCs) from FM patients. Mitochondrial dysfunction was also associated with increased expression of autophagic genes and the elimination of dysfunctional mitochondria by mitophagy (50). In other work, FM patients were evaluated clinically with Visual Analogical Scale of pain (VAS), and Fibromyalgia Impact Questionnaire (FIQ). FM patients with CoQ₁₀ deficiency showed a significant reduction on symptoms after CoQ₁₀ treatment (51, 52).

Recently, a randomized, double-blind, placebo-controlled trial was carried out to evaluate the effects of forty days of CoQ₁₀ supplementation (300 mg/day) on clinical and gene expression in 20 FM patients (53). An important clinical improvement was evident after CoQ₁₀ versus placebo treatment showing a reduction in pain, tender points, fatigue, and morning tiredness. Furthermore, CoQ₁₀ supplementation induced a recovery of inflammation, antioxidant enzymes, mitochondrial biogenesis and AMPK (5' adenosine monophosphate-activated protein kinase) gene expression levels. These results lead to the hypothesis that CoQ₁₀ have a potential

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therapeutic effect in FM, and indicate new potential molecular targets for the therapy of this disease. Therefore, determination of CoQ₁₀ deficiency and subsequent supplementation in FM may result in significant clinical improvement.

5.4. Cardiovascular disease

Oxidative stress plays a central role in the pathogenesis of cardiovascular diseases including heart failure and hypertension. Heart failure is often characterized by a loss of contractile function due to an energy depletion status in the mitochondria that has been associated with low endogenous CoQ₁₀ levels. Myocardial deficiency of CoQ₁₀ has been demonstrated in endomyocardial biopsy samples from patients with cardiomyopathy, and deficiency of CoQ₁₀ correlated with the severity of disease, suggesting that therapy with CoQ₁₀ can result in improving quality of life of cardiac patients by enhancing myocardial contractility (14). Numerous studies have investigated the benefit of CoQ₁₀ supplementation for improving cardiovascular function via enhanced energy production, improved contractility of cardiac muscle, and its potent antioxidant activity, particularly prevention of low-density lipoproteins (LDL) oxidation. In 1994, Langsjoen *et al* published a study summarizing eight years of research on the benefits of CoQ₁₀ in clinical cardiology (54). Since this study, numerous other studies have demonstrated the usefulness of CoQ₁₀ supplementation for various cardiovascular conditions. Research has shown CoQ₁₀ levels are depleted in both serum and myocardial tissue samples of patients with chronic heart failure (55, 56). Two important meta-analyses reported significant benefit of CoQ₁₀ on heart failure from various causes (57, 58). Dilated cardiomyopathy is a form of cardiac muscle disease characterized by ventricular dilation, contractile dysfunction, and eventual congestive heart failure. In patients with stable moderate congestive heart failure, oral CoQ₁₀ supplementation was shown to ameliorate cardiac contractility and endothelial dysfunction (59).

5.4.1. Atherosclerosis

CoQ₁₀ in its reduced form, ubiquinol (CoQ₁₀H₂), inhibits protein and DNA oxidation but it is the effect on lipid peroxidation that has been most deeply studied. Ubiquinol inhibits the peroxidation of cell membrane lipids and lipoprotein lipids present in the circulation. Dietary supplementation with CoQ₁₀ results in increased resistance of LDL to the initiation of lipid peroxidation (60). Moreover, CoQ₁₀ has a direct anti-atherogenic effect, which has also been demonstrated in apolipoprotein E-deficient mice fed with a high-fat diet (61). CoQ₁₀ supplement at a dose of 150 mg/day can decrease oxidative stress, increase antioxidant enzyme activity and decrease the inflammatory marker IL-6 in patients with atherosclerosis (62, 63).

5.4.2. Dyslipidemia and statin drugs

Elevated cholesterol and the associated dyslipidemia are commonly treated with 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibiting drugs (“statins”). Because both cholesterol and CoQ₁₀ synthesis depend on HMG-CoA reductase, both can

be blocked. Different mechanisms have been proposed to explain statin-induced myopathy, including reduction of mevalonate pathway products, induction of apoptosis, mitochondrial dysfunction, and genetic predisposition (64). Depletion in CoQ₁₀ may account for the statin-induced myopathies observed in some patients, the most serious of which is rhabdomyolysis. It is therefore important that clinicians understand the mechanism of action of these drugs as well as their overall effect at a biochemical level. From 1990-2004, 13 controlled trials have demonstrated significant CoQ₁₀ depletion secondary to statin therapy (65). Consequently, supplementing with CoQ₁₀ is highly recommended to prevent the myopathic side effects associated with the statin drugs. Recently, it has also been reported statin side effects on energy and exertional fatigue (66). However, clinical evidence supporting CoQ₁₀'s use in the treatment of statin-induced myopathy is limited and controversial (67).

In a study by Oh *et al.* (68), 133 patients from a 291-subject sample were found to be intolerant to statin monotherapy. The investigators showed that genetic variations in the COQ2 gene, which is involved in CoQ₁₀ biosynthesis, were significantly associated with an increased prevalence of statin intolerance. These preliminary pharmacogenetic results support the hypothesis that statin intolerance is associated with genetic variation in the COQ2 gene. It is therefore possible that an unidentified relationship exists between statin intolerance and CoQ₁₀ deficiency. One way to address this problem is through a genomic analysis of susceptibility genes, which would reveal the likelihood of a pharmacogenetic link to statin intolerance. This approach may help in the prevention of muscle-related symptoms through the supplementation of both statins and CoQ₁₀.

5.4.3. Hypertension

Depending on the class, various antihypertensive drugs can have adverse effects such as depression, cough, and cardiac and renal dysfunction (69, 70). Furthermore, many patients need to take more than one drug to control their blood pressure, increasing their risk of side effects. Some researchers believe CoQ₁₀ supplementation may reduce the need to take multiple antihypertensive drugs (71).

CoQ₁₀ appears to lower blood pressure. The exact mechanism is not known, but one theory is that it reduces peripheral resistance by preserving nitric oxide (70). Nitric oxide relaxes peripheral arteries, lowering blood pressure. In some forms of hypertension, superoxide radicals that inactivate nitric oxide are overproduced; CoQ₁₀, with its antioxidant effects, may prevent the inactivation of nitric oxide by these free radicals. Alternatively, CoQ₁₀ may boost the production of the prostaglandin prostacyclin (PGI₂) a potent vasodilator and inhibitor of platelet aggregation, or it may enhance the sensitivity of arterial smooth muscle to PGI₂, or both (72). A meta-analysis of clinical trials investigating the use of CoQ₁₀ for hypertension assessed overall efficacy. Blood pressure reduction was noted in all 12 trials, regardless of whether CoQ₁₀ was given alone or as an adjunct to standard antihypertensive medication, without significant side

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effects (73). In a recent randomized, double-blind, placebo-controlled 12-week crossover trial, the authors conclude that it is possible that CoQ₁₀ may improve blood pressure control under some circumstances, but any effects are likely to be smaller than reported in previous meta-analyses (74). In some cases, it seems reasonable to recommend this product as an adjunct to conventional antihypertensive therapy. However, larger, well-designed clinical trials of CoQ₁₀'s antihypertensive effects on specific clinical outcomes such as the risk of stroke or myocardial infarction are needed to define its true therapeutic value (67).

5.5. Neurological conditions

5.5.1. Parkinson's disease

A number of preclinical studies in both *in vitro* and *in vivo* models of Parkinson's disease (PD) have demonstrated that CoQ₁₀ can protect the nigrostriatal dopaminergic system and some clinical trials have looked at the neuroprotective effects of CoQ₁₀ in patients with early and mid-stage PD (75). Research suggests CoQ₁₀ may play a role in the cellular dysfunction found in PD, providing a protective agent for Parkinsonian patients (76). Significantly reduced levels of CoQ₁₀ have been observed in blood and platelet mitochondria (77), and plasma (78) of PD patients. Therefore, deficiency of CoQ₁₀ should be explored as a potential peripheral biomarker of antioxidant status in PD (79).

Since 1998, at least four clinical trials on the efficacy of CoQ₁₀ in PD have been conducted (80-83). Results seem to indicate a positive effect, warranting larger double-blind, placebo-controlled trials. Recently, it has been demonstrated that cellular pathophysiological alterations associated with mitochondrial dysfunction in induced pluripotent stem cell-derived neural cells from familial PD patients and at-risk individuals could be rescued with CoQ₁₀ (84).

5.5.2. Huntington's disease

Huntington's disease (HD) is a neurodegenerative genetic disorder caused by an expansion of CAG repeats in the HD gene encoding for huntingtin (Htt), resulting in progressive death of striatal neurons, with clinical symptoms of chorea, dementia and dramatic weight loss. Metabolic and mitochondrial dysfunction caused by the expanded polyglutamine sequence have been described along with other mechanisms of neurodegeneration previously described in human tissues and animal models of HD (85). Strong evidence exists for early oxidative stress in HD, coupled with mitochondrial dysfunction, each exacerbating the other and leading to an energy deficit (86). If oxidative damage plays a role in HD, then therapeutic strategies that reduce reactive oxygen species may ameliorate the neurodegenerative process. One such strategy using CoQ₁₀ has been proposed. High-dose CoQ₁₀ is safe and tolerable in HD patients. In addition, there are parallels in reducing markers of oxidative stress in both HD mice and HD patients after CoQ₁₀ treatment (86).

5.5.3. Alzheimer's disease

Increasing evidence suggests that Alzheimer's disease (AD) is associated with oxidative damage that is

caused in part by mitochondrial dysfunction (87). Studies have shown CoQ₁₀ to be neuroprotective in AD through protection of oxidative damage and attenuation of mitochondrial dysfunction (88).

However, in a recent double-blind, placebo-controlled clinical trial (Trial Registration clinicaltrials.gov Identifier: NCT00117403) antioxidant treatment, including CoQ₁₀, did not influence cerebrospinal fluid biomarkers related to amyloid or tau pathology (89).

5.5.4. Friedreich's ataxia

There is extensive evidence that mitochondrial respiratory chain dysfunction, oxidative damage and iron accumulation play significant roles in the disease mechanism. Therapeutic avenues for patients with Friedreich's ataxia (FRDA) are beginning to be explored in particular targeting antioxidant protection, enhancement of mitochondrial oxidative phosphorylation, iron chelation and more recently increasing frataxin transcription. The use of quinone therapy has been the most extensively studied to date with clear benefits demonstrated using evaluations of both disease biomarkers and clinical symptoms (90).

An open-label, pilot trial explored the use of 400 mg CoQ₁₀ plus 2,100 IU vitamin E daily in 10 patients with FRDA for 47 months. A sustained improvement in mitochondrial energy synthesis was observed that was associated with a slowing of disease progression and improved cardiac function (91). However, results are less satisfactory in shorter studies. Idebenone, a structural analog of CoQ₁₀ with a benzoquinone nucleus and a hydroxydecyl side chain did not significantly alter neurological function in FRDA during the 6-month study. Larger studies of longer duration may be needed to assess the therapeutic potential of drug candidates on neurological function in FRDA (92). In a recent review, Parkinson *et al.* (93) conclude that although much time and expense has been expended on clinical trials of antioxidant therapies in FRDA, definitive answers as to efficacy remain elusive. Prescribing patterns consequently remain inconsistent and many patients currently incur significant costs in procuring antioxidant supplements privately, without robust clinical evidence.

5.6. Cancer

Decreased levels of CoQ₁₀ have been found in plasma of women with breast cancer and in cancerous breast tissue, and low levels correlated with a worse prognosis (94). Case reports demonstrated 390 mg CoQ₁₀ daily resulted in tumor regression and disappearance of previously diagnosed metastasis. One to three years later, depending on the case, metastases had not reappeared (95, 96). In 117 melanoma patients without metastasis, plasma CoQ₁₀ levels were significantly lower than in control subjects and were associated with primary tumor thickness, with the highest CoQ₁₀ levels associated with thinner tumors. In addition, patients who developed metastases had lower CoQ₁₀ levels than those who did not, and subjects with lower baseline CoQ₁₀ levels had shorter disease-free intervals (96). Low plasma levels of CoQ₁₀ have been demonstrated in cervical intraepithelial neoplasia and cervical cancer (97).

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Mechanisms for CoQ₁₀'s benefit for cancer may include immune system enhancement and antioxidant activity. CoQ₁₀ can be depleted by the use of the chemotherapeutic drug doxorubicin (Adriamycin®), resulting in cardiotoxicity if a high enough cumulative dose is achieved. Supplemental CoQ₁₀ (100-200 mg/day) can prevent cardiac damage, as well as diarrhea and stomatitis that are caused by this agent, without decreasing its chemotherapeutic effectiveness (98). A systematic review of controlled trials in cancer patients revealed that CoQ₁₀ provides protection against cardiotoxicity and liver toxicity in patients receiving anthracycline chemotherapy drugs, such as doxorubicin (99). Moreover, it has been reported that chemotherapeutic drugs such as camptothecin, etoposide, doxorubicin and methotrexate induced an increase in CoQ₁₀ levels in cancer cell lines by upregulation of COQ7, COQ4 and COQ8 gene expression, as part of an antioxidant response against free radical production (100). On the other hand, compositions containing reduced CoQ₁₀ (in foods and beverages) have been proposed for preventing cancer and for mitigating the adverse reactions of anticancer agents (101).

5.7. Diabetes

Diabetes is a chronic metabolic disorder that continues to present as a major health problem worldwide. It is characterized by absolute or relative deficiencies in insulin secretion and/or insulin action and is associated with chronic hyperglycemia and disturbances of carbohydrate, lipid, and protein metabolism. Many studies suggest a central role for oxidative stress in the pathogenesis of this multifaceted metabolic disorder. This has prompted investigations in the use of antioxidants as a complementary therapeutic approach (102). Serum CoQ₁₀ levels in type 2 diabetic patients are often decreased and may be associated with subclinical diabetic cardiomyopathy, reversible by CoQ₁₀ supplementation (103). In three separate randomized, double-blind clinical trials, a total of 194 dyslipidemic type 2 diabetic patients received 200 mg CoQ₁₀ or placebo daily for 12 weeks. One study also compared CoQ₁₀ stand-alone treatment to a CoQ₁₀-fenofibrate combination and to fenofibrate (a lipid lowering medication) alone. Primary outcomes were endothelial function of the brachial artery (104), blood pressure (105), glycemic control, (105) and forearm microcirculatory function (106). CoQ₁₀ supplementation in this population raised plasma CoQ₁₀ levels, improved endothelial function in the brachial artery, significantly decreased both systolic and diastolic blood pressure, decreased glycosylated hemoglobin (HbA1C), and in combination with fenofibrate markedly improved both endothelial and non-endothelial forearm vasodilation.

Furthermore, it has been demonstrated that twelve weeks treatment with ubiquinone improves clinical outcomes and nerve conduction parameters of diabetic polyneuropathy; furthermore, it reduces oxidative stress without significant adverse events (107). These data identify CoQ₁₀ as a potential candidate for future treatment of peripheral neuropathy in type 2 diabetes.

5.8. Male infertility

Both the bioenergetic and the antioxidant role of CoQ₁₀ suggest a possible involvement in sperm

biochemistry and male infertility (108). CoQ₁₀ can be quantified in seminal fluid, where its concentration correlates with sperm count and motility (109). It was found that distribution of CoQ₁₀ between sperm cells and seminal plasma was altered in varicocele patients, who also presented a higher level of oxidative stress and lower total antioxidant capacity. The redox status of CoQ₁₀ in seminal fluid was also determined: an inverse correlation was found between ubiquinol/ubiquinone ratio and hydroperoxide levels and between this ratio and the percentage of abnormal sperm forms. Subsequently, CoQ₁₀ was administered to a group of idiopathic asthenozoospermic infertile patients. Treatment led to a significant increase in the concentration of CoQ₁₀, both in seminal plasma and sperm cells, and improvement in sperm motility (110). In a recent study, it has been demonstrated that CoQ₁₀ improves semen quality and pregnancy rate (111).

5.9. Periodontal disease

Periodontal disease is an inflammatory disease process resulting from the interaction of a bacterial attack and host inflammatory response. Arrays of molecules are considered to mediate the inflammatory response at one time or another, among these are free radicals and reactive oxygen species (ROS). Periodontal pathogens can induce ROS overproduction and thus may cause collagen and periodontal cell breakdown. When ROS are scavenged by antioxidants, there can be a reduction of collagen degradation. Ubiquinol serves as an endogenous antioxidant which increases the concentration of CoQ₁₀ in the diseased gingiva and effectively suppresses advanced periodontal inflammation (112).

5.10. Migraine

There are strong similarities between migraine and encephalomyopathies due to mitochondrial disorders, in which patients suffer genetic abnormalities in mitochondrial energy production to produce lactic acidosis, stroke, and migraine headaches. The theory of migraine as a mitochondrial disorder seems to have abundant evidence (113). Arising from these extensive neurophysiological studies, treatment of metabolic encephalomyopathies with pharmacological doses of riboflavin and CoQ₁₀ has shown positive benefits. The same treatment has now been applied to migraine, adding clinical support to the theory that migraine is a mitochondrial disorder. Currently, riboflavin and CoQ₁₀ supplementation has been recommended widely as safe and effective prophylactic therapy for migraine. Evidence indicates impaired energy metabolism may be present in brains of migraine sufferers. Rozen *et al* supplemented migraine patients with 150 mg CoQ₁₀ daily for three months and demonstrated a 50-percent reduction in number of days with migraine headache, regardless of whether patients experienced aura or not (114). Deficiency of CoQ₁₀ may be common in pediatric and adolescent migraine. Determination of deficiency and consequent supplementation may result in clinical improvement (115).

5.11. Pregnancy

Plasma CoQ₁₀ levels rise with each trimester of pregnancy and fetal wasting with subsequent spontaneous abortion has been correlated with low levels of CoQ₁₀

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(116). Supplementation with CoQ₁₀ reduces the risk of developing pre-eclampsia (gestational hypertension in association with significant amounts of protein in the urine) in women at risk for the condition (117).

5.12. Down's syndrome

Down syndrome (DS) is a chromosomal abnormality (trisomy 21) associated with a complex phenotype. Oxidative stress is known to play a major role in this pathology both due to genetic and epigenetic factors, suggesting that oxidative imbalance contributes to the clinical manifestation of DS (118). Structural changes and abnormal function of mitochondria have been documented in DS cells, patients, and animal models. DS cells in culture exhibit a wide array of functional mitochondrial abnormalities. Two studies have investigated the effect of CoQ₁₀ treatment on DNA damage in DS patients. Results suggest that the effect of CoQ₁₀ treatment in DS not only reflects antioxidant efficacy, but likely modulates DNA repair mechanisms (119).

5.13. Aging

Decrease of CoQ₁₀ levels during aging could be one of the main factors in the development of chronic diseases in old people. Furthermore, since CoQ₁₀ is not only an antioxidant but also is involved in a plethora of cellular processes appropriate uptake of CoQ₁₀ into cellular is crucial for improvement of cell activity during aging. Maintenance of CoQ₁₀ functional levels at cell membranes either by dietary supplementation or by improving endogenous synthesis can be a key strategy to enhance health during aging (120).

6. COSMETICS

Topical antioxidants have been used to treat photoaged and chronologically aged skin (121). Free radicals are known to promote oxidation of nucleic acids, proteins, and lipids and can damage intracellular structures including DNA (122). Free radicals also up-regulate transcription factors such as activator protein 1 (AP-1) and nuclear transcription factor-kappa B (NF-κB) (123). AP-1 is responsible for production of metalloproteinases that break down existing collagen, contributing to skin wrinkling (124). NF-κ B up-regulates transcription of proinflammatory mediators such as interleukin-1 (IL-1), IL-6, IL-8, and tumor necrosis factor-α (125). Acting through cell-surface receptors, these proinflammatory mediators further activate AP-1 and NF-κB, resulting in more damage. It is the sum of these events that are responsible for skin aging (126).

In the skin CoQ₁₀ acts as an antioxidant with 10-fold higher levels in the epidermis than in the dermis (127). The reduction in the efficiency of antioxidant systems has been proposed as a factor of skin ageing. Therefore, in the cosmetic industry the antioxidant CoQ₁₀ is widely used in anti-ageing products (128). Lipid nanoparticles, solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC), are innovative carrier systems for CoQ₁₀ that are derived from oil/water emulsions (129).

7. DRUG-NUTRIENT INTERACTIONS

Cholesterol-lowering drugs such as lovastatin and pravastatin inhibit the enzyme HMG-CoA reductase, required for synthesis of cholesterol as well as CoQ₁₀, resulting in decreased serum CoQ₁₀ (130). Beta blockers, propranolol and metoprolol, (131) phenothiazines and tricyclic antidepressants have been shown to inhibit CoQ₁₀-dependent enzymes (132). CoQ₁₀'s effects on platelet function may increase the risk of bleeding in patients taking antiplatelet drugs such as aspirin (133). On the other hand, since it acts like vitamin K, it may counteract the anticoagulant effects of warfarin (134). CoQ₁₀ may have an additive antihypertensive effect when given with antihypertensive drugs (135). CoQ₁₀ may improve beta-cell function and enhance insulin sensitivity, which may reduce insulin requirements for diabetic patients (105).

8. TOXICITY

CoQ₁₀ treatment is safe, even at the highest doses cited in the literature. Most clinical trials have not reported significant adverse effects that necessitated stopping therapy (136). However, gastrointestinal effects such as abdominal discomfort, nausea, vomiting, diarrhea, and anorexia have occurred (136). Allergic rash and headache have also been reported (136). In addition, CoQ₁₀'s antiplatelet effect may increase the risk of bleeding (137). It undergoes biotransformation in the liver and is eliminated primarily via the biliary tract (137), therefore it can accumulate in patients with hepatic impairment or biliary obstruction.

9. COENZYME Q₁₀-RELATED COMPOUNDS

Intestinal absorption of dietary CoQ₁₀ is very limited and only chronic ingestion of relatively large doses of CoQ₁₀ increase CoQ₁₀ concentrations especially in heart and brain mitochondria in rodent models (4). For this reason, the development of less hydrophobic structural derivatives of CoQ₁₀, and therefore with better pharmacokinetics profiles, are emerging as promising drugs for treating diseases with mitochondrial dysfunction. Idebenone and MitoQ have been evaluated in clinical trials for safety, toxicity and their effect for treating different diseases (101).

Several advancements have been made to enhance the bioavailability of CoQ₁₀ using various new formulations and approaches like size reduction, solubility enhancement (by solid dispersion, prodrug, complexation, ionization) and use of novel drug carriers such as liposomes, microspheres, nanoparticles, nanoemulsions and self-emulsifying system (138).

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11. REFERENCES

1. F. L. Crane: Biochemical functions of coenzyme Q10. *J Am Coll Nutr* 20(6), 591-8 (2001)
2. D. A. Groneberg, B. Kindermann, M. Althammer, M. Klapper, J. Vormann, G. P. Littarru and F. Doring: Coenzyme Q10 affects expression of genes involved in cell signalling, metabolism and transport in human CaCo-2 cells. *Int J Biochem Cell Biol* 37(6), 1208-18 (2005)
3. C. Schmelzer, I. Lindner, G. Rimbach, P. Niklowitz, T. Menke and F. Doring: Functions of coenzyme Q10 in inflammation and gene expression. *Biofactors* 32(1-4), 179-83 (2008)
4. H. N. Bhagavan and R. K. Chopra: Coenzyme Q10: absorption, tissue uptake, metabolism and pharmacokinetics. *Free Radic Res* 40(5), 445-53 (2006)
5. L. Ernster and G. Dallner: Biochemical, physiological and medical aspects of ubiquinone function. *Biochim Biophys Acta* 1271(1), 195-204 (1995)
6. U. C. Tran and C. F. Clarke: Endogenous synthesis of coenzyme Q in eukaryotes. *Mitochondrion* 7 Suppl, S62-71 (2007)
7. K. Folkers: Relevance of the biosynthesis of coenzyme Q10 and of the four bases of DNA as a rationale for the molecular causes of cancer and a therapy. *Biochem Biophys Res Commun* 224(2), 358-61 (1996)
8. M. Turunen, J. Olsson and G. Dallner: Metabolism and function of coenzyme Q. *Biochim Biophys Acta* 1660(1-2), 171-99 (2004)
9. M. Santos-Gonzalez, C. Gomez Diaz, P. Navas and J. M. Villalba: Modifications of plasma proteome in long-lived rats fed on a coenzyme Q10-supplemented diet. *Exp Gerontol* 42(8), 798-806 (2007)
10. V. Emmanuele, L. C. Lopez, A. Berardo, A. Naini, S. Tadesse, B. Wen, E. D'Agostino, M. Solomon, S. DiMauro, C. Quinzii and M. Hirano: Heterogeneity of Coenzyme Q10 Deficiency: Patient Study and Literature Review. *Arch Neurol* 69(8), 978-83 (2012)
11. S. Sacconi, E. Trevisson, L. Salviati, S. Ayme, O. Rigal, A. G. Redondo, M. Mancuso, G. Siciliano, P. Tonin, C. Angelini, K. Aure, A. Lombes and C. Desnuelle: Coenzyme Q10 is frequently reduced in muscle of patients with mitochondrial myopathy. *Neuromuscul Disord* 20(1), 44-8 (2010)
12. R. S. Sohal and M. J. Forster: Coenzyme Q, oxidative stress and aging. *Mitochondrion* 7 Suppl, S103-11 (2007)
13. C. M. Quinzii, S. DiMauro and M. Hirano: Human coenzyme Q10 deficiency. *Neurochem Res* 32(4-5), 723-7 (2007)
14. K. Folkers, J. Wolaniuk, R. Simonsen, M. Morishita and S. Vadhanavikit: Biochemical rationale and the cardiac response of patients with muscle disease to therapy with coenzyme Q10. *Proc Natl Acad Sci U S A* 82(13), 4513-6 (1985)
15. P. Niklowitz, T. Menke, T. Wiesel, E. Mayatepek, J. Zschocke, J. G. Okun and W. Andler: Coenzyme Q10 in plasma and erythrocytes: comparison of antioxidant levels in healthy probands after oral supplementation and in patients suffering from sickle cell anemia. *Clin Chim Acta* 326(1-2), 155-61 (2002)
16. A. J. Duncan, S. J. Heales, K. Mills, S. Eaton, J. M. Land and I. P. Hargreaves: Determination of coenzyme Q10 status in blood mononuclear cells, skeletal muscle, and plasma by HPLC with di-propoxy-coenzyme Q10 as an internal standard. *Clin Chem* 51(12), 2380-2 (2005)
17. S. L. Molyneux, C. M. Florkowski, P. M. George, A. P. Pilbrow, C. M. Frampton, M. Lever and A. M. Richards: Coenzyme Q10: an independent predictor of mortality in chronic heart failure. *J Am Coll Cardiol*, 52(18), 1435-41 (2008)
18. C. Weber, A. Bysted and G. Holmer: Coenzyme Q10 in the diet--daily intake and relative bioavailability. *Mol Aspects Med* 18 Suppl, S251-4 (1997)
19. M. F. Beal: Coenzyme Q10 administration and its potential for treatment of neurodegenerative diseases. *Biofactors* 9(2-4), 261-6 (1999)
20. A. Ochiai, S. Itagaki, T. Kurokawa, M. Kobayashi, T. Hirano and K. Iseki: Improvement in intestinal coenzyme q10 absorption by food intake. *Yakugaku zasshi* 127(8), 1251-4 (2007)
21. Y. Zhang, F. Aberg, E. L. Appelkvist, G. Dallner and L. Ernster: Uptake of dietary coenzyme Q supplement is limited in rats. *J Nutr*, 125(3), 446-53 (1995)
22. L. K. Kwong, S. Kamzalov, I. Rebrin, A. C. Bayne, C. K. Jana, P. Morris, M. J. Forster and R. S. Sohal: Effects of coenzyme Q(10) administration on its tissue concentrations, mitochondrial oxidant generation, and oxidative stress in the rat. *Free Radic Biol Med* 33(5), 627-38 (2002)
23. A. Palamakula, M. Soliman and M. M. Khan: Regional permeability of coenzyme Q10 in isolated rat gastrointestinal tracts. *Pharmazie* 60(3), 212-4 (2005)
24. Weber C, Bysted A and H. G.: Intestinal absorption of coenzyme Q10 administered in a meal or as capsules to healthy subjects. *Nutr Res Rev* 17, 941-945 (1997)
25. M. Weis, S. A. Mortensen, M. R. Rassing, J. Moller-Sonnergaard, G. Poulsen and S. N. Rasmussen:

Clinical applications of coenzyme Q10

- Bioavailability of four oral coenzyme Q10 formulations in healthy volunteers. *Mol Aspects Med* 15 Suppl, s273-80 (1994)
26. W. Lyon, O. Van den Brink, S. Pepe, M. Wowk, S. Marasco and F. L. Rosenfeldt. Similar therapeutic serum levels attained with emulsified and oil-based preparations of coenzyme Q10. *Asia Pac J Clin Nutr* 10(3), 212-5 (2001)
27. M. V. Miles: The uptake and distribution of coenzyme Q10. *Mitochondrion* 7 Suppl, S72-7 (2007)
28. J. N. Hathcock and A. Shao: Risk assessment for coenzyme Q10 (Ubiquinone). *Regul Toxicol Pharmacol* 45(3), 282-8 (2006)
29. M. V. Miles, B. J. Patterson, M. B. Schapiro, F. J. Hickey, M. Chalfonte-Evans, P. S. Horn and S. L. Hotze: Coenzyme Q10 absorption and tolerance in children with Down syndrome: a dose-ranging trial. *Pediatr Neurol* 35(1), 30-7 (2006)
30. K. Hosoe, M. Kitano, H. Kishida, H. Kubo, K. Fujii and M. Kitahara: Study on safety and bioavailability of ubiquinol (Kaneka QH) after single and 4-week multiple oral administration to healthy volunteers. *Regul Toxicol Pharmacol* 47(1), 19-28 (2007)
31. G. P. Littarru and L. Tiano: Clinical aspects of coenzyme Q10: an update. *Nutrition* 26(3), 250-4 (2010)
32. S. DiMauro, C. M. Quinzii and M. Hirano: Mutations in coenzyme Q10 biosynthetic genes. *J Clin Invest* 117(3), 587-9 (2007)
33. A. Rotig, J. Mollet, M. Rio and A. Munnich: Infantile and pediatric quinone deficiency diseases. *Mitochondrion* 7 Suppl, S112-21 (2007)
34. C. M. Quinzii and M. Hirano: Primary and secondary CoQ(10) deficiencies in humans. *Biofactors* 37(5), 361-5 (2011)
35. A. Naini, V. J. Lewis, M. Hirano and S. DiMauro: Primary coenzyme Q10 deficiency and the brain. *Biofactors* 18(1-4), 145-52 (2003)
36. S. Ogasahara, A. G. Engel, D. Frens and D. Mack: Muscle coenzyme Q deficiency in familial mitochondrial encephalomyopathy. *Proc Natl Acad Sci U S A* 86(7), 2379-82 (1989)
37. C. M. Quinzii, A. G. Kattah, A. Naini, H. O. Akman, V. K. Mootha, S. DiMauro and M. Hirano: Coenzyme Q deficiency and cerebellar ataxia associated with an aprataxin mutation. *Neurology* 64(3), 539-41 (2005)
38. K. Gempel, H. Topaloglu, B. Talim, P. Schneiderat, B. G. Schoser, V. H. Hans, B. Palmafy, G. Kale, A. Tokatli, C. Quinzii, M. Hirano, A. Naini, S. DiMauro, H. Prokisch, H. Lochmuller and R. Horvath: The myopathic form of coenzyme Q10 deficiency is caused by mutations in the electron-transferring-flavoprotein dehydrogenase (ETFDH) gene. *Brain* 130(Pt 8), 2037-44 (2007)
39. A. Aeby, Y. Sznajder, H. Cave, E. Rebuffat, R. Van Coster, O. Rigal and P. Van Bogaert. Cardiofaciocutaneous (CFC) syndrome associated with muscular coenzyme Q(10) deficiency. *J Inherit Metab Dis* 30(5), 827 (2007)
40. D. S. Kerr: Treatment of mitochondrial electron transport chain disorders: a review of clinical trials over the past decade. *Mol Genet Metab* 99(3), 246-55 (2010)
41. A. Berbel-Garcia, J. R. Barbera-Farre, J. P. Etessam, A. M. Salio, A. Cabello, E. Gutierrez-Rivas and Y. Campos: Coenzyme Q 10 improves lactic acidosis, stroke-like episodes, and epilepsy in a patient with MELAS (mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes). *Clin Neuropharmacol* 27(4), 187-91 (2004)
42. R. Gold, P. Seibel, G. Reinelt, R. Schindler, P. Landwehr, A. Beck and H. Reichmann: Phosphorus magnetic resonance spectroscopy in the evaluation of mitochondrial myopathies: results of a 6-month therapy study with coenzyme Q. *Eur Neurol* 36(4), 191-6 (1996)
43. R. S. Chen, C. C. Huang and N. S. Chu: Coenzyme Q10 treatment in mitochondrial encephalomyopathies. Short-term double-blind, crossover study. *Eur Neurol* 37(4), 212-8 (1997)
44. N. Bresolin, C. Doriguzzi, C. Ponzetto, C. Angelini, I. Moroni, E. Castelli, E. Cossutta, A. Binda, A. Gallanti, S. Gabellini and *et al.*: Ubidecarenone in the treatment of mitochondrial myopathies: a multi-center double-blind trial. *J Neurol Sci* 100(1-2), 70-8 (1990)
45. A. Rodriguez-Hernandez, M. D. Cordero, L. Salvati, R. Artuch, M. Pineda, P. Briones, L. Gomez Izquierdo, D. Cotan, P. Navas and J. A. Sanchez-Alcazar: Coenzyme Q deficiency triggers mitochondria degradation by mitophagy. *Autophagy* 5(1), 19-32 (2009)
46. J. Garrido-Maraver, M. D. Cordero, I. Dominguez Monino, S. Pereira-Arenas, A. V. Lechuga-Vieco, D. Cotan, M. De la Mata, M. Oropesa-Avila, M. De Miguel, J. Bautista Lorite, E. Rivas Infante, M. Alvarez-Dolado, P. Navas, S. Jackson, S. Francisci and J. A. Sanchez-Alcazar: Screening of effective pharmacological treatments for MELAS syndrome using yeasts, fibroblasts and cybrids models of the disease. *Br J Pharmacol* 167(6), 1311-28 (2012)
47. M. De la Mata, J. Garrido-Maraver, D. Cotan, M. D. Cordero, M. Oropesa-Avila, L. G. Izquierdo, M. De Miguel, J. B. Lorite, E. R. Infante, P. Ybot, S. Jackson and J. A. Sanchez-Alcazar: Recovery of MERRF fibroblasts and cybrids pathophysiology by Coenzyme Q10. *Neurotherapeutics* 9(2), 446-63 (2012)
48. D. Cotan, M. D. Cordero, J. Garrido-Maraver, M. Oropesa-Avila, A. Rodriguez-Hernandez, L. Gomez Izquierdo, M. De la Mata, M. De Miguel, J. B. Lorite, E. R.

Clinical applications of coenzyme Q10

- Infante, S. Jackson, P. Navas and J. A. Sanchez-Alcazar: Secondary coenzyme Q10 deficiency triggers mitochondria degradation by mitophagy in MELAS fibroblasts. *FASEB J* 25(8), 2669-87 (2011)
49. R. H. Haas: The evidence basis for coenzyme Q therapy in oxidative phosphorylation disease. *Mitochondrion* 7 Suppl, S136-45 (2007)
50. M. D. Cordero, M. De Miguel, A. M. Moreno Fernandez, I. M. Carmona Lopez, J. Garrido Maraver, D. Cotan, L. Gomez Izquierdo, P. Bonal, F. Campa, P. Bullon, P. Navas and J. A. Sanchez Alcazar: Mitochondrial dysfunction and mitophagy activation in blood mononuclear cells of fibromyalgia patients: implications in the pathogenesis of the disease. *Arthritis Res Ther* 12(1), R17 (2010)
51. M. D. Cordero, F. J. Cano-Garcia, E. Alcocer-Gomez, M. De Miguel and J. A. Sanchez-Alcazar: Oxidative stress correlates with headache symptoms in fibromyalgia: coenzyme Q10 effect on clinical improvement. *PLoS One* 7(4), e35677 (2012)
52. M. D. Cordero, E. Alcocer-Gomez, M. de Miguel, F. J. Cano-Garcia, C. M. Luque, P. Fernandez-Riejo, A. M. Fernandez and J. A. Sanchez-Alcazar: Coenzyme Q(10): a novel therapeutic approach for Fibromyalgia? case series with 5 patients. *Mitochondrion* 11(4), 623-5 (2011)
53. M. D. Cordero, E. Alcocer-Gomez, M. de Miguel, O. Culic, A. M. Carrion, J. M. Alvarez-Suarez, P. Bullon, M. Battino, A. Fernandez-Rodriguez and J. A. Sanchez-Alcazar: Can Coenzyme Q Improve Clinical and Molecular Parameters in Fibromyalgia? *Antioxid. Redox Signaling* (2013) in press
54. H. Langsjoen, P. Langsjoen, R. Willis and K. Folkers: Usefulness of coenzyme Q10 in clinical cardiology: a long-term study. *Mol Aspects Med* 15 Suppl, s165-75 (1994)
55. K. Folkers, G. P. Littarru, L. Ho, T. M. Runge, S. Havanonda and D. Cooley: Evidence for a deficiency of coenzyme Q10 in human heart disease. *Int Z Vitaminforsch* 40(3), 380-90 (1970)
56. K. Folkers, S. Vadhanavikit and S. A. Mortensen: Biochemical rationale and myocardial tissue data on the effective therapy of cardiomyopathy with coenzyme Q10. *Proc Natl Acad Sci USA* 82(3), 901-4 (1985)
57. S. A. Mortensen: Overview on coenzyme Q10 as adjunctive therapy in chronic heart failure. Rationale, design and end-points of "Q-symbio"--a multinational trial. *Biofactors* 18(1-4), 79-89 (2003)
58. S. Sander, C. I. Coleman, A. A. Patel, J. Kluger and C. M. White: The impact of coenzyme Q10 on systolic function in patients with chronic heart failure. *J Card Fail* 12(6), 464-72 (2006)
59. G. P. Littarru and L. Tiano: Bioenergetic and antioxidant properties of coenzyme Q10: recent developments. *Mol Biotechnol* 37(1), 31-7 (2007)
60. D. Mohr, V. W. Bowry and R. Stocker: Dietary supplementation with coenzyme Q10 results in increased levels of ubiquinol-10 within circulating lipoproteins and increased resistance of human low-density lipoprotein to the initiation of lipid peroxidation. *Biochim Biophys Acta* 1126(3), 247-54 (1992)
61. P. K. Witting, K. Pettersson, J. Letters and R. Stocker: Anti-atherogenic effect of coenzyme Q10 in apolipoprotein E gene knockout mice. *Free Radic Biol Med* 29(3-4), 295-305 (2000)
62. B. J. Lee, Y. C. Huang, S. J. Chen and P. T. Lin: Coenzyme Q10 supplementation reduces oxidative stress and increases antioxidant enzyme activity in patients with coronary artery disease. *Nutrition* 28(3), 250-5 (2012)
63. B. J. Lee, Y. C. Huang, S. J. Chen and P. T. Lin: Effects of coenzyme Q10 supplementation on inflammatory markers (high-sensitivity C-reactive protein, interleukin-6, and homocysteine) in patients with coronary artery disease. *Nutrition* 28(7-8), 767-72 (2012)
64. E. Mas and T. A. Mori: Coenzyme Q(10) and statin myalgia: what is the evidence? *Curr Atheroscler Rep* 12(6), 407-13 (2010)
65. I. P. Hargreaves, A. J. Duncan, S. J. Heales and J. M. Land: The effect of HMG-CoA reductase inhibitors on coenzyme Q10: possible biochemical/clinical implications. *Drug Saf* 28(8), 659-76 (2005)
66. B. A. Golomb, M. A. Evans, J. E. Dimsdale and H. L. White: Effects of statins on energy and fatigue with exertion: results from a randomized controlled trial. *Arch Intern Med* 172(15), 1180-2 (2012)
67. M. Wyman, M. Leonard and T. Morledge: Coenzyme Q10: a therapy for hypertension and statin-induced myalgia? *Cleve Clin J Med* 77(7), 435-42 (2010)
68. J. Oh, M. R. Ban, B. A. Miskie, R. L. Pollex and R. A. Hegele: Genetic determinants of statin intolerance. *Lipids Health Dis* 6, 7 (2007)
69. A. Hadj, S. Pepe and F. Rosenfeldt: The clinical application of metabolic therapy for cardiovascular disease. *Heart Lung Circ* 16 Suppl 3, S56-64 (2007)
70. S. Pepe, S. F. Marasco, S. J. Haas, F. L. Sheeran, H. Krum and F. L. Rosenfeldt: Coenzyme Q10 in cardiovascular disease. *Mitochondrion* 7 Suppl, S154-67 (2007)
71. P. Langsjoen, R. Willis and K. Folkers: Treatment of essential hypertension with coenzyme Q10. *Mol Aspects Med* 15 Suppl, S265-72 (1994)
72. K. Lonrot, I. Porsti, H. Alho, X. Wu, A. Hervonen and J. P. Tolvanen: Control of arterial tone after long-term coenzyme Q10 supplementation in senescent rats. *Br J Pharmacol* 124(7), 1500-6 (1998)

Clinical applications of coenzyme Q10

73. F. L. Rosenfeldt, S. J. Haas, H. Krum, A. Hadj, K. Ng, J. Y. Leong and G. F. Watts: Coenzyme Q10 in the treatment of hypertension: a meta-analysis of the clinical trials. *J Hum Hypertens* 21(4), 297-306 (2007)
74. J. M. Young, C. M. Florkowski, S. L. Molyneux, R. G. McEwan, C. M. Frampton, M. G. Nicholls, R. S. Scott and P. M. George: A randomized, double-blind, placebo-controlled crossover study of coenzyme Q10 therapy in hypertensive patients with the metabolic syndrome. *Am J Hypertens* 25(2), 261-70 (2012)
75. J. Liu, L. Wang, S. Y. Zhan and Y. Xia: Coenzyme Q10 for Parkinson's disease. *Cochrane Database Syst Rev* (12), CD008150 (2011)
76. C. W. Shults, R. H. Haas and M. F. Beal: A possible role of coenzyme Q10 in the etiology and treatment of Parkinson's disease. *Biofactors* 9(2-4), 267-72 (1999)
77. C. W. Shults, R. H. Haas, D. Passov and M. F. Beal: Coenzyme Q10 levels correlate with the activities of complexes I and II/III in mitochondria from parkinsonian and nonparkinsonian subjects. *Ann Neurol* 42(2), 261-4 (1997)
78. M. Sohmiya, M. Tanaka, N. W. Tak, M. Yanagisawa, Y. Tanino, Y. Suzuki, K. Okamoto and Y. Yamamoto: Redox status of plasma coenzyme Q10 indicates elevated systemic oxidative stress in Parkinson's disease. *J Neurol Sci* 223(2), 161-6 (2004)
79. L. K. Mischley, J. Allen and R. Bradley: Coenzyme Q10 deficiency in patients with Parkinson's disease. *J Neurol Sci* 318(1-2), 72-5 (2012)
80. M. W. Horstink and B. G. van Engelen: The effect of coenzyme Q10 therapy in Parkinson disease could be symptomatic. *Arch Neurol* 60(8), 1170-2; author reply 1172-3 (2003)
81. T. Muller, T. Buttner, A. F. Gholipour and W. Kuhn: Coenzyme Q10 supplementation provides mild symptomatic benefit in patients with Parkinson's disease. *Neurosci Lett* 341(3), 201-4 (2003)
82. C. W. Shults, M. F. Beal, D. Fontaine, K. Nakano and R. H. Haas: Absorption, tolerability, and effects on mitochondrial activity of oral coenzyme Q10 in parkinsonian patients. *Neurology* 50(3), 793-5 (1998)
83. C. W. Shults, M. Flint Beal, D. Song and D. Fontaine: Pilot trial of high dosages of coenzyme Q10 in patients with Parkinson's disease. *Exp Neurol* 188(2), 491-4 (2004)
84. O. Cooper, H. Seo, S. Andrabi, C. Guardia-Laguarta, J. Graziotto, M. Sundberg, J. R. McLean, L. Carrillo-Reid, Z. Xie, T. Osborn, G. Hargus, M. Deleidi, T. Lawson, H. Bogetofte, E. Perez-Torres, L. Clark, C. Moskowitz, J. Mazzulli, L. Chen, L. Volpicelli-Daley, N. Romero, H. Jiang, R. J. Uitti, Z. Huang, G. Opala, L. A. Scarffe, V. L. Dawson, C. Klein, J. Feng, O. A. Ross, J. Q. Trojanowski, V. M. Lee, K. Marder, D. J. Surmeier, Z. K. Wszolek, S. Przedborski, D. Krainc, T. M. Dawson and O. Isacson: Pharmacological rescue of mitochondrial deficits in iPSC-derived neural cells from patients with familial Parkinson's disease. *Sci Transl Med* 4(141), 141ra90 (2012)
85. L. Naia, M. J. Ribeiro and A. C. Rego: Mitochondrial and metabolic-based protective strategies in Huntington's disease: the case of creatine and coenzyme Q. *Rev Neurosci* 23(1), 13-28 (2012)
86. E. C. Stack, W. R. Matson and R. J. Ferrante: Evidence of oxidant damage in Huntington's disease: translational strategies using antioxidants. *Ann N Y Acad Sci* 1147, 79-92 (2008)
87. T. L. Wadsworth, J. A. Bishop, A. S. Pappu, R. L. Woltjer and J. F. Quinn: Evaluation of coenzyme Q as an antioxidant strategy for Alzheimer's disease. *J Alzheimers Dis* 14(2), 225-34 (2008)
88. J. Lee, J. H. Boo and H. Ryu: The failure of mitochondria leads to neurodegeneration: Do mitochondria need a jump start? *Adv Drug Deliv Rev* 61(14), 1316-23 (2009)
89. D. R. Galasko, E. Peskind, C. M. Clark, J. F. Quinn, J. M. Ringman, G. A. Jicha, C. Cotman, B. Cottrell, T. J. Montine, R. G. Thomas and P. Aisen: Antioxidants for Alzheimer Disease: A Randomized Clinical Trial With Cerebrospinal Fluid Biomarker Measures. *Arch Neurol* 69(7), 836-41 (2012)
90. J. M. Cooper and A. H. Schapira: Friedreich's ataxia: coenzyme Q10 and vitamin E therapy. *Mitochondrion* 7 Suppl, S127-35 (2007)
91. P. E. Hart, R. Lodi, B. Rajagopalan, J. L. Bradley, J. G. Crilley, C. Turner, A. M. Blamire, D. Manners, P. Styles, A. H. Schapira and J. M. Cooper: Antioxidant treatment of patients with Friedreich ataxia: four-year follow-up. *Arch Neurol* 62(4), 621-6 (2005)
92. D. R. Lynch, S. L. Perlman and T. Meier: A phase 3, double-blind, placebo-controlled trial of idebenone in Friedreich ataxia. *Arch Neurol* 67(8), 941-7 (2010)
93. M. H. Parkinson, J. B. Schulz and P. Giunti: Coenzyme Q10 and idebenone use in Friedreich's ataxia. *J Neurochem* 126 Suppl 1, 125-41 (2013)
94. P. Jolliet, N. Simon, J. Barre, J. Y. Pons, M. Boukef, B. J. Paniel and J. P. Tillement: Plasma coenzyme Q10 concentrations in breast cancer: prognosis and therapeutic consequences. *Int J Clin Pharmacol Ther*, 36(9), 506-9 (1998)
95. K. Lockwood, S. Moesgaard, T. Hanioka and K. Folkers: Apparent partial remission of breast cancer in 'high risk' patients supplemented with nutritional antioxidants, essential fatty acids and coenzyme Q10. *Mol Aspects Med* 15 Suppl, s231-40 (1994)

Clinical applications of coenzyme Q10

96. L. Rusciani, I. Proietti, A. Rusciani, A. Paradisi, G. Sbordoni, C. Alfano, S. Panunzi, A. De Gaetano and S. Lipa: Low plasma coenzyme Q10 levels as an independent prognostic factor for melanoma progression. *J Am Acad Dermatol* 54(2), 234-41 (2006) doi:10.1016/j.jaad.2005.08.031
97. P. R. Palan, M. S. Mikhail, D. W. Shaban and S. L. Romney: Plasma concentrations of coenzyme Q10 and tocopherols in cervical intraepithelial neoplasia and cervical cancer. *Eur J Cancer Prev* 12(4), 321-6 (2003)
98. N. Domae, H. Sawada, E. Matsuyama, T. Konishi and H. Uchino: Cardiomyopathy and other chronic toxic effects induced in rabbits by doxorubicin and possible prevention by coenzyme Q10. *Cancer Treat Rep* 65(1-2), 79-91 (1981)
99. L. Roffe, K. Schmidt and E. Ernst: Efficacy of coenzyme Q10 for improved tolerability of cancer treatments: a systematic review. *J Clin Oncol* 22(21), 4418-24 (2004)
100. G. Brea-Calvo, A. Rodriguez-Hernandez, D. J. Fernandez-Ayala, P. Navas and J. A. Sanchez-Alcazar: Chemotherapy induces an increase in coenzyme Q10 levels in cancer cell lines. *Free Radic Biol Med* 40(8), 1293-302 (2006)
101. J. M. Villalba, C. Parrado, M. Santos-Gonzalez and F. J. Alcain: Therapeutic use of coenzyme Q10 and coenzyme Q10-related compounds and formulations. *Expert Opin Investig Drugs* 19(4), 535-54 (2010)
102. S. Golbidi, S. A. Ebadi and I. Laher: Antioxidants in the treatment of diabetes. *Curr Diabetes Rev* 7(2), 106-25 (2011)
103. Y. Miyake, A. Shouzu, M. Nishikawa, T. Yonemoto, H. Shimizu, S. Omoto, T. Hayakawa and M. Inada: Effect of treatment with 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors on serum coenzyme Q10 in diabetic patients. *Arzneimittelforschung* 49(4), 324-9 (1999)
104. G. F. Watts, D. A. Playford, K. D. Croft, N. C. Ward, T. A. Mori and V. Burke: Coenzyme Q(10) improves endothelial dysfunction of the brachial artery in Type II diabetes mellitus. *Diabetologia* 45(3), 420-6 (2002)
105. J. M. Hodgson, G. F. Watts, D. A. Playford, V. Burke and K. D. Croft: Coenzyme Q10 improves blood pressure and glycaemic control: a controlled trial in subjects with type 2 diabetes. *Eur J Clin Nutr* 56(11), 1137-42 (2002)
106. D. A. Playford, G. F. Watts, K. D. Croft and V. Burke: Combined effect of coenzyme Q10 and fenofibrate on forearm microcirculatory function in type 2 diabetes. *Atherosclerosis* 168(1), 169-79 (2003)
107. J. Hernandez-Ojeda, E. G. Cardona-Munoz, L. M. Roman-Pintos, R. Troyo-Sanroman, P. C. Ortiz-Lazareno, M. A. Cardenas-Meza, S. Pascoe-Gonzalez and A. G. Miranda-Diaz: The effect of ubiquinone in diabetic polyneuropathy: A randomized double-blind placebo-controlled study. *J Diabetes Complications* 26(4), 352-8 (2012)
108. A. Mancini and G. Balercia: Coenzyme Q(10) in male infertility: physiopathology and therapy. *Biofactors* 37(5), 374-80 (2011)
109. A. Mancini, L. De Marinis, A. Oradei, M. E. Hallgass, G. Conte, D. Pozza and G. P. Littarru: Coenzyme Q10 concentrations in normal and pathological human seminal fluid. *J Androl* 15(6), 591-4 (1994)
110. A. Mancini, L. De Marinis, G. P. Littarru and G. Balercia: An update of Coenzyme Q10 implications in male infertility: biochemical and therapeutic aspects. *Biofactors* 25(1-4), 165-74 (2005)
111. M. R. Safarinejad: The effect of coenzyme Q(1)(0) supplementation on partner pregnancy rate in infertile men with idiopathic oligoasthenoteratozoospermia: an open-label prospective study. *Int Urol Nephrol* 44(3), 689-700 (2012)
112. S. Prakash, J. Sunitha and M. Hans: Role of coenzyme Q(10) as an antioxidant and bioenergizer in periodontal diseases. *Indian J Pharmacol* 42(6), 334-7 (2010)
113. H. G. Markley: CoEnzyme Q10 and riboflavin: the mitochondrial connection. *Headache* 52 Suppl 2, 81-7 (2012)
114. T. D. Rozen, M. L. Oshinsky, C. A. Gebeline, K. C. Bradley, W. B. Young, A. L. Shechter and S. D. Silberstein: Open label trial of coenzyme Q10 as a migraine preventive. *Cephalalgia* 22(2), 137-41 (2002)
115. A. D. Hershey, S. W. Powers, A. L. Vockell, S. L. Lecates, P. L. Ellinor, A. Segers, D. Burdine, P. Manning and M. A. Kabbouche: Coenzyme Q10 deficiency and response to supplementation in pediatric and adolescent migraine. *Headache* 47(1), 73-80 (2007)
116. G. Noia, G. P. Littarru, M. De Santis, A. Oradei, C. Mactromarino, C. Trivellini and A. Caruso: Coenzyme Q10 in pregnancy. *Fetal Diagn Ther* 11(4), 264-70 (1996)
117. E. Teran, I. Hernandez, B. Nieto, R. Tavera, J. E. Ocampo and A. Calle: Coenzyme Q10 supplementation during pregnancy reduces the risk of pre-eclampsia. *Int J Gynaecol Obstet* 105(1), 43-5 (2009)
118. L. Tiano, P. Carnevali, L. Padella, L. Santoro, F. Principi, F. Bruge, F. Carle, R. Gesuita, O. Gabrielli and G. P. Littarru: Effect of Coenzyme Q10 in mitigating oxidative DNA damage in Down syndrome patients, a double blind randomized controlled trial. *Neurobiol Aging* 32(11), 2103-5 (2011)
119. L. Tiano and J. Busciglio: Mitochondrial dysfunction and Down's syndrome: is there a role for coenzyme Q(10) ? *Biofactors* 37(5), 386-92 (2011)

Clinical applications of coenzyme Q10

120. G. Lopez-Lluch, J. C. Rodriguez-Aguilera, C. Santos-Ocana and P. Navas: Is coenzyme Q a key factor in aging? *Mech Ageing Dev* 131(4), 225-35 (2010)
121. S. R. Pinnell: Cutaneous photodamage, oxidative stress, and topical antioxidant protection. *J Am Acad Dermatol* 48(1), 1-19 (2003)
122. N. Camougrand and M. Rigoulet: Aging and oxidative stress: studies of some genes involved both in aging and in response to oxidative stress. *Respir Physiol* 128(3), 393-401 (2001)
123. M. Meyer, H. L. Pahl and P. A. Baeuerle: Regulation of the transcription factors NF-kappa B and AP-1 by redox changes. *Chem Biol Interact* 91(2-3), 91-100 (1994)
124. G. J. Fisher and J. J. Voorhees: Molecular mechanisms of photoaging and its prevention by retinoic acid: ultraviolet irradiation induces MAP kinase signal transduction cascades that induce Ap-1-regulated matrix metalloproteinases that degrade human skin *in vivo*. *J Invest Dermatol Symp Proc* 3(1), 61-8 (1998)
125. U. Senftleben and M. Karin: The IKK/NF-kappa B pathway. *Crit Care Med* 30(1 Suppl), S18-26 (2002)
126. G. J. Fisher, S. Kang, J. Varani, Z. Bata-Csorgo, Y. Wan, S. Datta and J. J. Voorhees: Mechanisms of photoaging and chronological skin aging. *Arch Dermatol* 138(11), 1462-70 (2002)
127. Y. Shindo, E. Witt, D. Han, W. Epstein and L. Packer: Enzymic and non-enzymic antioxidants in epidermis and dermis of human skin. *J Invest Dermatol* 102(1), 122-4 (1994)
128. A. Moore: The biochemistry of beauty. The science and pseudo-science of beautiful skin. *EMBO Rep* 3(8), 714-7 (2002)
129. J. Pardeike, K. Schwabe and R. H. Muller: Influence of nanostructured lipid carriers (NLC) on the physical properties of the Cutanova Nanorepair Q10 cream and the *in vivo* skin hydration effect. *Int J Pharm* 396(1-2), 166-73 (2010)
130. S. A. Mortensen, A. Leth, E. Agner and M. Rohde: Dose-related decrease of serum coenzyme Q10 during treatment with HMG-CoA reductase inhibitors. *Mol Aspects Med* 18 Suppl, S137-44 (1997)
131. T. Kishi, T. Watanabe and K. Folkers: Bioenergetics in clinical medicine XV. Inhibition of coenzyme Q10-enzymes by clinically used adrenergic blockers of beta-receptors. *Res Commun Chem Pathol Pharmacol* 17(1), 157-64 (1977)
132. A. M. Moreno-Fernandez, M. D. Cordero, J. Garrido-Maraver, E. Alcocer-Gomez, N. Casas-Barquero, M. I. Carmona-Lopez, J. A. Sanchez-Alcazar and M. de Miguel: Oral treatment with amitriptyline induces coenzyme Q deficiency and oxidative stress in psychiatric patients. *J Psychiatr Res* 46(3), 341-5 (2012)
133. V. L. Serebruany, J. V. Ordenez, W. R. Herzog, M. Rohde, S. A. Mortensen, K. Folkers and P. A. Gurbel: Dietary coenzyme Q10 supplementation alters platelet size and inhibits human vitronectin (CD51/CD61) receptor expression. *J Cardiovasc Pharmacol* 29(1), 16-22 (1997)
134. U. Singh, S. Devaraj and I. Jialal: Coenzyme Q10 supplementation and heart failure. *Nutr Rev* 65(6 Pt 1), 286-93 (2007)
135. R. A. Bonakdar and E. Guarneri: Coenzyme Q10. *Am Fam Physician* 72(6), 1065-70 (2005)
136. T. Hidaka, K. Fujii, I. Funahashi, N. Fukutomi and K. Hosoe: Safety assessment of coenzyme Q10 (CoQ10). *Biofactors* 32(1-4), 199-208 (2008)
137. S. Greenberg and W. H. Frishman: Co-enzyme Q10: a new drug for cardiovascular disease. *J Clin Pharmacol* 30(7), 596-608 (1990)
138. S. Beg, S. Javed and K. Kohli: Bioavailability enhancement of coenzyme Q10: an extensive review of patents. *Recent Pat Drug Deliv Formul* 4(3), 245-55 (2010)
139. K. Adarsh, H. Kaur and V. Mohan: Coenzyme Q10 (CoQ10) in isolated diastolic heart failure in hypertrophic cardiomyopathy (HCM). *Biofactors* 32(1-4), 145-9 (2008)
140. T. Celik and A. Iyisoy: Coenzyme Q10 and coronary artery bypass surgery: what we have learned from clinical trials. *J Cardiothorac Vasc Anesth*, 23(6), 935-6 (2009)
141. G. Caso, P. Kelly, M. A. McNurlan and W. E. Lawson: Effect of coenzyme q10 on myopathic symptoms in patients treated with statins. *Am J Cardiol* 99(10), 1409-12 (2007)
142. R. Belardinelli, A. Mucaj, F. Lacialaprice, M. Solenghi, G. Seddaiu, F. Principi, L. Tiano and G. P. Littarru: Coenzyme Q10 and exercise training in chronic heart failure. *Eur Heart J* 27(22), 2675-81 (2006)
143. C. Henchcliffe and M. F. Beal: Mitochondrial biology and oxidative stress in Parkinson disease pathogenesis. *Nat Clin Pract Neurol* 4(11), 600-9 (2008)
144. J. M. Cooper, L. V. Korlipara, P. E. Hart, J. L. Bradley and A. H. Schapira: Coenzyme Q10 and vitamin E deficiency in Friedreich's ataxia: predictor of efficacy of vitamin E and coenzyme Q10 therapy. *Eur J Neurol* 15(12), 1371-9 (2008)
145. P. S. Sandor, L. Di Clemente, G. Coppola, U. Saenger, A. Fumal, D. Magis, L. Seidel, R. M. Agosti and J. Schoenen: Efficacy of coenzyme Q10 in migraine prophylaxis: a randomized controlled trial. *Neurology* 64(4), 713-5 (2005)
146. P. Mitchell: Protonmotive redox mechanism of the cytochrome b-c1 complex in the respiratory chain:

Clinical applications of coenzyme Q10

protonmotive ubiquinone cycle. *FEBS Lett* 56(1), 1-6 (1975)

147. L. Gille and H. Nohl: The existence of a lysosomal redox chain and the role of ubiquinone. *Arch Biochem Biophys* 375(2), 347-54 (2000)

148. C. Gomez-Diaz, J. C. Rodriguez-Aguilera, M. P. Barroso, J. M. Villalba, F. Navarro, F. L. Crane and P. Navas: Antioxidant ascorbate is stabilized by NADH-coenzyme Q10 reductase in the plasma membrane. *J Bioenerg Biomembr* 29(3), 251-7 (1997)

149. A. Mellors and A. L. Tappel: The inhibition of mitochondrial peroxidation by ubiquinone and ubiquinol. *J Biol Chem* 241(19), 4353-6 (1966)

150. L. Papucci, N. Schiavone, E. Witort, M. Donnini, A. Lapucci, A. Tempestini, L. Formigli, S. Zecchi-Orlandini, G. Orlandini, G. Carella, R. Brancato and S. Capaccioli: Coenzyme q10 prevents apoptosis by inhibiting mitochondrial depolarization independently of its free radical scavenging property. *J Biol Chem* 278(30), 28220-8 (2003)

151. L. Kaas, P. A. Struijs, D. Ring, C. N. van Dijk and D. Eygendaal: Treatment of Mason type II radial head fractures without associated fractures or elbow dislocation: a systematic review. *J Hand Surg Am* 37(7), 1416-21 (2012)

152. C. Schmelzer, I. Lindner, C. Vock, K. Fujii and F. Doring: Functional connections and pathways of coenzyme Q10-inducible genes: an in-silico study. *IUBMB Life* 59(10), 628-633 (2007)

153. S. R. Thomas, J. Neuzil and R. Stocker: Cosupplementation with coenzyme Q prevents the prooxidant effect of alpha-tocopherol and increases the resistance of LDL to transition metal-dependent oxidation initiation. *Arterioscler. Thromb. Vasc. Biol.* 16(5), 687-96 (1996)

154. M. Turunen, L. Wehlin, M. Sjoberg, J. Lundahl, G. Dallner, K. Brismar and P. J. Sindelar: beta2-Integrin and lipid modifications indicate a non-antioxidant mechanism for the anti-atherogenic effect of dietary coenzyme Q10. *Biochem Biophys Res Commun* 296(2), 255-60 (2002)

155. S. J. Hamilton, G. T. Chew and G. F. Watts: Therapeutic regulation of endothelial dysfunction in type 2 diabetes mellitus. *Diab Vasc Dis Res* 4(2), 89-102 (2007)

156. M. E. Jones: Pyrimidine nucleotide biosynthesis in animals: genes, enzymes, and regulation of UMP biosynthesis. *Annu Rev Biochem* 49, 253-79 (1980)

157. M. D. Cordero, E. Alcocer-Gomez, O. Culic, A. M. Carrion, M. de Miguel, E. Diaz-Parrado, E. M. Perez-Villegas, P. Bullon, M. Battino and J. A. Sanchez-Alcazar: NLRP3 Inflammasome is activated in Fibromyalgia: the

effect of Coenzyme Q10. *Antioxid. Redox Signaling* (2013) *in press.*

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